

## **Solid and liquid media for isolating and cultivating acidophilic and acid-tolerant sulfate-reducing bacteria**

Nancucheo, Ivan; Rowe, Owen F.; Hedrich, Sabrina; Johnson, David

### **FEMS Microbiology Letters**

DOI:

[10.1093/femsle/fnw083](https://doi.org/10.1093/femsle/fnw083)

Published: 01/05/2016

Peer reviewed version

[Cyswllt i'r cyhoeddiad / Link to publication](#)

*Dyfyniad o'r fersiwn a gyhoeddwyd / Citation for published version (APA):*

Nancucheo, I., Rowe, O. F., Hedrich, S., & Johnson, D. (2016). Solid and liquid media for isolating and cultivating acidophilic and acid-tolerant sulfate-reducing bacteria. *FEMS Microbiology Letters*, 363, fnw083. <https://doi.org/10.1093/femsle/fnw083>

#### **Hawliau Cyffredinol / General rights**

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

#### **Take down policy**

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

# **Solid and liquid media for isolating and cultivating acidophilic and acid-tolerant sulfate-reducing bacteria**

Ivan Ćancucho<sup>1</sup>, Owen F. Rowe<sup>2</sup>, Sabrina Hedrich<sup>3</sup> and David Barrie Johnson

School of Biological Sciences, College of Natural Sciences, Bangor University,  
Bangor, LL57 2UW, U.K.

Current addresses:

<sup>1</sup>Facultad de Ingeniería y Tecnología, Universidad San Sebastián, Lientur 1457 Concepción, Chile

<sup>2</sup>Department of Food and Environmental Sciences, Division of Microbiology and Biotechnology, Viikki Biocenter 1, University of Helsinki, Helsinki, Finland & the Department of Ecology and Environmental Science, Umeå University, SE-901 87, Umeå, Sweden

<sup>3</sup>Federal Institute for Geosciences and Natural Resources, Resource Geochemistry, Stilleweg 2, 30655 Hannover, Germany

**Correspondence:** Barrie Johnson. College of Natural Sciences, Bangor University, Bangor, LL57 2UW, U.K. Tel: +44 1248 382358; fax: +44 1248 370731; e-mail: d.b.johnson@bangor.ac.uk

**Keywords:** acidophile; glycerol; media; sulfate reducing bacteria; hydrogen sulfide; zinc

## Abstract

Growth media have been developed to facilitate the enrichment and isolation of acidophilic and acid-tolerant sulfate reducing bacteria (aSRB) from environmental and industrial samples, and to allow their cultivation *in vitro*. The main features of the “standard” solid and liquid devised media are: (i) use of glycerol rather than an aliphatic acid as electron donor; (ii) inclusion of stoichiometric concentrations of zinc ions to both buffer pH and to convert potentially harmful hydrogen sulfide produced by the aSRB to insoluble zinc sulfide; (ii) inclusion of *Acidocella aromatica* (an heterotrophic acidophile that does not metabolize glycerol or yeast extract) in the gel underlayer of double layered (overlay) solid media, to remove acetic acid produced by aSRB that incompletely oxidize glycerol and also aliphatic acids (mostly pyruvic) released by acid hydrolysis of the gelling agent used (agarose). Colonies of aSRB are readily distinguished from those of other anaerobes due to their deposition and accumulation of metal sulfide precipitates. Data presented illustrate the effectiveness of the overlay solid media described for isolating aSRB from acidic anaerobic sediments and low pH sulfidogenic bioreactors.

## Introduction

Sulfate reducing bacteria (SRB) comprise a large number of phylogenetically-diverse prokaryotes that have in common their ability to grow by catalyzing the dissimilatory reduction of sulfate to sulfide in anoxic environments (Muyzer & Stams, 2008). They have been detected in, and isolated from, many different marine and freshwater environments, and soils, and also from many “extreme” environments. Thermophilic, psychrophilic, halophilic and alkalophilic species have been described. Acidic environments have presented something of a conundrum, however, as although there have been numerous reports of sulfidogenic activity in anoxic sediments in, for example, sulfate-rich streams draining mine

sites, attempts to isolate acidophilic or acid-tolerant strains of SRB (aSRB) have, until relatively recently, mostly been unsuccessful (Dopson & Johnson, 2012).

One of the hazards faced by SRB is that the sulfide they produce (particularly H<sub>2</sub>S, which is the dominant form present at pH < 7) is toxic to them, as to other life forms, at relatively low (milli-molar) concentrations (Koschorreck, 2008). In many situations (e.g. enrichment culture formulations for neutrophilic SRB) ferrous iron acts as a sink for sulfide, removing it as insoluble, non-bioavailable, iron sulfide (FeS). However, the solubility product of FeS is such that it does not precipitate at pH < 5, and therefore cannot act as an effective sink for sulfide in moderately to extremely acidic situations (Lewis, 2010). A second generic problem for acidophiles is the toxicity of small molecular weight organic acids (Ingledew & Norris, 1992). Many SRB are “incomplete substrate oxidizers”, in that they partially oxidize organic substrates and release the end product(s) of their metabolism (frequently acetate) into their growth milieu. At pH values below their respective pK<sub>a</sub> values (4.75 for acetic acid) small molecular weight organic acids occur predominantly as non-dissociated, lipophilic molecules. Acetic acid is toxic to many chemolithotrophic bacteria when present in micro-molar concentrations, though some heterotrophic acidophiles (e.g. *Acidocella (Ac.) aromatica*) are more tolerant of this and some other aliphatic acids, and use it as a carbon and energy source (Jones *et al.*, 2013). Many media formulations used to enrich neutrophilic SRB utilize organic acids, such as lactate, as carbon and energy sources (e.g. Postgate, 1963). As with acetic acid, lactate exists predominantly as non-dissociated lactic acid at low pH, and again micro-molar concentrations of this potential electron donor are sufficient to partially or completely inhibit the growth of SRB (and most other bacteria) in acidic media. In contrast, organic substrates, such as glycerol, that are uncharged at low pH have been used successfully to enrich for acid-tolerant strains (e.g. Sen & Johnson, 1999).

A variety of solid media formulations have been developed to facilitate the isolation and enumeration of acidophilic chemolithotrophic and heterotrophic microorganisms from environmental and industrial samples (Johnson & Hallberg, 2007). Most of these media employ an “overlay” technique, in which a double-layered gel is used in a standard Petri

55 plate, the lower layer of which is inoculated with an active culture of a heterotrophic  
acidophile (usually an *Acidiphilium* (A.) sp.) while the upper layer is not. The rationale is that  
the heterotrophic acidophile metabolizes the small molecular weight compounds (such as  
pyruvic acid) that derive from acid hydrolysis of commonly-used gelling agents such as agar.  
Using this technique, it has been possible to routinely isolate and cultivate chemolithotrophic  
60 bacteria (such as *Leptospirillum* spp.) that had previously been considered not to grow on  
solid media (Johnson, 1995).

Here we describe solid and liquid media that have been developed in the authors'  
laboratory and used successfully over a number of years to isolate and enumerate aSRB  
from environmental samples, and also to cultivate isolates as axenic cultures in the  
65 laboratory.

## Materials and methods

### Solid media

70 The standard solid medium developed to isolate aSRB from environmental samples and  
laboratory enrichment cultures was a variant of overlay media previously described for  
aerobic acidophiles (Johnson & Hallberg, 2007). The main differences were (i) higher pH of  
the "standard" aSRB medium (~3.7 compared to ~ 2.7 for the aerobic medium plates); (ii)  
75 using the type strain of *Ac. aromatica* rather than *A. cryptum* strain SJH in the underlay gel;  
(iii) inclusion of 4 mM glycerol (as electron donor) and 7 mM zinc (as the sink for hydrogen  
sulfide). Two solutions were prepared and sterilised separately by autoclaving (121°C, 20  
mins) and a third solution (acidic ferrous sulfate) sterilized by filtration through 0.2 µm (pore  
size) membranes:

80 *Solution A* - 20 mL of concentrated basal salts solution, containing (g/L) MgSO<sub>4</sub>·7H<sub>2</sub>O  
(25), (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> (22.5), Na<sub>2</sub>SO<sub>4</sub>·10H<sub>2</sub>O (7.5); KH<sub>2</sub>PO<sub>4</sub> (2.5), KCl (2.5) and Ca(NO<sub>3</sub>)<sub>2</sub>·4H<sub>2</sub>O

(0.7), mixed with 770 mL of reverse osmosis (RO)-grade water, 4 mL of a 1 M glycerol solution, 0.1 g of yeast extract, 7 mL of 1 M zinc sulfate and 0.875 g of magnesium sulfate.

One millilitre of a concentrated trace elements solution was added, the mixture adjusted to

85 pH 3.5 with sulfuric acid, and autoclaved. The trace elements solution contained (g/L):

ZnSO<sub>4</sub>·7H<sub>2</sub>O (10); CuSO<sub>4</sub>·5H<sub>2</sub>O (1.0); MnSO<sub>4</sub>·4H<sub>2</sub>O (1.0); CoSO<sub>4</sub>·7H<sub>2</sub>O (1.0);

Cr<sub>2</sub>(SO<sub>4</sub>)<sub>3</sub>·15H<sub>2</sub>O (0.5); H<sub>3</sub>BO<sub>3</sub> (0.6); Na<sub>2</sub>MoO<sub>4</sub>·2H<sub>2</sub>O (0.5); NiSO<sub>4</sub>·6H<sub>2</sub>O (1); Na<sub>2</sub>SeO<sub>4</sub>·10H<sub>2</sub>O

(1); Na<sub>2</sub>WO<sub>4</sub>·2H<sub>2</sub>O (0.1); NaVO<sub>3</sub> (0.1).

*Solution B* - 5 g of agarose (Sigma Type I) suspended in 200 mL of RO water.

90 Solutions A and B were combined when they had cooled to about 50°C, and 0.1 mL of 1

M ferrous sulfate (*Solution C*) added. The combined solutions were mixed and split into two portions of approximately similar volumes. Ten millilitres of a culture of *Ac. aromatica*<sup>T</sup> (pre-

grown in an acidic (pH 3) medium containing 5 mM fructose, basal salts and trace elements (as above) was added to one portion, and *ca.* 20 mL aliquots of the inoculated molten gel

95 poured into sterile Petri plates. When this layer had solidified, the other (sterile) combined

solution, which had been retained in a molten state by storing in a 50°C water bath, was

poured on top, again in aliquots of *ca.* 20 mL/plate (Fig. 1).

Variants on this “standard” (glycerol/zinc) aSRB solid medium formulation were: (i) a non-overlay version; (ii) one in which zinc sulfate was replaced with 7 mM (final concentration)

100 ferrous sulfate; (iii) more acidic variants (final pH values of ~2.8 or 2.3, achieved by adjusting

the pH of solution A to either 2.5 or 2.0, respectively) in which zinc sulfate was replaced with copper sulfate (CuS has a smaller solubility product than ZnS and precipitates at pH 2.8

whereas ZnS does not). Because of the greater sensitivity of *Ac. aromatica* to copper than to zinc (Jones *et al.*, 2013) the concentration of copper sulfate added was 0.25 mM; (iv) a

105 circum-neutral pH non-overlay variant (adjusting the pH of solution A to 7.0). The pH values

of the gelled media were measured using a calibrated flat-tipped combined pH electrode

(Hanna instruments, UK) coupled to an Accumet 50 pH/E<sub>H</sub> meter.

## Liquid media

110

The standard liquid medium used to grow aSRB isolates *in vitro* contained basal salts/trace elements/magnesium sulfate (as above) supplemented with 4 mM (final concentration) glycerol, 0.01% (w/v) yeast extract and 7 mM zinc sulfate. The pH of the medium was adjusted to either 3.5 or 4.0 with 1 M sulfuric acid, de-aerated under vacuum, and autoclaved  
115 at 121°C for 20 minutes. Ferrous sulfate was added to the cooled medium to give a final concentration of 100 µM (resulting in slight reductions in media pH, to ~ 3.4 and 3.7, respectively). The liquid medium was stored at ambient temperature in an anaerobic glove-box (Plas Labs, Lansing, MI) under a nitrogen/carbon dioxide atmosphere prior to use. A second liquid medium in which zinc sulfate was replaced with 7 mM ferrous sulfate (and pH  
120 adjusted between 3.0 and 5.0) was also used in some experiments.

### Isolation of aSRB on solid media

(i) *Environmental samples.* Sediments in an acidic stream draining an abandoned copper  
125 mine (Cantareras, located in the Iberian Pyrite Belt in south-west Spain; Rowe *et al.*, 2007) were sampled at the entrance, and 10, 30 and 60 m downstream of the mine adit. Sediment samples were placed in sterile Falcon tubes, which were filled to capacity and sealed at the mine site. In the laboratory, 0.5 g of each sediment was mixed with 0.5 ml of pH 2.5 basal salts solution and the suspensions serially diluted and spread onto overlay aSRB solid  
130 media. Inoculated plates were incubated under anaerobic conditions at 30°C using the AnaeroGen system (Oxoid, U.K.), for up to one month. Colonies that grew on SRB plates were examined under a binocular microscope, and sulfidogens tentatively identified by their deposition of metal sulfides (ZnS or CuS) which gave distinctive coloration and metallic sheens to the colonies (Fig. 2). The identities of putative aSRB isolates were confirmed by  
135 amplifying and sequencing their 16S rRNA genes (Rowe *et al.*, 2007). Other overlay plates, formulated to support the growth of chemolithotrophic and heterotrophic acidophiles

(Johnson & Hallberg, 2007) were inoculated at the same time and incubated at 30°C under aerobic conditions.

(ii) *Laboratory bioreactor cultures.* Liquid samples from two continuous-flow sulfidogenic bioreactors, maintained at between pH 2.8 and 4.5 and used to selectively precipitate transition metals from synthetic mine waters (Ñancucheo & Johnson, 2012a) were serially-diluted and spread onto glycerol/zinc overlay aSRB medium. Terminal restriction enzyme fragment length polymorphism (T-RFLP) analysis was carried out in parallel to obtain an overview of the compositions of the bacterial communities in the bioreactors (Ñancucheo & Johnson, 2012a). The bioreactors had originally been inoculated with pure cultures of two aSRB: *Desulfosporosinus acididurans*<sup>T</sup> (Sánchez-Andrea *et al.*, 2015) and “*Desulfobacillus acidavidus*” strain CL4 (Ñancucheo & Johnson, 2012a), and also with dissected samples of a sulfidogenic microbial mat taken from a stream draining the Cantareras mine.

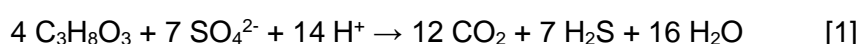
(iii) *Pure cultures of aSRB.* Acidophilic and acid-tolerant SRB that had been isolated previously were also tested for growth on the various solid media. These were *D. acididurans*<sup>T</sup> (originally isolated from a geothermal site in Monserrat, West Indies), and “*Db. acidavidus*” strain CL4 and *Firmicute* strain C5, both of which had been isolated from the microbial mat at Cantareras.

## Results and discussion

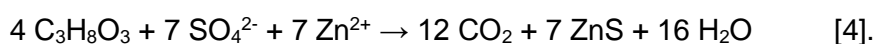
The solid and liquid media described have been used routinely to isolate and cultivate aSRB in the authors' laboratory for several years. *Ac. aromatica* is used in the underlayer gel, as, unlike *Acidiphilium* spp and other *Acidocella* spp., this heterotrophic acidophile uses only a limited range of organic donors (including fructose and acetic acid) but does not grow on yeast extract, glucose, glycerol or many other small molecular weight organic compounds that are commonly metabolized by acidophilic heterotrophic microorganisms. The aSRB present in plate inocula and *Ac. aromatica* do not compete, therefore, for the glycerol and



yeast extract included in the aSRB overlay media. However, acetic acid, which is produced  
 165 as a metabolic waste product by some strains of aSRB, is metabolized by the non-  
 sulfidogenic heterotrophic acidophile, thereby circumventing the problem of this (and other)  
 aliphatic acids inhibiting the growth of the sulfate-reducers. Both the standard solid and liquid  
 media contain zinc sulfate, which has two important functions: (i) removal of toxic hydrogen  
 sulfide as ZnS and (ii) buffering pH. In acidic media, sulfidogenesis is a proton-consuming  
 170 reaction, as (a) the end products of organic substrate (e.g. glycerol) oxidation are either  
 carbon dioxide (complete oxidizers; equation [1]) or a mixture of carbon dioxide and acetic  
 acid (incomplete oxidizers; equation [2]), and (b) sulfate is reduced to H<sub>2</sub>S rather than to HS<sup>-</sup>:



However, the reaction between soluble zinc ions and hydrogen sulfide generates protons  
 (equation [3]), which causes the net reaction in the presence of zinc (shown for the complete  
 oxidation of glycerol in equation [4]) to be pH-neutral:



For this to be effective, soluble zinc has to be present in greater or equimolar concentrations  
 185 to the amount of hydrogen sulfide produced. The “standard” solid and liquid media devised  
 for isolating aSRB contain a molar ratio of glycerol and zinc of 4:7, which provides sufficient  
 zinc to react with the theoretical maximum amount of H<sub>2</sub>S generated (i.e. for aSRB that  
 oxidize glycerol completely to CO<sub>2</sub>) and an excess in the case of incomplete oxidizers. All  
 strains of aSRB isolated in the authors’ laboratory have been found to tolerate zinc  
 190 concentrations well above 7 mM, so that the issue of potential zinc toxicity does not arise.  
 However, lower concentrations of zinc can also be used (if toxicity is suspected) in both solid

and liquid media, so long as the 4:7 ratio (assuming that glycerol is used as electron donor) is maintained. Measurement of the pH of solid and liquid glycerol/zinc media confirmed that pH changes were minor (increasing from ~ 3.7 to ~ 4.0) in grown cultures, in contrast to those containing ferrous sulfate where the pH increased to ~7. The presence of zinc in both solid and liquid media also served as a useful indicator of growth of aSRB. Formation of ZnS in liquid cultures is evidenced by the formation of silver/pink-coloured metallic precipitates (which tend to coat the walls of growth vessels), while ZnS-stained colonies of aSRB are also readily differentiated from non-sulfidogenic anaerobes. Continued accumulation of ZnS causes aSRB colonies to develop hard surface coats with protracted incubation (Fig. 2e).

### **Isolation of aSRB from environmental samples.**

Figure 3 shows data of direct isolation of bacteria from stream sediments taken from the abandoned Cantareras copper mine using a variety of overlay solid media. Isolates were categorized as: (i) iron-oxidizing aerobes (ferric iron-encrusted colonies on aerobically-incubated ferrous iron overlay plates); (ii) heterotrophic aerobes (colonies that grew on aerobically-incubated yeast extract overlay plates); (iii) aSRB (colonies encrusted with ZnS or CuS on anaerobically-incubated overlay plates); (iv) other anaerobes (colonies on anaerobically-incubated overlay plates that were not encrusted with ZnS or CuS). Confirmation of sulfide production by putative SRB isolates was confirmed when colonies were transferred into liquid media, and isolates identified from sequence analysis of their 16S rRNA genes.

Numbers of cultivatable acidophiles were relatively low in the sediment sample from the mine adit entrance ( $<10^3$ /g) but were generally far greater (in some cases by 2-3 orders of magnitude) downstream of the adit. This was particularly noticeable with the anaerobic acidophiles (both the aSRB and the non-sulfidogenic isolates) and corresponded to samples underlying streamer/mat growths that proliferated in the drainage channel at those sampling points (Rowe *et al.*, 2007). The surfaces of these stratified macroscopic growths were

220 dominated by acidophilic algae which were thought to provide much of the organic carbon that sustained the underlying streamer/mat microbial communities, which were predominantly heterotrophic (Nancucheo & Johnson, 2012b). The identities of the non-sulfidogenic anaerobic acidophiles were not determined.

Colony forming units of putative aSRB were about an order of magnitude fewer on  
225 glycerol/copper plates (pH 2.8) than on the higher pH zinc-containing plates (sediments taken at 10 and 30 m downstream of the adit entrance) but were present in slightly greater numbers in the other two sediment samples (Fig. 3). No colonies grew on the more acidic (pH 2.3) glycerol/copper plates nor on the circum-neutral pH solid medium. Colonies of aSRB on copper-containing plates (stained black due to the deposition of CuS) were much smaller  
230 than those on zinc-containing plates and displayed superior growth when transferred to the latter. This was considered due, at least in part, to the greater amount of electron donor (glycerol) in the “standard” solid medium, but could also reflect a preference of the isolates for growing at slightly higher pH.

### 235 Isolation of aSRB from laboratory bioreactor samples

Bacteria (both aSRB and non-sulfidogens) were isolated on solid media at regular intervals from acidic sulfidogenic bioreactors used to selectively remove transition metals from synthetic acidic mine drainage waters and operated at different pH values (Nancucheo &  
240 Johnson, 2012a; Hedrich & Johnson, 2014). Figure 4 shows T-RFLP profiles of the bacterial populations when one of the bioreactors was operated at pH 4.5, 3.0 and 2.8. The profiles were dominated by four restriction fragments of different lengths, each corresponding to a single bacterial species. Two of these were acidophilic sulfidogens (*D. acididurans* and *Peptococcaceae* strain CEB3; Petzsch *et al.*, 2015) while the other two (*Acidithiobacillus*  
245 *ferrooxidans* and *Alicyclobacillaceae* strain IR2) were facultative anaerobes that did not reduce sulfate. The two sulfidogens and strain IR2 were all isolated from bioreactor liquors on glycerol/zinc plates incubated under anaerobic conditions, while *At. ferrooxidans* was

isolated on ferrous iron-containing overlay plates incubated aerobically (Johnson & Hallberg, 2007). Plate counts reflected the relative abundance of bacteria indicated by semi-quantitative T-RFLP analysis.

### **Growth of pure cultures of aSRB on solid media**

Some strains of aSRB isolated and maintained in the authors' laboratory (e.g. strains CL4 and C5) grew on both overlay and non-overlay glycerol/zinc solid media, though colonies tended to grow more slowly and were smaller on the latter. In contrast, *D. acididurans* only grew on the overlay variant of the standard medium, though it also grew on non-overlay plates in which ferrous iron substituted for zinc sulfate. In the latter case, the colonies of *D. acididurans* were heavily blacked-stained (encrusted with FeS) rather than encrusted with silver/pink-coloured ZnS (Figs. 2a & 2b) and the pH of the solid medium increased to ~7 rather than being buffered at ~ pH 4. These differences in behaviour can be attributed to the fact that strains CL4 and C5 appear to oxidize glycerol completely to CO<sub>2</sub>, whereas *D. acididurans* is an incomplete oxidizer. In the pH-buffered glycerol/zinc plates, the acetic acid produced by *D. acididurans* inhibits the growth of the sulfidogen, unless (as in the overlay plate variant) it is removed by *Ac. aromatica* (Kimura *et al.*, 2006). However, in the absence of zinc, the pH of the medium increased (as evidenced by the formation of FeS) causing acetic acid to dissociate to the relatively non-toxic acetate anion, thereby allowing this sulfidogen to grow. Overlay plates are considered to be more versatile and efficient, particularly for isolating acidophilic sulfidogens from environmental samples, given that these may contain both complete and incomplete substrate-oxidizing aSRB.

While the solid and liquid media described herein have proven effective for isolating and cultivating aSRB, other formulations have also been used with varying success. Many sulfidogens (including *D. acididurans*), for example, can use hydrogen as electron donor, and in this case the potential problem of toxicity caused by incomplete oxidation of an organic electron donor obviously does not arise. There are several pragmatic issues, however, which

can limit the use of this inorganic electron donor, including the fact that, unlike with glycerol, it is extremely difficult or impossible to control the molar ratio of this electron donor to the H<sub>2</sub>S sink (zinc ions) which severely limits the potential for pH buffering in batch liquid cultures and solid media. Experiments carried out in which glycerol/zinc plates incubated in anaerobic jars with or without hydrogen gas has not resulted in any significant increases in numbers of aSRB colonies obtained from environmental samples, suggesting that many, if not most hydrogen-oxidizing aSRB, can also grow on glycerol.

## Acknowledgement

The authors wish to acknowledge the contributions of many past and current members of the *Bangor Acidophile Research Team* in general, and Dr. Barry M. Grail in particular, for their contributions to the work described in this report.

## References

- Dopson M & Johnson DB (2012) Biodiversity, metabolism and applications of acidophilic sulfur-metabolizing micro-organisms. *Environ Microbiol* 14: 2620-2631.
- Hedrich S & Johnson DB (2014) Remediation and selective recovery of metals from acidic mine waters using novel modular bioreactors. *Environ Sci Technol* 48: 12206-12212.
- Johnson DB (1995) Selective solid media for isolating and enumerating acidophilic bacteria. *J Microbiol Meth* 23: 205-218.
- Johnson DB & Hallberg KB (2007) Techniques for detecting and identifying acidophilic mineral-oxidizing microorganisms. In: Rawlings DE & Johnson DB. (eds.). *Biomining*. Heidelberg: Springer-Verlag, 237-262.
- Jones RM, Hedrich S & Johnson DB (2013) *Acidocella aromatica* sp. nov.: an acidophilic heterotrophic alphaproteobacterium with unusual phenotypic traits. *Extremophiles* 17: 841-850.
- Kimura S, Hallberg KB & Johnson DB (2006) Sulfidogenesis in low pH (3.8 - 4.2) media by a mixed population of acidophilic bacteria. *Biodegradation* 17: 57-65.
- Koschorreck M (2008) Microbial sulphate reduction at a low pH. *FEMS Microbiol Ecol* 64:

329-342.

Lewis AE (2010) Review of metal sulfide precipitation. *Hydrometallurgy* 104: 222-234.

Muyzer G & Stams AJM (2008) The ecology and biotechnology of sulfate-reducing bacteria.

310 *Nature Rev Microbiol* 6: 441-453.

Ñancucheo I & Johnson DB (2012a) Selective removal of transition metals from acidic mine waters by novel consortia of acidophilic sulfidogenic bacteria. *Microb Biotechnol* 5: 34-44.

Ñancucheo I & Johnson DB (2012b). Acidophilic algae isolated from mine-impacted environments and their roles in sustaining heterotrophic acidophiles. *Front Microbiol* 3:

315 235.

Norris PR & Ingledew WJ (1992) Acidophilic bacteria: adaptations and applications, In: Herbert RA & Sharp AJ (eds.). *Molecular biology and biotechnology of extremophiles*. Glasgow: Blackie, 115-142.

Petzsch P, Poehlein A, Johnson DB, Daniel R, Schlömann M & Mühling M (2015) Genome of the acidophilic sulfate reducing *Peptococcaceae* bacterium strain CEB3. *Genome Announcements* 3: doi:10.1128/genomeA.00886-15.

320

Postgate JR. (1963) Versatile medium for the enumeration of sulfate-reducing bacteria. *Appl Microbiol* 11: 265-267.

Rowe OF, Sánchez-España J, Hallberg KB & Johnson DB (2007) Microbial communities and geochemical dynamics in an extremely acidic, metal-rich stream at an abandoned sulfide mine (Huelva, Spain) underpinned by two functional primary production systems. *Environ Microbiol* 9: 1761-1771.

325

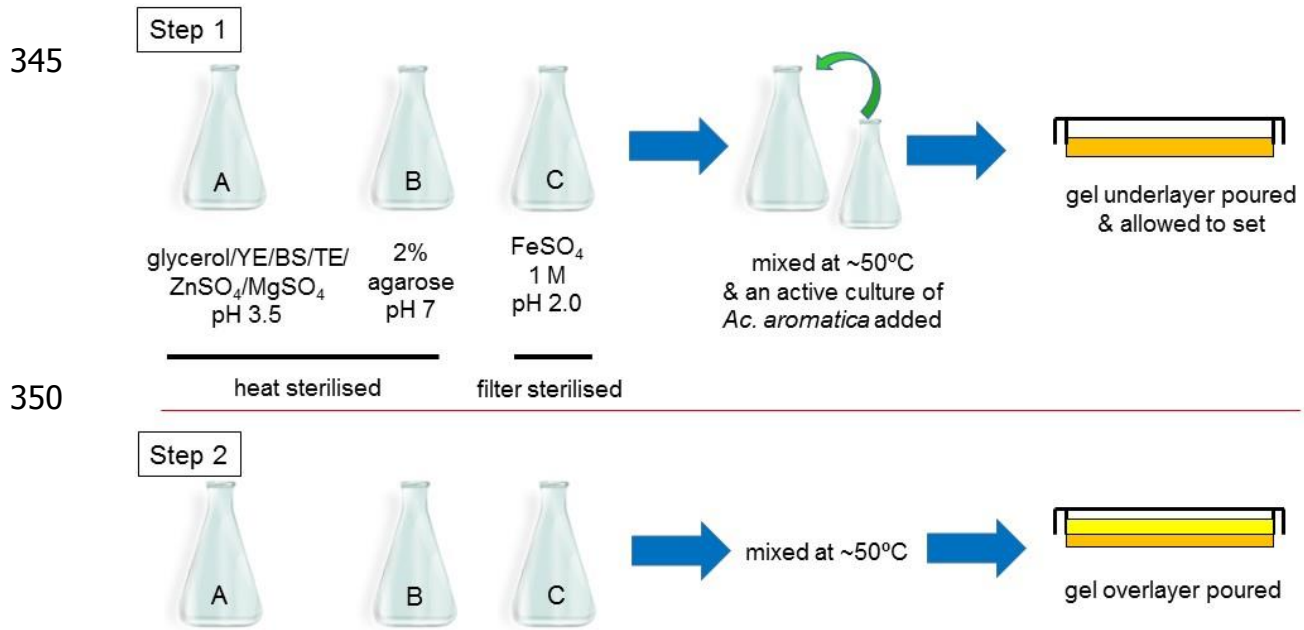
Sánchez-Andrea I, Stams AJM, Hedrich S, Ñancucheo I & Johnson DB (2015) *Desulfosporosinus acididurans* sp. nov., an acidophilic sulfate-reducing bacterium isolated from acidic sediments. *Extremophiles* 19: 39-47.

330

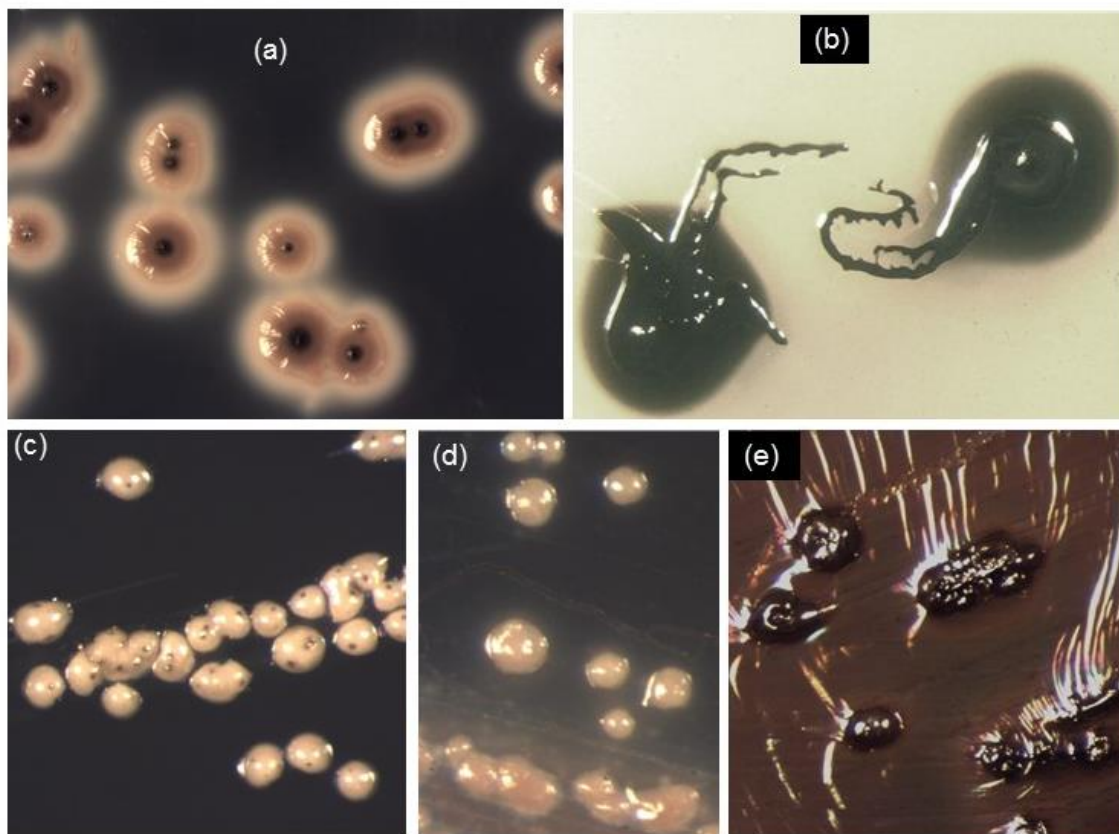
Sen AM & Johnson DB (1999) Acidophilic sulphate-reducing bacteria: candidates for bioremediation of acid mine drainage. In: pp. 709-718, In: Amils R & Ballester A (eds.). *Biohydrometallurgy and the Environment Toward the Mining of the 21st Century*. Process Metallurgy 9A. Amsterdam: Elsevier, 709-718.

335

340

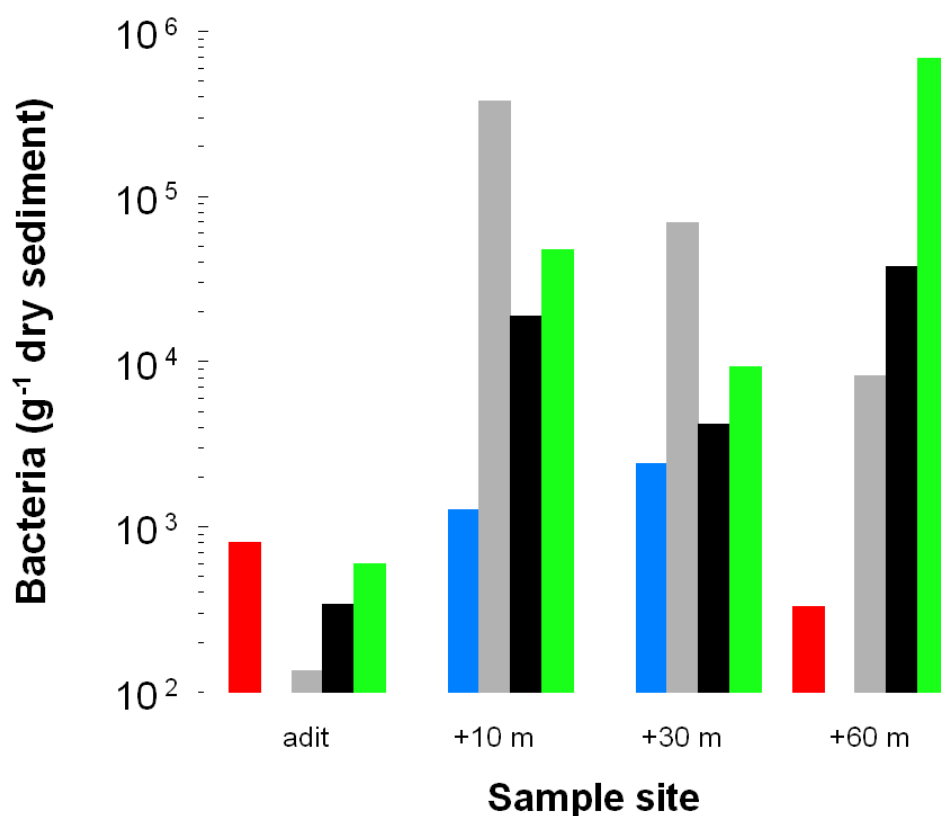


355 **Fig. 1.** Schematic representation of the approach used to prepare overlay medium for aSRB

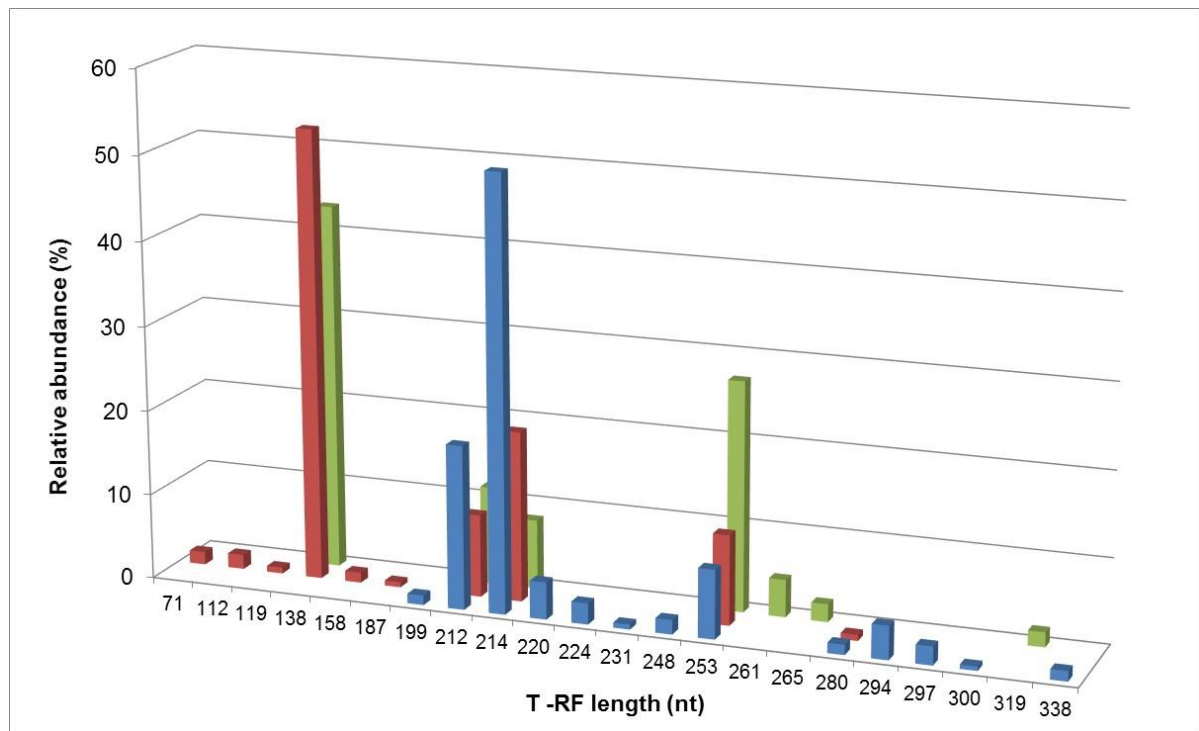


**Fig. 2.** Colonies of aSRB grown on solid media: (a) *D. acididurans*, grown on glycerol/zinc overlay medium; (b) *D. acididurans* grown on glycerol/ferrous iron non-overlay medium; (c) “*Db. acidavidus*” strain CL4, grown on glycerol/zinc overlay medium; (d) *Peptococcaceae* strain CEB3, grown on glycerol/zinc overlay medium; (e) *Firmicute* strain C5 grown on glycerol/zinc overlay medium for 6 weeks, showing colonies covered with shell-like coating of ZnS.





**Fig. 3.** Relative numbers of acidophilic bacteria isolated on solid media from sediment samples taken from the abandoned Cantareras copper mine, at and downstream from the mine adit entrance. Key: red bars, iron-oxidizing aerobes; blue bars, aerobic heterotrophs; grey bars, aSRB (isolated on glycerol/zinc plates); black bars, aSRB (isolated on glycerol copper plates, pH ~2.8); green bars, non-sulfidogenic anaerobes. No colonies were obtained on more acidic (pH ~2.3) glycerol/copper plates, or on circum-neutral pH solid media. In cases where bars are absent, numbers of bacteria isolated were less than 10<sup>2</sup> g<sup>-1</sup>.



**Fig. 4.** Terminal restriction enzyme fragment length polymorphism (T-RFLP) profiles of bacterial 16S rRNA genes (digested with HaeIII) of planktonic bacteria in an acidic sulfidogenic bioreactor, operated at pH 4.5 (blue bars), pH 3.0 (red bars) and pH 2.8 (green bars). The main T-RFs corresponded to; 138 nt, *Peptococcaceae* strain CEB3; 212 nt, *Alicyclobacillaceae* strain IR2; 214 nt, *D. acididurans*; 253 nt, *At. ferrooxidans*.