

Shading impacts by coastal infrastructure on biological communities from subtropical rocky shores'

Pardal-Souza, André; Dias, Gustavo; Jenkins, Stuart; Ciotti, Aurea; Christofoletti, Ronaldo

Journal of Applied Ecology

DOI:

10.1111/1365-2664.12811

Published: 01/06/2017

Peer reviewed version

Cyswllt i'r cyhoeddiad / Link to publication

Dyfyniad o'r fersiwn a gyhoeddwyd / Citation for published version (APA):
Pardal-Souza, A., Dias, G., Jenkins, S., Ciotti, A., & Christofoletti, R. (2017). Shading impacts by coasta in frastructure on biological communities from subtropical rocky shores'. Journal of Applied Ecology, 54(3), 826-835. https://doi.org/10.1111/1365-2664.12811

Hawliau Cyffredinol / General rights
Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
 - You may not further distribute the material or use it for any profit-making activity or commercial gain
 You may freely distribute the URL identifying the publication in the public portal?

Take down policy
If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Shading impacts by coastal infrastructure on biological communities from

2 subtropical rocky shores

3

1

- 4 André Luiz Pardal-Souza, Centre of Natural and Human Sciences, Federal
- 5 University of ABC (CCNH/UFABC), Santo André/SP, Brazil.
- 6 andre.pardal@ufabc.edu.br
- 7 Gustavo Muniz Dias, Centre of Natural and Human Sciences, Federal
- 8 University of ABC (CCNH/UFABC), Santo André/SP, Brazil.
- 9 gmdias@ufabc.edu.br
- 10 Stuart Rees Jenkins, School of Ocean Sciences, Bangor University, Menai
- Bridge, Anglesey, United Kingdom. s.jenkins@bangor.ac.uk
- 12 Áurea Maria Ciotti, Center of Marine Biology, University of São Paulo
- 13 (CEBIMar/USP), São Sebastião/SP, Brazil. ciotti@usp.br
- 14 Corresponding author: Ronaldo Adriano Christofoletti, Marine Institute,
- 15 Federal University of São Paulo (IMar/UNIFESP), Santos/SP, Brazil. Rua Dr
- 16 Carvalho de Mendonça, 144, Santos (SP), Brazil, 11070-102. Phone
- +551338783899. christofoletti@unifesp.br

18

19 **Running title:** Effect of shading on biological communities

20

Summary

22

- 23 **1.** Artificial shading has been highlighted as an important human disturbance,
- 24 affecting both productivity and community organisation. However, studies of
- 25 shading have been poorly explored from an environmental impact perspective.
- 26 **2.** We compared community structure on subtropical rocky shores in areas
- 27 shaded by human constructions with those in unshaded areas. We then
- implemented a manipulative field experiment to determine the effects of shading
- on the macrobenthic community, biofilm biomass and larval recruitment.
- 30 **3.** Shading consistently affected the biological community of rocky shores. The
- 31 biomass and cover of macroalgae and the size of most sedentary grazers were
- 32 smaller in shaded habitat. In the infralittoral fringe, we recorded a shift in
- dominance from macroalgae in unshaded habitats to invertebrate filter-feeders
- in shaded ones. In a similar way, the community from the mesolittoral was also
- affected by shading but not at all locations.
- **4.** Experimental manipulation of shading led to a total loss of macroalgae from
- 37 the infralittoral fringe and no community replacement over a period of 220 days.
- 38 In the mesolittoral, oysters became more abundant in shaded conditions, while
- 39 barnacles decreased in abundance. Larval recruitment was also affected, with
- 40 oysters and barnacles recruiting more in shaded habitats.
- 41 **5.** Synthesis and applications. We demonstrate a clear impact of shading by
- 42 artificial man-made structures on patterns and processes regulating biodiversity
- on rocky shores and thus consequences for coastal ecosystem functioning. We
- argue that shading by artificial coastal structures, such as those proposed in the
- 45 port expansion in our study site in south-eastern Brazil, are potentially under-

estimated. Our work emphasises the importance of careful evaluations of artificial structures in order to promote sustainable coastal development. As a result, we do not recommend the proposed expansion by suspended structures of the port of São Sebastião, as the consequent shading will negatively affect the biodiversity and ecosystem functioning of the Araçá Bay and surrounding areas.

Key-words: Araçá Bay, biofilm, filter-feeder invertebrates, grazers, human disturbances, intertidal, larval recruitment, luminosity, macroalgae, port expansion.

Introduction

Sunlight shading affects the structure and functioning of biological communities in natural ecosystems, through a reduction in the incidence of solar radiation, thus disturbing the growth and biomass production by autotrophs in both terrestrial (Williams, Messier & Kneeshaw 1999; Pagès *et al.* 2003) and aquatic environments (Fitzprack & Kirkman 1995; Quinn *et al.* 1997; Ruiz & Romero 2001). Sunlight limitation can influence animal communities by affecting physical conditions (e.g., reducing heat stress) (Williams 1994; Kon, Kurokura & Tongnunui 2010), decreasing energetic resources of herbivores (Hill, Ryon & Schilling 1995; Harley 2002) and influencing larval recruitment of marine organisms (Thorson 1964; Saunders & Connell 2001; Blockley & Chapman 2006).

In both freshwater and marine environments, sunlight shading occurs naturally due to riparian vegetation (Beschta 1997) or macroalgae coverage (Clark, Edwards & Foster 2004). However, sunlight can also be blocked by man-made structures, such as bridges, piers, wharfs, docks and ports. Although the influence of the additional substrate provided by such structures on aquatic biodiversity has been addressed (Bulleri & Chapman 2010), their role in artificial shading has been overlooked. Results derived from salt marshes (Sanger, Holland & Gainey 2004; Struck et al. 2004), seagrass beds (Burdick & Short 1999; Shafer 1999), estuaries (Able, Manderson & Studholme 1998) and hard substrates (Glasby 1999; Blockley 2007) consistently show negative effects of shading by artificial structures on autotrophs and alterations in the structure of biological communities. Specifically on hard substrates, artificial shading has been associated with shifts in the structure and the diversity of the community, by reducing macroalgae cover (Glasby 1999; Blockley & Chapman 2006; Blockley 2007; Miller & Etter 2008) and also by increasing the overall abundances of some filter-feeding invertebrates and mobile consumers (Williams 1994; Glasby 1999; Takada 1999; Blockley 2007; Miller & Etter 2008).

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

The accelerating urbanisation of coastal areas worldwide is well recognised (Small & Nicholls 2003; NOAA 2004; EEA 2006). Intensification of international commercial trade has increased the demand for construction or expansion of ports and associated infrastructure (Van Gils & Klijn 2007; Hricko 2012). Such projects are usually of great magnitude and cause substantial disturbance to the natural environment where they are installed and in nearby locations. Loss of habitat and biodiversity, contamination by toxic substances, introduction of exotic species and deterioration of air and water quality (Darbra

& Casal 2004; Gupta, Gupta & Patil 2005; Grech *et al.* 2013) are some examples of potential impacts of ports. Quantifying the potential effects of such urban structures on the organisation and functioning of biological communities of natural ecosystems, has gained importance over the past decade, not only from an academic viewpoint, but as tools for coastal management (Bulleri & Airoldi 2005; Perkol-Finkel *et al.* 2012; Ferrario *et al.* 2016). However, in considering the impacts of coastal development the effects of enhanced shading, although recognised, are potentially under-estimated and have received far less attention than other sources of disturbance.

On the north coast of São Paulo State, Brazil, the expansion of the port of São Sebastião has been discussed for many decades (see Appendix S1 in supplementary information for details). The initial proposal was to increase port facilities through infilling of an adjacent area, Araçá Bay (Fig. S1). However, this small bay is a hot spot for benthic biodiversity in the Southwest Atlantic (Amaral et al. 2010, 2015), and after some debate, it was proposed to avoid infilling of the bay by construction of a structure suspended by pillars, covering 75% of Araçá Bay. Among other disturbances, such plans would substantially restrict sunlight to natural habitats. As a consequence we aimed to assess the effects of shading on the rocky intertidal benthic community. Through a descriptive and manipulative approach, we tested the hypotheses that shading (i) decreases abundance or biomass of primary producers; (ii) reduces body size of sedentary grazers, owing to bottom-up control from a reduced biofilm; and (iii) modifies the organisation of sessile macrobenthic communities through effects on autotrophs and on larval recruitment. We show consistent impacts, based on observational surveys and experimental manipulation, of sunlight shading on patterns and

processes driving structure and functioning of biological communities of the rocky intertidal, supporting the importance of the careful considerations of shading disturbance in discussions regarding sustainable development and policies for expansion of ports and man-made structures worldwide.

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

119

120

121

122

Material and methods

DESCRIPTIVE APPROACH

In September 2014, a survey was conducted in three subtropical sheltered rocky shores in the Southwest Atlantic that are partially shaded by man-made constructions, built at least five years before the study. Tidal range for the sites is about 1.4 m (Fig. S1; Table S1). All shores are dominated by filter-feeding invertebrates (e.g., barnacles, oysters and mussels) in the mesolittoral, while macroalgae (e.g., corticoid and turf forming algae) dominate the infralittoral fringe. Biological data were compared between two different habitats within each of the three shores: (i) unshaded, naturally sunny areas; and (ii) shaded, sunlight-restricted areas due to human-made structures. In the shaded habitat, we sampled the area under the structure where no direct sunlight reached the substrate. An area of approximately 50 m horizontal distance was sampled in each habitat; these areas of shore were immediately adjacent to each other (separated by 20 m) to avoid any differences in physical environment such as degree of wave exposure. Response variables measured included the biomass of primary producers, population structure of sedentary grazers and sessile community organization.

Primary producers. Biofilm biomass was estimated by a field-based remote sensing method using digital photographs (adapted from Murphy *et al.* 2005; Murphy & Underwood 2006; Murphy, Underwood & Jackson 2009; see Appendix S2 in supplementary materials for details). Ten images, each of 100 cm², were haphazardly taken in the upper mesolittoral of each location in unshaded and shaded habitats. Macroalgal biomass was estimated by destructive sampling. Five areas of 100 cm² in the infralittoral fringe in both habitats on the three shores were scraped clear and the macroalgae collected were dried at 60° C for 24h and weighed. The dry material was then burned at 500° C for 4h and we calculated the ash free dry weight.

Sedentary grazers. Population structure of three benthic grazers - the limpet *Lottia subrugosa* (d'Orbigny 1846) in the lower mesolittoral and the littorinid gastropods *Echinolittorina lineolata* (d'Orbigny 1840) and *Littoraria flava* (King 1832) in the upper mesolittoral was assessed through photography using quadrats of 100 cm² (n = 20 per habitat) for *L. subrugosa* and 25 cm² (n = 10 per habitat) for littorinids. Due to the small body size, the littorinids were collected and, photographed in the field on a white background to facilitate counting and measurement. Size was defined as the longest length of the shell. Comparisons of body size of consumers between unshaded and shaded areas were performed only when we obtained at least 50 specimens in each habitat on each shore.

Community organization. Sessile macrobenthic communities were compared between unshaded and shaded habitats in three tidal zones: infralittoral fringe,

lower mesolittoral and upper mesolittoral, delimited according to Christofoletti *et al.* (2011). Samples were taken haphazardly through photography (100 cm², n = 10 or 20 per habitat) and taxa abundance estimated as percentage cover using 100 regular intersection grids. Macroalgae were classified into functional groups, according to Littler & Arnold (1982) and Littler, Litter & Taylor (1983). Other taxa were identified to the lowest possible taxonomic group. We also compared the abundance of the functional groups 'macroalgae' and 'filterfeeding invertebrates' between different habitats to test the hypothesis of a dominance shift in the infralittoral fringe due to shading.

MANIPULATIVE APPROACH

Experimental manipulations to test the effect of shading on biofilm biomass and community organization were conducted on the shore of Pernambuco island, within Araçá Bay (São Sebastião, São Paulo State, Brazil; Fig. S1). The experiment lasted 221 days in two intertidal zones (infralittoral fringe and upper mesolittoral) and was performed with three treatments: (i) shaded, constructed with marine plywood sheets (40 x 40 cm) suspended 10 cm above the substrate by stainless steel bars in each corner; (ii) procedural control, provided by transparent acrylic sheets, mimicking the physical structure of the shaded treatment but allowing sunlight penetration to the substratum (≈ 90%); and (iii) control, unmanipulated areas. While all the 40 x 40 cm area is under the influence of the treatment we only used the central 100 cm² for analyses, to avoid artefacts caused by diffuse light at the perimeter. Although partial shading (diffuse light) is a natural consequence of many artificial structures our manipulative approach intended to simulate the effect of full

shade expected following potential port expansion We deployed 5 replicates of each treatment in each intertidal zone. Replicates were separated by at least 2 m and the treatments were randomly allocated within each tidal zone. Acrylic plates were regularly cleaned and damaged structures were replaced as soon as possible. To test the efficacy of manipulations, luminosity and temperature sensors were deployed on treatments at both tide heights. Both luminosity and temperature were reduced by shading, but did not differ between control treatments (more details in Appendix S3).

Biofilm biomass was evaluated in the upper mesolittoral using the same remote sensing technique and protocols (Appendix S2). Samples were undertaken 0, 15, 29, 44, 73, 149, 191 and 220 days after the start of the experiment. Benthic community composition was sampled on the same dates in the upper mesolittoral and, after 0, 15, 29, 75, 191 and 221 days in the infralittoral fringe, as allowed by tidal conditions. Because communities were dominated by a few species during the whole experiment, we tested how the treatments affected the areas covered by macroalgae, oysters, the barnacle *Chthamalus bisinuatus* (Pilsbry 1916) and open space.

Larval recruitment. To test whether alterations in adult populations were linked to effects of shading on larval recruitment, we utilized acrylic plates (8 x 8 cm) covered with gray slip-resistant tape (3MTM Safety-Walk, Minnesota, EUA). Plates were screwed in the upper mesolittoral, under the procedural control and shaded treatments, avoiding central sampling area, and close to replicates in control. Plates were replaced approximately every 30 days. In the laboratory, we identified and quantified recruits under a stereomicroscope. We tested the

effects of time and treatment on the recruitment rate (number of recruits per days in the field) of barnacles and oysters, the two most abundant taxa in the upper mesolittoral.

DATA ANALYSIS

In the descriptive approach, ecological parameters from primary producers, sedentary grazers and univariate data from benthic community (combined macroalgae and filter-feeding invertebrates) were analyzed using factorial ANOVA, considering the factors 'habitat' (fixed, 2 levels: unshaded and shaded) and 'location' (random, 3 levels). Specifically for the analysis of body size of *Lottia subrugosa* and *Echinolittorina lineolata*, location had 2 levels as we did not sample enough specimens at one of the shores. Variance homogeneity was tested by Cochran's procedure and transformations were applied when needed. Where heterogeneous variances persisted, the same procedure was still performed using raw data, and attention is drawn to the potential for increased probability of type I errors in these cases. Body size of *Littoraria flava* was compared through a t-test after confirming homoscedasticity (Levene's test), since we sampled this species in enough number only in one shore.

Data from the sessile macrobenthic communities were converted to a similarity matrix based on Bray-Curtis distance and compared between 'habitat' (fixed, 2 levels: unshaded and shaded) and 'location' (random, 3 levels) using PERMANOVA test (999 permutations) (Anderson 2001). Comparisons of communities from different habitats within locations were performed by post-hoc pair-wise tests. Non-metric multidimensional scaling (nMDS) was used for

visualization of data. SIMPER analysis was applied to identity the taxa which contributed most to dissimilarity between habitats.

In the experimental approach, repeated measures ANOVA was used to assess the differences between treatments through time on biofilm biomass, area covered by macroalgae, oysters, barnacle *Chthamalus bisinuatus* and open space. Mauchly's sphericity test was applied to verify time autocorrelation. When this assumption was violated, we corrected statistical significances with Greenhouse-Geisser adjustment. Post-hoc Student-Newman-Keuls (SNK) test was used for multiple comparisons of means for both statistical techniques. The effects of 'treatment' and 'time' (random factor) on recruitment rate during the manipulative approach were evaluated using a factorial ANOVA.

Results

DESCRIPTIVE APPROACH

Primary producers

There was no influence of shading on biofilm biomass on shores under the influence of man-made constructions (Table 1). On the other hand, shading decreased the macroalgal biomass, which was about eight times lower in the shaded (mean \pm SE: 1.01 \pm 0.43 g) than in the unshaded habitat (8.33 \pm 1.28 g) (Table 1).

Sedentary grazers

Shading affected body size of the limpet *Lottia subrugosa* and the littorinids *Echinolittorina lineolata* (Table 1) and *Littoraria flava* (t-test, df = 121, t = 5.36, P < 0.001) (Fig. 1). Grazers from four of the five sampled populations were smaller in the shaded habitat, although, *L. subrugosa* in one of the localities was bigger in shaded habitat (SNK 'Habitat x Location' interaction: Table 1).

There was a general trend across all three species at all shores for greater abundance in unshaded areas (Fig 1). However only for *Echinolittorina lineolata*, on one rocky shore, was this effect of shade significant (SNK 'Habitat x Location' interaction: Table 1; Fig. 1).

Community organization

There was a significant, although spatially variable, effect of shading on community organization. In the infralittoral fringe, there was a significant effect of shading at all three shores, while the effect was significant on two shores in the lower mesolittoral and one shore in the upper mesolittoral (Post-hoc pairwise test 'Habitat x Location' interaction: Table 2; Fig. 2).

In the infralittoral fringe, cover of combined macroalgae decreased from about 70% in unshaded habitat to 17% in shaded habitat. On the other hand, combined invertebrate filter-feeders increased from 5% in unshaded to 65% in shaded habitats (ANOVA, effect of 'Habitat', macroalgae: $F_{1,54} = 23,470.89$; filter-feeding invertebrates: $F_{1,54} = 491.84$; P < 0.01). Macroalgae morphofunctional groups combined were responsible for more than 45% of dissimilarity between habitats (SIMPER: Table S2). For both other zones, while

the same species occurred in shaded and unshaded habitats, the relative abundance of organisms was affected by shading (SIMPER: Table S2). In the lower mesolittoral, oysters were more abundant in shaded than in unshaded areas, while *Brachidontes* sp. showed an opposite pattern. These two species contributed 55% of the dissimilarity between the sessile communities from shaded and unshaded habitats at this shore height. In the upper mesolittoral, the barnacle *Microeuraphia rizophorae* (De Oliveira 1940) was more abundant in shaded than in unshaded habitats, while *Chthamalus bisinuatus* showed an opposite pattern. These two species contributed 91% of the dissimilarity between the sessile communities from shaded and unshaded habitats in the upper mesolittoral.

MANIPULATIVE APPROACH

Biofilm biomass

Biofilm biomass showed a high degree of variation among replicates and over time (Fig. 3). Although there was a significant effect of treatment (Table 3), this was a consequence of the control being placed in plots of higher NDVI at the start of the experiment (Fig. 3). There was no significant interaction between treatment and time which might indicate divergence of the treatments over time (Table 3).

Community organization

In the infralittoral fringe, macroalgae covered almost 100% at the beginning of the experiment and quickly decreased in the shading treatment

during the first 2 months, completely disappearing at day 221 (SNK 'Treatment x Time' interaction: Table 3, Fig. 4). Shading also affected the structure of the macrobenthic community in the upper mesolittoral. In the shaded treatment, oysters increased in abundance from 1.2% at the beginning of the experiment to 37.8% after 220 days, while there was no change in other treatments. Conversely, the barnacle *Chthamalus bisinuatus* decreased in abundance due to shading, reaching a cover of about 10% by the end of the experiment (SNK 'Treatment x Time' interaction: Table 3; Fig. 4).

Larval recruitment

Larval recruitment rate was affected by shade. Oyster larvae recruited more in the shaded treatment than in the control treatments (SNK 'Treatment': Table 4; Fig. 5). For the barnacle *Chthamalus bisinuatus*, larval recruitment was smaller in the control than in shaded and procedural control (SNK 'Treatment': Table 4; Fig. 5). Also, there was variation among sampling dates, with larger numbers of *C. bisinuatus* larvae recruiting in April/2015 than in all 5 months before, which did not differ from one another (SNK 'Time': Table 4).

Discussion

Many studies have shown how artificial structures can influence local ecosystems by the addition of substrate (see Bulleri & Chapman 2010 for review). We showed important ecological changes on natural rocky substrates associated with such structures and demonstrated through experiments that the changes were a consequence of shading. Under shading disturbance, biomass

of primary producers, body size of primary consumers, community structure and larval recruitment changed, and the results were consistent for both descriptive and manipulative approaches. The changes in communities are likely explained by physical factors and biological processes such as competition, recruitment rates and physiological status of the organisms, as discussed below. Our observational and experimental results contribute quantitatively to the extensive debate on coastal management under pressure from urbanization (Bulleri & Chapman 2010; Dugan *et al.* 2011).

The results highlighted more pronounced changes in the intertidal communities found at the lower levels in the shore, suggesting that this zone is more vulnerable to shading. There was a strong negative influence of shading on macroalgae, expected since light restriction limits photosynthesis and prevents autotrophs survival, as observed for other ecosystems (e.g., Glasby 1999; Shafer 1999; Struck *et al.* 2004; Blockley 2007; Miller & Etter 2008). Surveys showed that in areas shaded by human-made structures, macroalgae coverage and biomass were low, while experimental manipulation of shade caused progressive loss of existing macroalgae, with total loss in about 6 months. Differences in abundance of macroalgae between unshaded and shaded habitats can also be linked to recruitment, as macroalgae tend to recruit less in light-limited areas (Clark, Edwards & Foster 2004; Blockey & Chapman 2006) due to high post-settlement mortality of spores and low growth (Goldberg & Foster 2002).

With the reduction in macroalgae in the infralittoral zone affected by manmade structures, dominance shifted toward filter-feeding invertebrates. This shift to invertebrates was not observed following shade manipulation, possibly as a consequence of limited recruitment over the 220 day experimental period, but also potentially owing to high mortality of early settlers caused by sedimentation observed in experimental plots (Airoldi 2003). It was not clear the extent to which such sedimentation was an artefact of the shade structures, since sedimentation on rock throughout Araçá Bay is common depending on prevailing weather and sea conditions. Whether shading leads to dominance by filter feeding invertebrates, or to open bare space, there was a clear reduction in local biodiversity, considering the loss of macroalgae species and their associated fauna (Christie, Norderhaug & Fredriksen 2009). An additional consequence of loss of turf macroalgae cover is the opening up of bare space and an enhanced probability of invasive species establishment (Arenas *et al.* 2006). This shading may be an additional mechanism by which urban infrastructure in port facilities can facilitate the introduction and spread of exotic species (Bulleri & Airoldi 2005; Vaselli, Bulleri & Benedetti-Cecchi 2008; Dafforn, Johnston & Glasby 2009).

We predicted, based on previous observations (Hill, Ryon & Schilling 1995; Harley 2002), that shading would have a negative effect on the intertidal epilithic biofilm, but we found no support for this hypothesis. The intertidal biofilm is a complex system and the influence of light on its dynamics is still controversial. Biofilm biomass can increase due to shading or reduced sunlight regimes (Jenkins *et al.* 2001; Thompson, Norton & Hawkins 2004) or show restricted growth when exposed to excess sunlight, due to photoinhibition or thermal stress (Serôdio, Vieira & Cruz 2008; Coelho, Vieira & Serôdio 2009). Also, it is important to note that our manipulation did not exclude biofilm grazers. Previous work in the study region has shown that fast-moving grazers

mask the effect of sedentary grazers on biofilm, while grazing pressures of both together mask environmental influences (Christofoletti, Almeida & Ciotti 2011). Although the influence of shading on biofilm was not seen, our prediction of negative shade effects on grazers was confirmed at most locations. Another, non-exclusive hypothesis to explain bigger animals in sunny habitats is that grazer size is related to increased heat stress in sunny habitats, which would select specimens with larger shells due to optimized water storage (Vermeij 1973; Tanaka, Duque-Estrada & Magalhães 2002) and also increasing growth rate due a higher metabolism (Chelazzi, Williams & Gray 1999).

Shading can also promote differences between communities by changing the recruitment regime (Chapman & Blockley 2006). Many late-stage larvae show active choice at settlement (Keough & Downes 1982); available light is an important cue with many larvae of marine organisms exhibiting negative phototactic behaviour, thus settling in light-limited habitats (Thorson 1964; Young & Chia 1984; Saunders & Connell 2001). Blockley & Chapman (2006) showed that shading increased recruitment of some filter-feeding invertebrates but reduced macroalgae recruitment on seawalls. Such results are consistent with the abundance of these taxa in established shaded communities in the present study, supporting the conclusion that changes in community organisation in the infralittoral fringe and mesolittoral following shading may be linked to larval recruitment. Oysters became more abundant in the upper mesolittoral in the shaded treatment and barnacle recruitment increased in shaded habitats. However, interestingly, the abundance of adult barnacles reduced under shading. Such patterns, both in the natural environment and experiments indicate the need to differentiate between effects of shade on settlement/ early post settlement mortality and the development of adult populations.

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

428

429

430

431

432

433

434

435

436

Our study shows the consequences of shading from artificial structures on the biota of intertidal rocky shores. Coastlines worldwide are being increasingly modified through constructions ranging from artificial sea defences to port facilities (Bulleri & Chapman 2010; Dugan et al. 2011). Such urban infrastructures alter landscape via loss, addition or fragmentation of habitat (Chapman 2006; Goodsell, Chapman & Underwood 2007), and modify biodiversity, since artificial and natural habitats do not support the same communities (Bulleri & Chapman 2004). Our study was prompted by the proposal for expansion of the port of São Sebastião which would result in a shaded area of approximately 1 km² of the Araçá Bay, impacting rocky shores, soft sediment habitats and mangroves. Results indicate such a development would cause substantial decreases in macroalgae cover on hard substrate leading to a reduction in primary production, carbon exchange and habitat for associated fauna. Shading is also predicted to increase filter-feeding invertebrate cover strongly influencing the dynamics of the pelagic environment. The Araçá Bay, like many other coastal sites threatened by development is a hot spot in marine biodiversity (Amaral et al. 2010, 2015) supporting a wide range of ecosystem services. Understanding the impact of shading contributes to a wider view of the consequences of such development and hence informs discussions regarding sustainable development, in this and other regions around the world. Specifically regarding the expansion plans of the port of São Sebastião, our results suggest that covering the bay with a suspended structure will have major negative effects on biodiversity and ecosystem functioning. As a

result, we recommend that stakeholders carefully evaluate the expansion plan, and, based on the likely shade effects (as well as many other potential impacts) across much of the bay, we do not recommend it.

440

441

437

438

439

Acknowledgments

442 This work was supported by research funds granted by São Paulo 443 Research Foundation (FAPESP) to A.L.P.S (#2013/19122-0), to A.M.C 444 (#2013/50199-8), to R.A.C. (#2013/11594-9) and to Biota Araçá Project 445 (#2011/50317-5). We thank Deborah, Jaqueline, Gabriella, Neemias, Ivan, Elso 446 and Joseilto for support with field work. Finally, we are grateful to Isabella and 447 A.C.Z Amaral for help with some figures, and to M.O. Tanaka, G.H. Pereira-448 Filho and the anonymous referees for their valuable comments on the 449 manuscript.

450

451

Data accessibility

- Study sites locations: uploaded as online supporting information (Table S1).
- Data archived in Figshare https://dx.doi.org/10.6084/m9.figshare.3205285.v1
- 454 (Pardal-Souza *et al.* 2016).

455

456

References

- 457 Able, K.W., Manderson, J.P. & Studholme, A.L. (1998) The distribution of 458 shallow water juvenile fishes in an urban estuary: the effects of manmade
- structures in the lower Hudson river. *Estuaries*, **21**, 731–744.

- 460 Airoldi, L. (2003) The effects of sedimentation on rocky coast assemblages.
- Oceanography and Marine Biology: an Annual Review, **41**, 161–236.
- 462 Amaral, A.C.Z., Migotto, A.E., Turra, A. & Schaeffer-Novelli, Y. (2010) Araçá:
- biodiversidade, impactos e ameaças. *Biota Neotropica*, **10**, 219–264.
- 464 Amaral, A.C.Z., Turra, A., Ciotti, A.M., Rossi-Wongtschowski, C.L.B. &
- 465 Schaeffer-Novelli, Y. (2015) Vida na Baía do Araçá: diversidade e importância,
- 466 1st edn. Lume, São Paulo.
- 467 Anderson, M.J. (2001) A new method for non-parametric multivariate analysis of
- variance. Austral Ecology, **26**, 32–46.
- 469 Arenas, F., Bishop, J.D.D., Carlton, J.T., Dyrynda, P.J., Farnham, W.F.,
- 470 Gonzalez, D.J., Jacobs, M.W., Lambert, C., Lambert, G., Nielsen, S.E.,
- 471 Pederson, J.A., Porter, J.S., Ward, S. & Wood, C.A. (2006) Alien species and
- other notable records from a rapid assessment survey of marinas on the south
- 473 coast of England. Journal of the Marine Biological Association of the United
- 474 *Kingdom*, **86**, 1329–1337.
- 475 Beschta, R.L. (1997) Riparian shade and stream temperature: an alternative
- 476 perspective. *Rangelands*, **19**, 25–28.
- 477 Blockley, D.J. & Chapman, M.G. (2006) Recruitment determines differences
- 478 between assemblages on shaded or unshaded seawalls. Marine Ecology
- 479 *Progress Series*, **327**, 27–36.
- 480 Blockley, D.J. (2007) Effect of wharves on intertidal assemblages on seawalls in
- 481 Sydney Harbour, Australia. *Marine Environmental Research*, **63**, 409–427.

- 482 Bulleri, F. & Airoldi, L. (2005) Artificial marine structures facilitate the spread of
- 483 a non-indigenous green alga, Codium fragile ssp. tomentosoides, in the north
- 484 Adriatic Sea. *Journal of Applied Ecology*, **42**, 1063–1072.
- 485 Bulleri, F. & Chapman, M.G. (2004) Intertidal assemblages on artificial and
- natural habitats in marinas on the north-west coast of Italy. *Marine Biology*, **145**,
- 487 **381–391**.
- 488 Bulleri, F. & Chapman, M.G. (2010) The introduction of coastal infrastructure as
- a driver of change in marine environments. Journal of Applied Ecology, 47, 26-
- 490 35.
- 491 Burdick, D.M. & Short, F.T. (1999) The effects of boat docks on eelgrass beds
- in coastal waters of Massachusetts. *Environmental Management*, **23**, 231–240.
- 493 Chapman, M.G. (2006) Intertidal seawalls as habitats for molluscs. Journal of
- 494 *Molluscan Studies*, **72**, 247–257.
- 495 Chelazzi, G., Williams, G.A. & Gray, D.G. (1999) Field and laboratory
- 496 measurement of heart rate in a tropical limpet, Cellana grata. Journal of the
- 497 Marine Biological Association of the United Kingdom, **79**, 749–751.
- 498 Christie, H., Norderhaug, K.M. & Fredriksen, S. (2009) Macrophytes as habitat
- for fauna. *Marine Ecology Progress Series*, **396**, 221–233.
- 500 Christofoletti, R.A., Almeida, T.V.V. & Ciotti, A.M. (2011) Environmental and
- grazing influence on spatial variability of intertidal biofilm on subtropical rocky
- shores. *Marine Ecology Progress Series*, **424**, 15–23.
- 503 Christofoletti, R.A., Takahashi, C.K., Oliveira, D.N. & Flores, A.A.V. (2011)
- 504 Spatial and temporal variation of sedentary consumers and sessile organisms

- on shores of a subtropical area of the south-west Atlantic. *Journal of the Marine*
- 506 Biological Association of the United Kingdom, **91**, 961–967.
- 507 Clark, R.P., Edwards, M.S. & Foster, M.S. (2004) Effects of shade from multiple
- kelp canopies on an understory algal assemblage. Marine Ecology Progress
- 509 Series, **267**, 107–119.
- 510 Coelho, H., Vieira, S. & Serôdio, J. (2009) Effects of desiccation on the
- 511 photosynthetic activity of intertidal microphytobenthos biofilms as studied by
- optical methods. Journal of Experimental Marine Biology and Ecology, 381, 98-
- 513 104.
- 514 Dafforn, K.A., Johnston, E.L. & Glasby, T.M. (2009) Shallow moving structures
- promote marine invader dominance. *Biofouling*, **25**, 277–287.
- 516 Darbra, R.M. & Casal, J. (2004) Historical analysis of accidents in seaports.
- 517 Safety Science, **42**, 85–98.
- 518 Dugan, J.E., Airoldi, L., Chapman, M.G., Walker, S.J. & Schlacher, T. (2011)
- 519 Estuarine and coastal structures: environmental effects, a focus on shore and
- 520 nearshore structures. Treatise on Estuarine and Coastal Science (eds E.
- Wolanski & D.S. McLusky), pp. 17-41. Academic Press, Waltham.
- 522 EEA (2006) The changing faces of Europe's coastal areas. EEA Report 6/2006.
- 523 OPOEC, Luxembourg.
- 524 Ferrario, F., Iveša, L., Jaklin, A., Perkol-Finkel, S. & Airoldi, L. (2016) The
- overlooked role of biotic factors in controlling the ecological performance of
- artificial marine habitats. *Journal of Applied Ecology*, **53**, 16–24.

- 527 Fitzpatrick, J. & Kirkman, H. (1995) Effects of prolonged shading stress on
- 528 growth and survival of seagrass *Posidonia australis* in Jervis Bay, New South
- Wales, Australia. *Marine Ecology Progress Series*, **127**, 279–289.
- 530 Glasby, T.M. (1999) Effects of shading on subtidal epibiotic assemblages.
- Journal of Experimental Marine Biology and Ecology, **234**, 275–290.
- Goldberg, N.A. & Foster, M.S. (2002) Settlement and postsettlement processes
- 533 limit the abundance of the geniculate coralline alga *Calliarthron* on subtidal
- walls. Journal of Experimental Marine Biology and Ecology, **278**, 31–45.
- Goodsell, P.J., Chapman, M.G. & Underwood, A.J. (2007) Differences between
- biota in anthropogenically fragmented habitats and in naturally patchy habitats.
- 537 Marine Ecology Progress Series, **351**, 15–23.
- 538 Grech, A., Bos, M., Brodie, J., Coles, R., Dale, A., Gilbert, R., Hamann, M.,
- Marsh, H., Neil, K., Pressey, R.L., Rasheed, M.A., Sheaves, M. & Smith, A.
- 540 (2013) Guiding principles for the improved governance of port and shipping
- impacts in the Great Barrier Reef. *Marine Pollution Bulletin*, **75**, 8–20.
- 542 Gupta, A.K., Gupta, S.K. & Patil, R.S. (2005) Environmental management plan
- for port and harbour projects. Clean Technologies and Environmental Policy, 7,
- 544 133–141.
- 545 Harley, C.D. (2002) Light availability indirectly limits herbivore growth and
- abundance in a high rocky intertidal community during the winter. *Limnology*
- 547 and Oceanography, **47**, 1217–1222.
- 548 Hill, W.R., Ryon, M.G. & Schilling, E.M. (1995) Light limitation in a stream
- 549 ecosystem: responses by primary producers and consumers. Ecology, 76,
- 550 1297–1309.

- 551 Hricko, A. (2012) Progress & Pollution: Port cities prepare for the Panama
- 552 Canal expansion. *Environmental Health Perspectives*, **120**, 470–473.
- Jenkins, S.R., Arenas, F., Arrontes, J., Bussel, J., Castro, J., Coleman, R.A.,
- Hawkins, S.J., Kay, S., Martínez, B., Oliveros, J., Roberts, M.F., Sousa, S.,
- 555 Thompson, R.C. & Hartnoll, R.G. (2001) European-scale analysis of seasonal
- variability in limpet grazing activity and microalgal abundance. *Marine Ecology*
- 557 Progress Series, **211**, 193–203.
- Keough, M.J. & Downes, B.J. (1982) Recruitment of marine invertebrates: the
- role of active larval choices and early mortality. *Oecologia*, **54**, 348–352.
- Kon, K., Kurokura, H. & Tongnunui, P. (2010) Effects of the physical structure of
- 561 mangrove vegetation on a benthic faunal community. Journal of Experimental
- 562 *Marine Biology and Ecology*, **383**, 171–180.
- 563 Littler, M.M. & Arnold, K.E. (1982) Primary productivity of marine macroalgal
- functional-form groups from Southwestern North America. *Journal of Phycology*,
- 565 **18**, 307–311.
- Littler, M.M., Littler, D.S. & Taylor, P.R. (1983) Evolutionary strategies in a
- 567 tropical barrier reef system: functional-form groups of marine macroalgae.
- 568 *Journal of Phycology*, **19**, 229–237.
- 569 Miller, R.J. & Etter, R.J. (2008) Shading facilitates sessile invertebrate
- dominance in the rocky subtidal Gulf of Maine. *Ecology*, **89**, 452–462.
- 571 Murphy, R.J. & Underwood, A.J. (2006) Novel use of digital colour-infrared
- imagery to test hypotheses about grazing by intertidal herbivorous gastropods.
- Journal of Experimental Marine Biology and Ecology, **330**, 437–447.

- 574 Murphy, R.J., Underwood, A.J. & Jackson, A.C. (2009) Field-based remote
- 575 sensing of intertidal epilithic chlorophyll: techniques using specialized and
- 576 conventional digital cameras. Journal of Experimental Marine Biology and
- 577 *Ecology*, **380**, 68–76.
- 578 Murphy, R.J., Underwood, A.J., Pinkerton, M.H. & Range, P. (2005) Field
- spectrometry: new methods to investigate epilithic microalgae on rocky shores.
- Journal of Experimental Marine Biology and Ecology, **325**, 111–124.
- NOAA (2004) Population trends along the coastal United States: 1980-2008.
- 582 Coastal Trends Report Series. NOAA's National Ocean Service Management
- 583 and Budget Office Special Projects.
- Pagès, J.P., Pache, G., Joud, D., Magnan, N. & Michalet, R. (2003) Direct and
- 585 indirect effects of shade on four forest tree seedlings in the French Alps.
- 586 *Ecology*, **84**, 2741–2750.
- Pardal-Souza, André (2016): No light at the end of the tunnel: Shading impacts
- 588 on biological communities from subtropical rocky shores. figshare.
- 589 https://dx.doi.org/10.6084/m9.figshare.3205285.v1
- 590 Perkol-Finkel, S., Ferrario, F., Nicotera, V. & Airoldi, L. (2012) Conservation
- challenges in urban seascapes: promoting the growth of threatened species on
- coastal infrastructures. *Journal of Applied Ecology*, **49**, 1457–1466.
- 593 Quinn, J.M., Cooper, A.B., Stroud, M.J. & Burrell, G.P. (1997) Shade effects on
- 594 stream periphyton and invertebrates: An experiment in streamside channels.
- New Zealand Journal of Marine and Freshwater Research, **31**, 665–683.

- Ruiz, J.M. & Romero, J. (2001) Effects of in situ experimental shading on the
- 597 Mediterranean seagrass Posidonia oceanica. Marine Ecology Progress Series,
- 598 **215**, 107–120.
- 599 Sanger, D.M., Holland, A.F. & Gainey, C. (2004) Cumulative impacts of dock
- 600 shading on Spartina alterniflora in South Carolina estuaries. Environmental
- 601 *Management*, **33**, 741–748.
- Saunders, R.J. & Connell, S.D. (2001) Interactive effects of shade and surface
- orientation on the recruitment of spirorbid polychaetes. Austral Ecology, 26,
- 604 109–115.
- Serôdio, J., Vieira, S. & Cruz, S. (2008) Photosynthetic activity, photoprotection
- and photoinhibition in intertidal microphytobenthos as studied in situ using
- variable chlorophyll fluorescence. *Continental Shelf Research*, **28**, 1363–1375.
- 608 Shafer, D.J. (1999) The effects of dock shading on the seagrass Halodule
- 609 wrightii in Perdido Bay, Alabama. Estuaries, **22**, 936–943.
- 610 Small, C. & Nicholls, R.J. (2003) A global analysis of human settlement in
- coastal zones. *Journal of Coastal Research*, **19**, 584–599.
- 612 Struck, S.D., Craft, C.B., Broome, S.W., Sanclements, M.D. & Sacco, J.N.
- 613 (2004) Effects of bridge shading on estuarine marsh benthic invertebrate
- 614 community structure and function. *Environmental Management*, **34**, 99–111.
- Takada, Y. (1999) Influence of shade and number of boulder layers on mobile
- organisms on a warm temperate boulder shore. Marine Ecology Progress
- 617 Series, **189**, 171–179.
- Tanaka, M.O., Duque-Estrada, T.E. & Magalhães, C.A. (2002) Dynamics of the
- 619 acmaeid limpet Collisella subrugosa and vertical distribution of size and

- abundance along a wave exposure gradient. Journal of Molluscan Studies, 68,
- 621 55–64.
- Thompson, R.C., Norton, T.A. & Hawkins, S.J. (2004) Physical stress and
- 623 biological control regulate the producer-consumer balance in intertidal biofilms.
- 624 *Ecology*, **85**, 1372–1382.
- Thorson, G. (1964) Light as an ecological factor in the dispersal and settlement
- of larvae of marine bottom invertebrates. *Ophelia*, **1**, 167–208.
- Van Gils, M. & Klijn, E.H. (2007) Complexity in decision making: The case of the
- 628 Rotterdam harbour expansion. Connecting decisions, arenas and actors in
- spatial decision making. *Planning Theory & Practice*, **8**, 139–159.
- Vaselli, S., Bulleri, F. & Benedetti-Cecchi, L. (2008) Hard coastal-defence
- 631 structures as habitats for native and exotic rocky-bottom species. Marine
- 632 Environmental Research, 66, 395–403.
- 633 Vermeij, G.J. (1973) Morphological patterns in high-intertidal gastropods:
- adaptive strategies and their limitations. *Marine Biology*, **20**, 319–346.
- Williams H., Messier, C. & Kneeshaw D.D. (1999) Effects of light availability and
- 636 sapling size on the growth and crown morphology of understory Douglas-fir and
- lodgepole pine. Canadian Journal of Forest Research, **29**, 222–231.
- 638 Williams, G.A. (1994) The relationship between shade and molluscan grazing in
- 639 structuring communities on a moderately-exposed tropical rocky shore. *Journal*
- of Experimental Marine Biology and Ecology, **178**, 79–95.
- Young, C.M. & Chia, F.S. (1984) Microhabitat-associated variability in survival
- 642 and growth of subtidal solitary ascidians during the first 21 days after
- settlement. *Marine Biology*, **81**, 61-68.

Supporting Information

645	TABLES	3
0.0		_

Table 1. Effect of habitats (unshaded and shaded) (2-way ANOVA) on primary producers and sedentary grazers on subtropical rocky shores. **P < 0.01; ***P

< 0.001; ns = not significant. \D ata transformed to \D n (x +1)

652	Effect	df	MS	F	P	df	MS	F	Р
653				Primar	y producers				
654			Biofilm biomas	ss	Macro	algae b	iomass		
655	Habitat = H	1	0.004	0.16	0.72	1	4.02	137.25	· **
656	Location = L	2	0.094	3.84	0.20	2	0.49	16.92	0.05
657	H*L	2	0.024	2.92	0.06	2	0.03	0.25	0.78
658	Error	54	0.008			24	0.11		
659	Cochran's tes	it	C = 0.4	47 (***)			C = 0.4	45 (ns)	
660				Graze	rs body size				
661			L. subrugosa	L. subrugosa					
662	Habitat = H	1	0.52	0.001	0.97	1	34.66	3.28	0.32
663	Location = L	1	11.52	0.027	0.89	1	38.18	3.61	0.30
664	H*L	1	416.12	42.56	***	1	10.55	11.95	***
665	Error	1066	9.77			606	0.88		
666	Cochran's test $C = 0.5$.51 (***) $C = 0.39 (***)$					
667				Graze	rs abundance				
668			L. subrugosa			E. line	olata§		
669	Habitat = H	1	2,226.05	5.26	0.26	1	1.97	2.17	0.28
670	Location = L	1	61.25	0.14	0.76	2	2.99	3.30	0.23
671	H*L	1	423.20	3.37	0.07	2	0.90	9.79	***
672	Error	76	125.46			54	0.09		
673	Cochran's tes	st	C = 0.4	47 (***)			C = 0.2	28 (ns)	
674									

Table 2. Effects of habitats (unshaded and shaded) (PERMANOVA) on sessile macrobenthic communities on different zonation ranges in subtropical rocky shores. All data was transformed to arcsine $(\sqrt{x}) + 1$. ***P < 0.001

Effect	df	MS	Pseudo-F	P	df	MS	Pseudo-F	P	df	MS	Pseudo-F	P
		(a) Infralitto	oral fringe			(b) Lower m	nesolittoral			(c) Upper r	nesolittoral	
Habitat = H	1	315.96	1.77	0.24	1	138.69	0.55	0.60	1	150.98	0.63	0.57
Location = L	2	156.02	7.36	***	2	1,521.10	147.30	***	2	376.64	21.03	***
H*L	2	177.87	8.40	***	2	252.02	26.06	***	2	240.12	13.41	***
Error	54	21.17			114	1,102.40			54	17.91		
			Pair-	wise tes	ts 'unsł	naded' <i>vs.</i> 'sh	aded' inside Lo	cation				
		(a) Infralitto	oral fringe			(b) Lower m	nesolittoral			(c) Upper r	nesolittoral	
		t	P			t	P			t	P	
RS1		3.06	***			1.02	0.36			1.44	0.16	
RS2		3.43	***			4.84	***			5.31	***	
RS3		3.30	***			4.69	***			0.40	0.80	

Table 3. Effects of treatments (repeated measures ANOVA) on abundance of primary producers, sessile invertebrates and open space on shading manipulation in the upper mesolittoral and infralittoral fringe. $^*P < 0.05$; $^{**}P < 0.01$; $^{***}P < 0.001$; § P-values corrected by Greenhouse-Geisser adjustment. All response variables except biofilm were transformed to arcsine (\sqrt{x})

Effect	df	MS	F	P	MS	F	P	MS	F	Ρ
			Upper	mesolittoral						
		Biofilr	m		Oyste	ers§		C. bis	inuatus	§
Treatment = Tr	2	0.0240	8.87	**	0.60	12.31	***	0.62	4.13	*
Error	12	0.0028			0.05			0.15		
Time = Ti	7	0.0058	3.79	**	0.02	1.48	0.25	0.70	50.40	***
Tr*Ti	14	0.0021	1.36	0.19	0.10	6.45	**	0.05	4.10	**
Error	84	0.0015			0.01			0.01		
			Infralit	toral fringe						
		Macro	oalgae			Open	space			
Treatment = Tr	2	5.75	64.22	***	5.16	61.05	***			
Error	12	0.09			0.08					
Time = Ti	5	0.98	30.87	***	0.94	29.09	***			
Tr*Ti	10	0.34	10.64	***	0.32	10.03	***			
Error	60	0.03			0.03					

Table 4. Effects of treatments (2-way ANOVA) on recruitment rate of oysters and the barnacle *Chthamalus bisinuatus* on shading manipulation in the upper mesolittoral. $^*P < 0.05$; $^{**}P < 0.01$; $^{***}P < 0.001$

Effect	df	MS	F	Р	MS	F	Ρ
	Chtha	amalus i	Oyste	rs			
Treatment = Tr	2	0.98	10.97	**	0.48	5.32	*
Time = Ti	5	1.34	15.01	***	0.10	1.07	0.42
Tr*Ti	10	0.09	0.84	0.58	0.09	1.79	0.07
Error	72	0.10			0.05		
Cochran's test	C = 0	.25 (***))	C = 0	.83 (***))	

FIGURE CAPTIONS

Fig. 1. Body size and abundance (mean + standard error) of sedentary grazers (a) Lottia subrugosa, (b) Echinolittorina lineolata and (c) Littoraria flava in unshaded and shaded habitats in subtropical rocky shores (RS). $^*P < 0.01$; $^{***}P < 0.001$; ns = not significant; according to SNK test (items a and b) or t-test (c).

Fig. 2. nMDS diagrams of macrobenthic communities of different zonation ranges in unshaded and shaded habitats in subtropical rocky shores (RS).

Fig. 3. Biofilm biomass (NDVI) (mean ± standard error) in treatments of shading manipulation in the upper mesolittoral.

Fig. 4. Cover percentage (mean ± standard error) of macroalgae, open space, oysters and the barnacle *Chthamalus bisinuatus* in treatments of shading manipulation. Post-hoc comparisons of treatments shown only for beginning and end of experiment. SNK test: ns = not significant; ***control = procedural control ≠ shaded.

Fig. 5. Cumulative recruitment (mean number of accumulated recruits in each time) of oysters and the barnacle *Chthamalus bisinuatus* in treatments of shading manipulation.

FIGURES

Figure 1

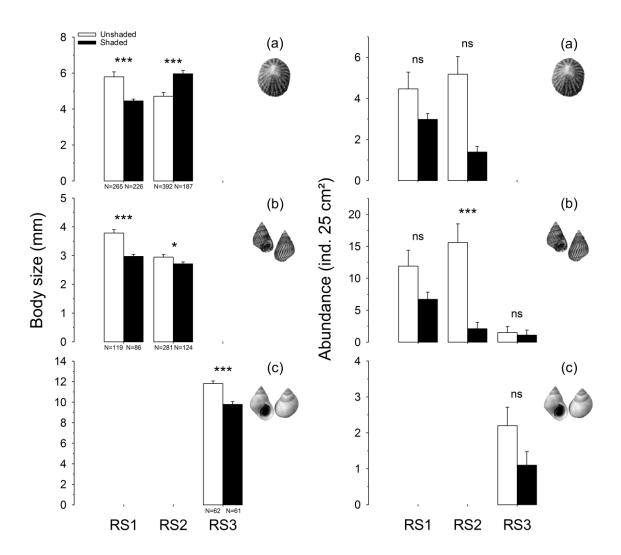


Figure 2

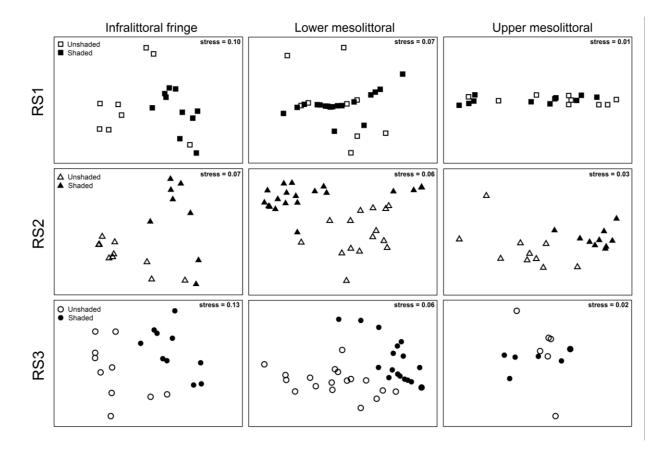


Figure 3

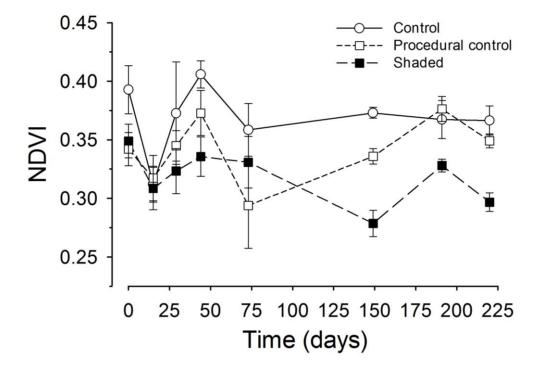


Figure 4

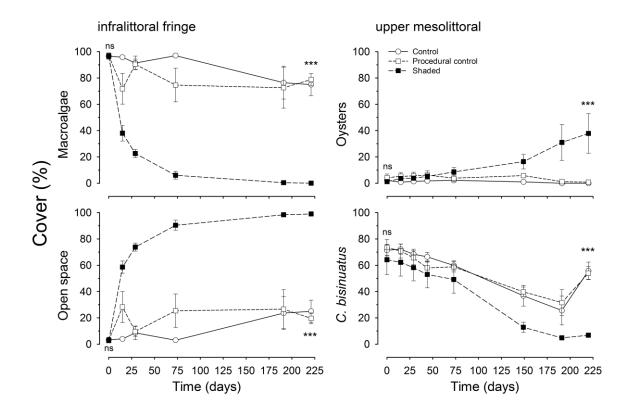


Figure 5

