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# Hydroacoustics for the discovery and quantification of Nassau grouper (*Epinephelus striatus*) spawning aggregations

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8 Abstract Fish spawning aggregations (FSAs) are vital 9 life-history events that need to be monitored to determine 10 the health of aggregating populations; this is especially true 11 of the endangered Nassau grouper (Epinephelus striatus). 12 Hydroacoustics were used to locate Nassau grouper FSAs 13 at sites on the west end of Little Cayman (LCW), and east 14 ends of Grand Cayman (GCE) and Cayman Brac (CBE). 15 Fish abundance and biomass at each FSA were estimated 16 via echo integration and FSA extent. Acoustic mean fish 17 abundance estimates  $(\pm SE)$  on the FSA at LCW 18  $(893 \pm 459)$  did not differ significantly from concurrent SCUBA estimates (1150  $\pm$  75). Mean fish densities 19 20 (number  $1000 \text{ m}^{-3}$ ) were significantly higher at LCW 21  $(33.13 \pm 5.62)$  than at the other sites (GCE: 7.01  $\pm$  2.1, 22 CBE: 4.61  $\pm$  1.16). We investigate different acoustic post-23 processing options to obtain target strength (TS), and we 24 examine the different TS to total length (TL) formulas 25 available. The SCUBA surveys also provided measures of 26 TL through the use of laser callipers allowing development

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of an in situ TS to TL formula for Nassau grouper at the 27 LCW FSA. Application of this formula revealed mean fish 28 29 TL was significantly higher at LCW (65.4  $\pm$  0.7 cm) than GCE (60.7  $\pm$  0.4 cm), but not CBE (61.1  $\pm$  2.5 cm). Use 30 of the empirical TS to TL formula resulted in underesti-31 mation of fish length in comparison with diver measure-32 ments, highlighting the benefits of secondary length data 33 and deriving specific TS to TL formulas for each popula-34 35 tion. FSA location examined with reference to seasonal marine protected areas (Designated Grouper Spawning 36 Areas) showed FSAs were partially outside these areas at 37 GCE and very close to the boundary at CBE. As FSAs 38 often occur at the limits of safe diving operations, 39 hydroacoustic technology provides an alternative method 40 to monitor and inform future management of aggregating 41 42 fish species.

KeywordsHydroacoustics · Nassau grouper (Epinephelus44striatus) · Fish spawning aggregations (FSAs) · Echo45integration46

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### Introduction

Fish spawning aggregations (FSAs) are broadly defined as 48 'a group of conspecific fish gathered for the purposes of 49 spawning with fish densities significantly higher than are 50 found during the non-reproductive periods' (Domeier and 51 Colin 1997). This reproductive strategy creates temporary 52 concentrations of fish (Johannes 1978; Kobara and Heyman 53 54 2008) that are highly susceptible to overfishing (Nemeth 2005; Starr et al. 2007; Sadovy de Mitcheson and Erisman 55 2012). The health of a FSA is a good indicator of the health 56 of the population as a whole (Gascoigne 2002), and any 57 depletion of a FSA has serious consequences for the 58

59 reproductive output of that population (Sadovy and Domeier 2005; Sadovy de Mitcheson 2016). FSAs there-60 61 fore are important life-history phenomena that must be 62 considered in any efforts to manage fisheries of aggregat-63 ing species (Sadovy and Colin 2012; Sadovy de Mitcheson 2016). We use the term FSA for fish that are gathered 64 together for the purpose of spawning. We acknowledge, 65 66 however, that the aggregations of fish detected may not 67 have been spawning per se at the specific times of the 68 surveys.

69 One of the best known examples of the demise of a 70 species due to FSA over fishing is that of the Nassau 71 grouper (Epinephelus striatus) (Sadovy de Mitcheson et al. 72 2008). These large top-level predators are an important 73 species within Caribbean reef ecosystems (Stallings 74 2008, 2009; Archer et al. 2012). Nassau grouper migrate to 75 specific sites during periods of winter full moons to reproduce in FSAs (Sala et al. 2001; Whaylen et al. 2004; 76 Starr et al. 2007) and were one of the first large-bodied 77 78 tropical reef-fish species scientifically documented to do so 79 (Smith 1972). It is estimated that 75% of all known Nassau 80 grouper spawning aggregations have either been eradicated 81 or reduced to negligible numbers (Sadovy de Mitcheson 82 et al. 2008). Following over-exploitation, these aggregations often fail to recover (Gibson 2007; Semmens et al. 83 84 2007), although recent evidence suggests that effective 85 management can lead to population increases (Kadison et al. 2010; Heppell et al. 2012). FSAs in the Cayman 86 87 Islands have been reported on the eastern and southwest 88 points of Grand Cayman, the northeast and southwest 89 points of Little Cayman and the southwest point of Cayman Brac (Bush et al. 2006). These sites were protected by 90 91 legislation in 2003 which prohibits fishing in these areas 92 (Whaylen et al. 2006), and due to winter spawning, it is 93 now forbidden to take a Nassau grouper from Cayman 94 waters during the months of December to April (Cayman 95 Islands Government 2016).

#### 96 Monitoring spawning aggregations

9 AQ1 Monitoring an FSA is an effective way to determine the 98 health of an aggregating population, but adequately mon-99 itoring an FSA requires a clear understanding of its loca-100 tion, extent, and dynamics. In-water monitoring is fraught 101 with difficulties including high temporal variability in fish 102 numbers and variable distribution across multiple sites, the expense of underwater visual census (UVC) surveys and 103 104 challenging underwater working conditions (including 105 strong currents, poor visibility and FSA locations below 106 safe diver depth limits) (Sadovy and Domeier 2005). This 107 is especially true in the Cayman Islands where FSAs occur 108 on the extreme tips of the islands at locations where cur-109 rents are strong and dives must occur at dawn and dusk to coincide with periods of peak fish activity. Further,110observer bias may be present in UVC surveys and fish may111avoid divers (Colin 1992; Murphy and Jenkins 2010).112

Hydroacoustics may be useful for assessing aggregating 113 reef fishes that are otherwise difficult to count (Johannes 114 et al. 1999). One of the main advantages of hydroacoustics 115 is the ability to collect large volumes of information in a 116 short amount of time (Trenkel et al. 2011: Jones et al. 117 2012). Further, unlike video or UVC, the acoustic tech-118 nique is unaffected by underwater visibility (Gledhill et al. 119 1996) nor are the fish influenced by the presence of a diver. 120 To date there has been limited use of hydroacoustics to 121 monitor spawning aggregations (e.g. Johnston et al. 2006; 122 Taylor et al. 2006; Ehrhardt and Deleveaux 2007) and 123 Taylor et al. (2006) noted the technology can provide an 124 accurate estimate of overall fish abundance and spatial 125 extent in comparison with diver visual counts. Studies 126 comparing hydroacoustics and UVC are sparse, however. 127 Taylor et al. (2006) reported similar acoustic density and 128 diver estimates over their entire survey region, although 129 total abundances differed likely due to differences in area 130 covered by the two methods and the patchy distribution of 131 the fish. Although hydroacoustic techniques hold great 132 promise, many authors highlight that ground-truthing is 133 required to identify the fish to species level (Simmonds and 134 MacLennan 2005; Ryan et al. 2009). 135

The International Union for the Conservation of Nature 136 (IUCN) lists the Nassau grouper as endangered and rec-137 ommends annual monitoring at as many traditional aggre-138 gation sites as possible, including adjacent areas where 139 aggregations have not previously been reported and as part 140 of the assessment of the effectiveness of protected areas 141 (Carpenter et al. 2015). Given the need to develop effective 142 monitoring techniques that can rapidly, effectively, and 143 quantitatively assess FSA status, we investigated the 144 capacity of hydroacoustics to address these recommenda-145 tions. We examined FSA locations in relation to protected 146 zones in the Cayman Islands and compared acoustic data 147 with diver-collected data. Further, we evaluated the dif-148 ferent acoustic processing methods available to estimate 149 the sizes of fish within FSAs. 150

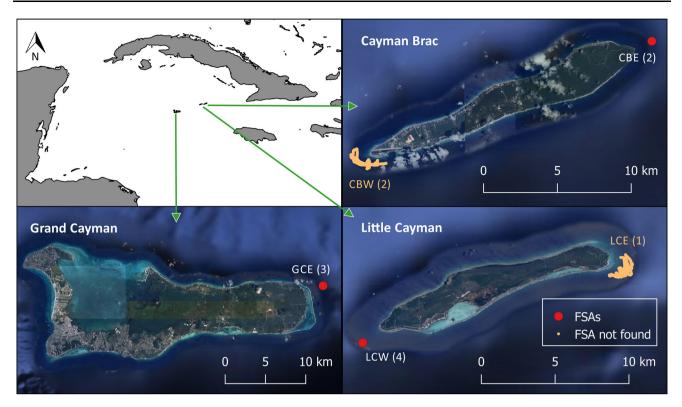
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#### Survey sites

The sites chosen in this study are all within the Designated153Grouper Spawning Areas (DGSA) of the Cayman Islands.154Surveys were focussed on the likely areas of the FSA,155based on site geomorphology and from local knowledge156via the Department of Environment (DoE) (Fig. 1). Most157survey effort was concentrated on the FSA located at the158



**Fig. 1** Areas in the Cayman Islands surveyed by hydroacoustics and in-water assessment techniques The numbers at each site represent the total number of hydroacoustic surveys undertaken at each location.

Table 1Dates and times of thesurveys conducted, with thenumber of days elapsed sincethe February full moon

*Red dots* show located fish spawning aggregations (FSAs); *peach colour* shows survey tracks that did not locate FSAs. Map data ©2016 Google

Survey name	Date	Start time	Stop time	Days after full moor	
GCE1	14/02/2014	12:40:43	15:19:39	0	
LCE1	15/02/2014	17:48:01	19:33:54	1	
LCW1	16/02/2014	12:04:38	12:52:39	2	
LCW2	16/02/2014	17:38:42	17:51:19	2	
LCW3	16/02/2014	18:38:18	19:12:52	2	
LCW4	17/02/2014	13:24:40	13:55:05	3	
CBW	17/02/2014	17:05:56	18:25:45	3	
CBE	18/02/2014	17:44:52	19:00:25	4	
CBW2	18/02/2014	10:43:05	13:04:03	4	
CBE2	19/02/2014	07:43:09	08:48:04	5	
GCE2	19/02/2014	17:13:11	18:28:32	5	
GCE3	20/02/2014	08:13:58	09:41:08	6	

Times are in Easter Standard Time (EST) (UTC/GMT -5 h)

west end of Little Cayman (LCW) as this is known to be
the most active of the FSAs, and for which concurrent fish
abundance and size data obtained via SCUBA were provided by the Grouper Moon project (http://www.reef.org/
groupermoonproject). Surveys were also conducted at

164 Little Cayman East (LCE), Grand Cayman East (GCE) and

165 Cayman Brac West (CBW) and East (CBE). The field

166 surveys in Cayman occurred between 14 and 20 February

167 2014 (Table 1).

#### Equipment

A Biosonics DTX split-beam echosounder with a 200-kHz 169 transducer (beam opening angle of  $6.8^{\circ}$ ), pole mounted 170 over the side of the survey vessel, was used for the surveys. 171 Data were collected with Biosonics visual acquisition 172 software (Biosonics Inc., Seattle, WA). Pulse duration was 0.4 ms, and the specified ping rate was 10 s<sup>-1</sup>. Survey 174 speed was kept to approximately 4 kn and sea state was 175

176 calm (Beaufort scale 3 or under) on all surveys. The echosounder was calibrated before the start of the surveys 177 178 on 13 February 2014 using a tungsten carbide 36-mm standard calibration sphere, following the standard meth-179 180 ods (Foote et al. 1987; Demer et al. 2015). The acoustic 181 return from the sphere was within acceptable tolerance to 182 the expected value given for the local environmental settings (TS = -39.6 vs. -39.8 dB, respectively (Biosonics 183 184 2004), with speed of sound calculated as  $1521.54 \text{ m s}^{-1}$ ). 185 Where diver observations were not available for species ground-truthing, underwater video was used (Thomas and 186 187 Thorne 2003; Doray et al. 2007; Jones et al. 2012). This consisted of a Sony 37CSHR camera with a live surface 188 189 feed mounted on an aluminium wing. Both the acoustic 190 data and the video data were time-stamped allowing 191 syncing of the visual and acoustic records in post-192 processing.

#### 193 Data processing

Potential Nassau grouper FSAs were initially identified through their stronger backscattering properties and school morphology (Fig. 2) than aggregations of other species (e.g. horse-eye jack, *Caranx latus*) and then verified by visual observation either by the use of the pelagic tow camera or through confirmation by the dive team at LCW.

200 Data were processed with the software package Sonar5-201 Pro (Balk and Lindem 2006), following the software-gui-202 ded analysis routine (see Parker-Stetter et al. 2009 for 203 details). The analysis was based upon echo integration 204 (also known as Sv/TS scaling) which divides the average reflection from all fish over a segment (the volume 205 206 backscattering coefficient, Sv) by the average target 207 strength (TS) from individual fish (Winfield et al. 2011). TS is defined as TS = mLogL + b where m and b are 208 209 constants for a given species and frequency, respectively, 210 and L =length as total length (TL), (Simmonds and MacLennan 2005). Initially, a threshold of -60 dB was 211 212 applied to the echograms to distinguish fishes from other 213 particulate targets such as plankton. This is a typical threshold applied for the detection of pelagic schooling 214 215 fishes (Reid 2000). Any noise due to issues such as bubbles 216 in the water column from wave action was removed by eye. 217 Sonar5 applies a time-varied gain correction of 218  $40\log(R)$  for TS values and  $20\log(R)$  for Sv values (Balk 219 and Lindem 2006). A bottom exclusion layer of 1 m was applied, and data from within this layer were not included 220 221 in the analysis due to the 'acoustic dead zone' (Ona and 222 Mitson 1996). For echo integration methodology, there are 223 two main options to obtain TS: using tracked fish as a 224 source or using 'single echoes detected' (SED) as source. 225 We used tracked fish as source to derive abundance esti-226 mates but examined the efficacy of both options to derive TS. We used the following criteria to track fish within the 227 FSAs: a minimum track length of three pings; a maximum 228 ping gap of two pings: a gating range of 0.3 m; a maximum 229 mean echo threshold of -25 dB; and a minimum mean 230 echo threshold of -40 dB. Due to difficulties in obtaining 231 232 sufficient numbers of tracks from within FSAs (likely due to high fish density and low signal-to-noise ratios in dense 233 areas of the aggregation), tracks were extracted and stored 234 from all passes of the FSAs per survey and then the tracked 235 fish were used to provide the survey-specific abundance 236 estimates. As tilt angle of fish can have a significant 237 bearing on TS, extreme tilt angles were filtered out of the 238 data following Gauthier and Horne (2004), so that any fish 239 with an aspect  $\pm 40^{\circ}$  from horizontal (dorsal aspect) were 240 removed from the analysis. We examined both the mean 241 TS of fish echoes in each track (calculated in the linear 242 domain) and the 75th percentile of TSs of each track. For 243 fish TS estimates using SED as source, SED were extracted 244 for each pass of an FSA and mean TS values subsequently 245 determined for the FSA from each survey. To assess 246 whether fish near the top of a school were shadowing those 247 beneath them, data were checked to ensure that echo 248 energy was consistent from the top to the bottom of the 249 school following Knudsen et al. (2009) (see electronic 250 supplementary information, ESM, Fig. S1). 251

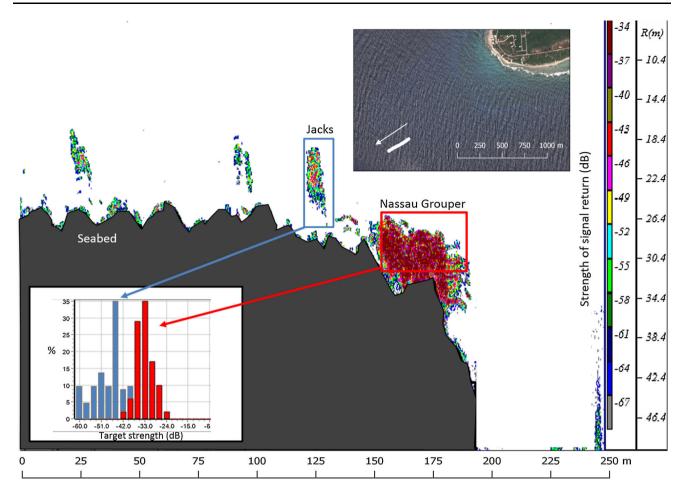
Three main equations were examined to convert TS to 252 fish TL by applying our mean TSs values (Table 2). Fur-253 ther, we scaled diver fish length (TL) measurements (taken 254 using a laser calliper system; Heppell et al. 2012) by our 255 mean TS data from tracked fish for the LCW FSA, by 256 sorting both datasets by increasing value and then plotting 257 one against the other to determine a survey-specific TS-TL 258 formula (see ESM Fig. S2) resulting in Formula 4 in 259 Table 2. 260

TL—weight regressions specific to the Nassau xo2 61 grouper—were used to calculate weight at TL for biomass 262 estimates using the formula W = aLb where W = weight 263 (g), L = TL (cm), a = 0.01122, b = 3.05 (Froese and 264 Pauly 2016). 265

Applying the TS-TL formula and then using the specific266TL-to-weight relationship for the Nassau grouper (Froese267and Pauly 2016) give the mean weight of fish in each FSA.268This number was then multiplied by the number of fish269estimated in each FSA to provide total biomass estimates270for each FSA surveyed.271

#### Spatial extents

Once the FSA was located using preliminary acoustic273transects, the aggregation was surveyed from different274angles to corroborate its extent. This approach follows275Doonan et al. (2003), who noted the advantages of a star-276shaped survey track in hydroacoustic surveys over277



**Fig. 2** An example echogram of the analysis of fish echoes resulting from a Nassau grouper fish spawning aggregation (FSA) (*red*) and those from an aggregation of horse-eye jacks (*blue*). The *inset* shows that grouper had a higher percentage of stronger echoes. Transect distance is shown along the *x*-axis, while depth [R(m)] and strength of

signal return (*colour strip*) are shown on the *y-axis*. The satellite image shows the location of the transect of the Little Cayman west (LCW) 1 survey, and the *arrow* shows the direction of travel. Map data ©2016 Google

Table 2 Target strength (TS) to length (L) formulae examined in this study

	Formula TS to L	Formula L to TS	Reference	Species	Frequency (kHz)
1	$TS = 19.1 \log 10(L) - 64.07$	L = (2261.8) * EXP[0.1206*(TS)]	Love (1971)	Multi species	200
2	TS = 0.7091 * L - 89.136	L = (TS/0.7091) + 89.136	Ehrhardt and Deleveaux (2007)	Epinephelus striatus	200
3a	$TS = 19.2 \log 10(L) - 64.05$	L = (2165)*EXP[0.12*(TS)]	Rivera et al. (2010)	Epinephelus guttatus	120
$3b^{a}$	$TS = 19.2 \log 10(L) - 64.25$	L = (2220)*EXP[0.1199*(TS)]	Rivera et al. (2010)	Epinephelus guttatus	200
4	$TS = 27.6 \log 10(L) - 147.32$	L = (207.06) * EXP[0.0362*(TS)]	This study	Epinephelus striatus	200

Length is total length in cm

<sup>a</sup> 3b is 3a reformulated for 200 kHz

278 schooling fishes. Alongside fish abundance values, the 279 geographical extents were also extracted, but these are 280 given in only two dimensions (height and length). Where 281 survey tracks crossed the FSA from different angles, the full 282 three-dimensional extent of the FSA was estimated by 283 drawing a polygon (Fig. 3) as per the arithmetic extrapo-284 lation method used by Taylor et al. (2006) and Ehrhardt and Deleveaux (2007). When the track crossed the FSA from285only one angle, it was assumed that the aggregation was286circular unless nearby pings showed no fish were present, in287which case the halfway point between the positive (FSA288detected) and negative (FSA not detected) pings was taken289to demarcate the FSA extent. If the FSA represented two or290more clear densities, separate polygons were drawn for each291

292 density class present. Once a polygon was drawn, fish 293 abundance was calculated by multiplying the mean number 294 of fish ha<sup>-1</sup> by the area of the polygon. When there were 295 multiple polygons of differing abundances, the result of 296 each was summed to give a total number of fish.

#### 297 Statistical analyses

298 Welch's ANOVAs (equal variances were not assumed) were used to compare fish densities (number of fish 1000  $\text{m}^{-3}$ , log 299 transformed) among sites and surveys at LCW, and a two-300 301 sample t test was used to compare densities at GCE surveys. 302 Diver fish abundance estimates were compared to the 303 acoustic abundance estimates by using a two-sample t test. 304 The TS values from the different acoustic processing meth-305 ods were compared for each site with two-sample t tests. 306 Values of fish TL gained from applying tracked fish mean TS 307 data coupled with our in situ formula were compared among 308 the different surveys and sites with Welch's ANOVA, and 309 Games-Howell pairwise comparisons were used to test 310 where the differences among sites existed.

#### 311 Results

#### 312 Numbers of fish in each FSA

313 FSAs were identified at LCW (all four surveys), GCE (two

314 of three surveys) and CBE (one of two surveys). No FSAs

were detected in the surveys of CBW or LCE. Visual 315 confirmation that the targets were Nassau grouper was 316 provided by the Grouper Moon dive team at LCW and at 317 GCE by the towed camera system. We did not achieve 318 visual confirmation of species present at CBE; however, 319 320 mean TS's and FSA morphology at that location were similar to those at the verified Nassau grouper FSA sites. 321 The highest acoustically measured fish abundance was 322 detected at LCW with a maximum abundance of 2194 fish 323 in the aggregation (survey LCW1) 2 d after the full moon 324 on 16 February 2014. Fish density was significantly greater 325 at LCW FSA than at the other two sites ( $F_2 = 25.49$ , 326 p = 0.000) which did not differ significantly from each 327 other. Fish densities did not differ significantly among 328 individual surveys at the LCW FSA ( $F_3 = 1.35$ , 329 p = 0.319) or the GCE FSA  $(T_8 = 1, p = 0.349)$ 330 (Table 3). 331

## Comparison between acoustic and diver abundance332data333

Diver-estimated numbers of fish at the LCW FSA were 334 made concurrent with acoustic surveys LCW2, LCW3 and 335 LCW4 (Table 3). Diver confirmation of species also occurred during LCW1, although numbers could not be recorded. No significant difference was detected at the 95% 338 confidence level between diver estimates and acoustics  $(T_3 = 0.55, p = 0.619)$ . 340

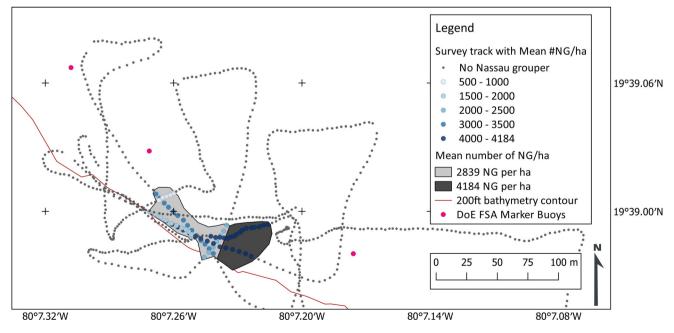


Fig. 3 Example of fish spawning aggregation (FSA) polygon determination in the arithmetic extrapolation method during the Little Cayman west (LCW) 4 survey. *NG* Nassau grouper. Department of

Environment Little Cayman FSA location marker buoys shown in *pink* and the 200 ft bathymetry contour shown in *brown. Crosses* indicate where latitude and longitude intersect

#### 341 Fish TS

342 Mean fish TS gained through tracked fish was compared 343 with mean fish TS via SED for each site (Fig. 4). There 344 was no significant difference in mean TS values at any site 345 (CBE:  $T_{12} = 0.03$ , p = 0.98, LCW:  $T_{47} = 1.44$ , p =0.157, GCE:  $T_{28} = 0.59$ , p = 0.557). The TS values from 346 the 75th percentile of echoes in a fish track were signifi-347 348 cantly higher than the mean TS at LCW ( $T_{192} = 3.78$ , 349 p = 0.000) and GCE ( $T_{429} = 6.91$ , p = 0.000), but not at 350 CBE  $(T_{19} = 1.13, p = 0.273)$  presumably due to the 351 smaller number of observations reducing statistical power.

#### 352 Converting TS to TL

Mean TS measurements from tracked fish were scaled by the diver LCW FSA diver length data. This resulted in: TS = 27.6log10(L) – 147.32 ( $R^2$  = 0.98; ESM Fig. S2). The results from applying this formula to TS data are plotted for the LCW dataset alongside the alternative equations given in Table 2 (Fig. 5).

The results of applying our in situ formula to the acoustic TS data are plotted per individual survey (Fig. 6a) and as mean values per site (Fig. 6b).

362 There was a significant difference in mean fish TL 363 calculated from mean TS of tracked fish between the sites  $(F_2 = 15.08, p = 0.000)$ , with significantly larger fish at 364 LCW than at GCE but not CBE, which did not differ from 365 366 each other. Using the von Bertalanffy growth curve for the 367 Nassau grouper sampled from aggregations in the Cayman Islands 1987–1992 (Bush et al. 2006), the estimated mean 368 369 fish TL of  $65.4 \pm 0.7$  cm seen at the LCW FSA corre-370 sponds to an age of 10 yr. The estimated mean sizes of fish

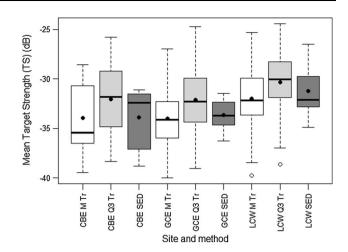


Fig. 4 Mean fish target strength (TS) found in fish spawning aggregations during each survey, per site and for each of the acoustic processing methods. *CBE* Grand Cayman Brac, *GCE* Grand Cayman east, *LCW* Little Cayman west, *Tr M* mean echo of tracked fish, *Q3 Tr* 75th percentile of echoes from tracked fish, *SED* single echoes detected. *Box plots* show mean values (*black circle*), median values (*solid horizontal line*), and the *lower and upper* ends of the box are the 25 and 75% quartiles, respectively. The whiskers indicate 1.5 times the inter-quartile range, and points beyond this range are shown by *empty circles* 

at	the	GCE	FSA	$(60.7 \pm 0.4 \text{ cm})$	and	CBE	371
(61.	$1 \pm 2$	.5 cm) c	orrespo	nd to those of 8-year	ar-old f	ìsh.	372

# FSA location relative to Cayman Islands DoE373Designated Grouper Spawning Areas374

The extent of the FSA located on Grand Cayman fell on the 375 extreme northern limit of the DGSA boundary on the 376

 Table 3
 Estimates of mean TS, mean lengths, weights, fish numbers and subsequent biomass values per survey where a FSA was identified as derived from mean TS from tracked fish

Survey name	Mean TS (dB)	Mean length (cm)	Mean weight (g)	Fish number	Biomass (kg)	Verification method	Fish density (#/ 1000 m^3)	Fish number/ isonified volume (Nv)	Mean depth (m)
LCW1	-31.98 (0.86)	65.22 (2.06)	3900.03 (390.9)	2194	8556.67	D (NP)	46.89 (24.60)	0.095 (0.05)	28.0 (1.4)
LCW2	-32.89 (1.43)	63.60 (3.30)	3782.35 (598.3)	398	1505.37	D (1225)	24.69 (12.76)	0.051 (0.024)	28.9 (2.1)
LCW3	-32.62 (1.25)	63.94 (2.97)	3746.54 (559.0)	122	457.08	D (1225)	18.20 (5.29)	0.031 (0.007)	26.2 (2.6)
LCW4	-30.50 (0.84)	68.86 (2.11)	4615.64 (443.8)	857	3955.60	D (1000)	32.87 (21.50)	0.072 (0.046)	29.0 (2.6)
LCW all	-32.01 (0.61)	65.40 (1.44)	4018.20 (268.1)	893	3588.25	D	33.13 (11.02)	0.067 (0.023)	28.1 (1.1)
CBE1	-33.95 (2.26)	61.12 (5.08)	3327.10 (849.2)	58	192.97	NP	4.61 (2.27)	0.009 (0.005)	30.4 (1.9)
GCE2	-33.95 (0.55)	60.90 (1.2)	3208.22 (191.6)	49	157.20	TC	4.01 (2.24)	0.0198 (0.011)	43.7 (2.2)
GCE3	-34.07 (0.48)	60.61 (1.08)	3162.43 (181.7)	40	126.50	TC	8.37 (5.82)	0.042 (0.028)	46.1 (1.1)
GCE all	-34.01 (0.36)	60.74 (0.8)	3183.32 (131.6)	45	143.25	TC	7.01 (4.12)	0.035 (0.019)	45.2 (1.1)

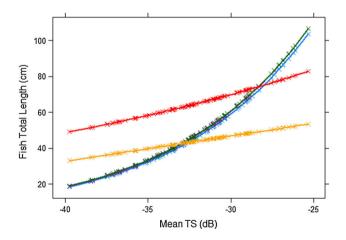
Fish density is number of fish per 1000 m<sup>3</sup>. Nv is number of fish per volume isonified (Sawada et al. 1993). Verification method shows how the fish were identified D diver (number in brackets), NP not possible, TC towed camera. Mean depth is the mean fish depth at each FSA. Numbers in brackets are 95% confidence levels

GCE2 survey and just outside the boundary during the
GCE3 survey. At CBE, the FSA was just within the
boundary close to its northern limit. The LCW FSA was
within the associated protection zone (Fig. 7).

#### 381 Discussion

382 The greatest fish abundances and densities were recorded at 383 the LCW FSA. This is as expected as this particular FSA is 384 well known throughout the Caribbean for the high numbers 385 of fish present there during spawning periods (Whaylen et al. 2004). It should be noted that these surveys occurred 386 387 closest to the full moon (2-3 d after the full moon), when 388 Nassau grouper FSAs are most active (Starr et al. 2007). 389 The surveys LCW1 and LCW4 both yielded very similar 390 patterns of fish distribution and had the highest abundance 391 estimates. These surveys occurred at similar times near the 392 middle of the day, while surveys LCW2 and LCW3, both 393 occurring near dusk, recorded lower abundances. Other 394 studies have found that groupers were more densely 395 aggregated at sunrise and sunset (Whaylen et al. 2006), and 396 it is possible that the main aggregation may therefore have 397 been missed by surveys LCW2 and LCW3, or that abun-398 dance estimates are more robust when fish are more dis-399 persed as has been seen in other studies (Rudstam et al. 400 2003).

401 At any given time in the LCW FSA, some proportion of 402 the fish are located on the plateau and across a wider area 403 than is represented by the main aggregation at the reef crest 404 (Whaylen et al. 2006); it is possible that the acoustics may 405 not have detected these individuals. In addition, as fish



**Fig. 5** Target Strength (TS) data from the Little Cayman west (LCW) surveys and corresponding fish total length using the following empirical formulas:  $TS = 19.2\log_{10}(L) - 64.05$  (*blue*; Rivera et al. 2010):  $TS = 19.1\log_{10}(L) - 64.07$  (*pink*, partially hidden due to similar values as green; Love 1971),: TS = 0.7091 \* L - 89.136 (*yellow*; Erhardt and Deleveaux 2007),  $TS = 27.6\log_{10}(L) - 147.32$  (*red*, this study)

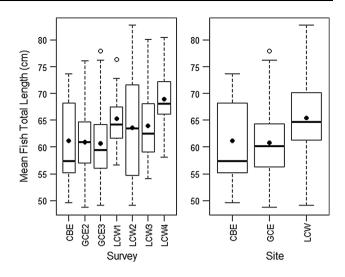
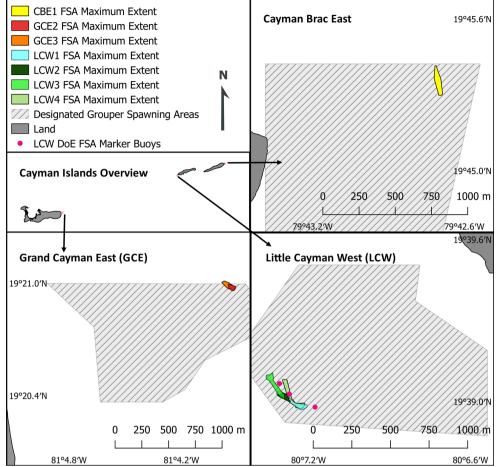


Fig. 6 Mean fish total length (TL) as calculated by applying our in situ formula **a** during each survey and **b** as grouped data per site. *Box plots* show median values (*solid horizontal line*), and the *lower* and upper ends of the box are the 25and 75% quartiles, respectively. The whiskers indicate 1.5 times the inter-quartile range and points beyond this range are shown by *empty circles* 

within 1 m of the seabed were not included in the study. 406 acoustic abundance estimates are best considered an index 407 of abundance rather than an absolute abundance and are 408 409 likely to be conservative compared to the total number of 410 all spawning fish. The LCW FSA was most active the day before the acoustic surveys (15 February, 1 d after the full 411 moon) with 4000 fish estimated by the dive team. Our peak 412 number of fish was detected the following day. The CBE 413 FSA was surveyed 4 d after the full moon, and the FSA at 414 415 GCE surveyed 5 and 6 d after the full moon; only small numbers of fish were found at either location. It is likely 416 that the acoustics results underestimate the total abun-417 dances of individuals in these FSAs as they do not account 418 for the most active times, i.e. closer to the full moon. 419 Therefore, we recommend that to fully evaluate a given 420 FSA, acoustic surveys should be conducted both over 421 422 several days and at multiple times per day to increase the 423 probability of capturing peak abundance at any given FSA. Note that we assumed that all echoes from within a FSA 424 were Nassau grouper, but it is possible that relatively low 425 numbers of other fish species were also present. 426

We evaluated the possibility of acoustic shadowing 427 leading to the differences between diver estimates and 428 acoustic estimates of fish numbers. No decrease in echo 429 energy from the top of the FSAs to the bottom was found, 430 indicating that the acoustic technique can be used to 431 accurately quantify fish in FSAs (Knudsen et al. 2009). 432 However, this is contrary to some other studies which have 433 434 reported a shadowing effect in dense schools of marine fishes (Zhao and Ona 2003; Utne and Ona 2006; Løland 435 et al. 2007). 436 Fig. 7Fish spawning<br/>aggregation locations and<br/>maximum extents detected via<br/>hydroacoustics in the Cayman<br/>Islands in relation to the<br/>positions of the Designated<br/>Grouper Spawning Areas<br/>(hatched area)CBE1 FSA Maximum Ext<br/>GCE2 FSA Maximum Ext<br/>GCE3 FSA Maximum Ext<br/>CLCW1 FSA Maximum Ext<br/>LCW2 FSA Maximum Ext<br/>LCW3 FSA Maximum Ext<br/>Designated Grouper Spa<br/>Land



43<sup>°</sup> AQ3 We examined three different methods in the acoustic post-processing to extract TS values, and it is interesting to 438 439 note that mean TS with SED as source did not differ sig-440 nificantly from the mean TS of tracked fish. When fish are 441 tilted further from the horizontal, TS is reduced so max TS 442 may be a better estimator than mean TS (Balk and Lindem 443 2006). However, to remove any effect of 'flash echoes' 444 (Lilja et al. 2004) and also the potential exaggerating effects on mean TS of multiple echoes (Soule et al. 1995; 445 446 Rudstam et al. 2003), a 75th percentile of the TS along a 447 tracked fish was also examined and unsurprisingly yielded 448 higher values overall than the other two methods. How-449 ever, we used the mean TS for subsequent calculations as 450 this method is most common in the literature (e.g. Guillard 451 et al. 2004; Rose 2009).

452 TS varies with tilt angle (Nielsen and Lundgren 1999), 453 and among fish species due to anatomical differences in the 454 size of the swim bladder (Simmonds and MacLennan 455 2005). Therefore, an empirical TS-TL relationship is 456 needed to convert TS to fish TL, which is known for many 457 species (Kracker 2007). Ideally, TS data should be 458 obtained from fish that are typical of the population to be 459 surveyed (Simmonds and MacLennan 2005). The LCW FSA presented a rare opportunity to do this as the fish 460 species (almost entirely Nassau grouper) could be deter-461 mined by divers who were also able to provide accurate 462 length measurements. By scaling our TS values by the 463 diver measurements, we derived an alternative in situ TS-464 TL equation allowing comparison to the other equations 465 examined. Application of either the Love (1971) or Rivera 466 et al. (2010) formula results in a significant underestima-467 tion of fish size in comparison with the diver data. 468 Although our equation contains a log function, it is more 469 similar to the Erhardt and Deleveaux (Ehrhardt and Dele-470 veaux 2007) than the other equations. This is likely to be 471 due to the relatively narrow range of fish sizes in both their 472 and our studies, as these are the lengths of reproductively 473 active fish. While applying our equation matches diver 474 lengths at LCW, we are hesitant to suggest without further 475 evaluation that it should be used in preference to other 476 equations in future studies due to a number of reasons. 477 First, there was a relatively narrow range of fish lengths 478 present in the FSA as seen by divers, and applying our 479 formula may have the effect of overestimating the size of 480 smaller fish and underestimating the size of larger fish 481 beyond the range experienced here. Second, there are 482 483 difficulties in extracting tracked fish TS data from the 484 centre of FSAs and it may be the case that the tracked fish, 485 more commonly located on the periphery of the aggrega-486 tion, may be of a different size or orientation than those in 487 the centre (Starr et al. 1995). Third, tracking fish is difficult 488 in vertical marine applications (Guillard et al. 2004), and 489 although we experienced calm sea states, vessel movement 490 is likely to have reduced the number of possible tracks and 491 increased variation in TS. We recommend further exami-492 nation of the TS-TL relationship for Nassau grouper and 493 that caged fish experiments, or similar, should be con-494 ducted across a larger range of fish sizes to obtain more 495 empirical data points from which a potentially more robust 496 equation can be determined. Future research examining the 497 novel combination of hydroacoustics and laser callipers 498 could prove useful for FSA monitoring and other assess-499 ments of fish populations. The effect of reproductive state 500 on TS of Nassau grouper would also be worthy of exami-501 nation, since the relationship of gonad size to swim bladder 502 volume of spawning sardines is as important as the rela-503 tionship of the swim bladder volume to fish length 504 (Machias and Tsimenidis 1995). Mean fish TL was sig-505 nificantly larger at LCW than at GCE, but not CBE. As younger fish tend to be smaller, a recovering population 506 507 may have a larger proportion of smaller fish (Heppell et al. 508 2012). Our results could indicate that the FSAs on GCE 509 and CBE may be recovering from previous exploitation (Bush et al. 2006) or that the generally smaller fish at those 510 511 locations are a result of larger fish being removed by 512 fishing.

513 Hydroacoustics allowed us to determine the location of 514 FSAs in three-dimensional space. Spawning aggregations 515 were consistently found just off the reef crest at around 516 30 m depth at LCW as has been described previously by 517 direct observation (Whaylen et al. 2004). The depths of 518 FSAs will be influenced by a number of factors such as 519 diurnal time of survey or lunar phase (Starr et al. 2007); 520 however, knowing the depths from our surveys may assist 521 managers in determining optimum future survey strategies. 522 The relatively deep FSA of GCE was also noted by Kobara 523 and Heyman (2008) and is most likely due to the spawning 524 suitability of the local geomorphologic characteristics at 525 the site. The depth at which this FSA occurs highlights the 526 difficulty of visual census approaches using SCUBA. FSAs 527 can move between repeat surveys within the same lunar 528 period, and some wider movement not detected in this 529 study could reasonably be expected. We recommend 530 including line fishing in the one-mile-radius restrictive 531 buffers around DGSAs or increasing the size of the DGSAs 532 as a further precautionary measure. If fishing occurs at the 533 edge of the protected areas, as is common practice fol-534 lowing closures to fishing (Kellner et al. 2007), it is possible that these FSAs, which may be recovering, could 535 still be at risk. 536

Hydroacoustics has proven capable of locating FSAs in 537 historic areas where it was unknown whether fish were still 538 aggregating. This also means that acoustics can be used to 539 540 search for aggregations in new locations and used in situ-541 ations when diving surveys are impractical or hazardous. We have shown that surveying FSAs with hydroacoustics 542 produces fish count information comparable to that from 543 544 diver estimates, and it provides additional information such 545 as fish size when ground-truthing is also provided, although further work is needed in this area. Repeating hydroa-546 coustics surveys could yield much information on how 547 exploited FSAs are recovering and could assist with the 548 vital monitoring of endangered aggregating populations. 549

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#### References

- Archer SK, Heppell SA, Semmens BX, Pattengill-Semmens CV, Bush PG, McCoy CM, Johnson BC (2012) Patterns of color phase indicate spawn timing at a Nassau grouper *Epinephelus striatus* spawning aggregation. Curr Zool 58:73–83
   566

   Balk H, Lindem T (2006) Sonar 4, Sonar 5, Sonar 6 e post-processing
   568
- Balk H, Lindem T (2006) Sonar 4, Sonar 5, Sonar 6 e post-processing systems. Operator manual. Lindem Data Acquisition, Oslo
- Biosonics (2004) Calibration of Biosonics Digital Scientific Echosounder using T/C calibration spheres. www.biosonicsinc.com/ doc\_library/docs/DTXcalibration2e.pdf
- Bush PG, Lane ED, Ebanks-Petrie GC, Luke K, Johnson B, McCoy C, Bothwell J, Parsons E (2006) The Nassau grouper spawning aggregation fishery of the Cayman Islands—an historical and management perspective. Proceedings of the Gulf and Caribbean Fisheries Institute 57:515–524
- Carpenter KE, Claro R, Cowan J, Sedberry G, Zapp-Sluis M (2015) *Epinephelus striatus*. The IUCN Red List of Threatened Species 2015: e.T7862A70324790
- Cayman Islands Government (2016) The National Conservation (General) Regulations Part 2.6. http://www.gov.ky/portal/pls/ portal/docs/1/12326595.PDF
- Colin PL (1992) Reproduction of the Nassau grouper, *Epinephelus striatus* (Pisces: Serranidae) and its relationship to environmental conditions. Environ Biol Fish 34:357–377
- Demer DA, Berger L, Bernasconi M, Bethke E, Boswell K, Chu D,<br/>Domokos R et al (2015) Calibration of acoustic instruments.<br/>Cooperative Research Report 326, International Council for the<br/>Exploration of the Sea, 133 pp587<br/>588<br/>589

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- Domeier ML, Colin P (1997) Tropical reef fish spawning aggregations defined and reviewed. Bull Mar Sci 60:698–726
- 593 Doonan I, Bull B, Coombs R (2003) Star acoustic surveys of localized
   594 fish aggregations. ICES J Mar Sci 60:132–146
  - Doray M, Josse E, Gervain P, Reynal L, Chantrel J (2007) Joint use of echosounding, fishing and video techniques to assess the structure of fish aggregations around moored fish aggregating devices in Martinique (Lesser Antilles). Aquat Living Resour 20:357–366
  - Ehrhardt NM, Deleveaux VKW (2007) The Bahamas' Nassau grouper (*Epinephelus striatus*) fishery—two assessment methods applied to a data-deficient coastal population. Fish Res 87:17–27
  - Foote KG, Knudsen HP, Vestnes G, MacLennan DN, Simmonds EJ
     (1987) Calibration of acoustic instruments for fish-density
     estimation: a practical guide. Cooperative Research Report
     144, International Council for the Exploration of the Sea, 57 pp
  - Froese R, Pauly D (2016) FishBase. World Wide Web electronic publication. www.fishbase.org, version (10/2016)
  - Gascoigne J (2002) Nassau grouper and queen conch in the Bahamas:
     status and management options. Report to the Bahamas Reef
     Environment Educational Foundation. Nassau, The Bahamas
  - Gauthier S, Horne JK (2004) Potential acoustic discrimination within
     boreal fish assemblages. ICES J Mar Sci 61:836–845
  - Gibson J (2007) Managing a Nassau grouper fishery—a case study
     from Belize. Proceedings of the Gulf and Caribbean Fisheries
     Institute 60:1–2
  - Gledhill CT, Lyczkowski-Shultz J, Rademacher K, Kargard E, Crist
     G, Grace MA (1996) Evaluation of video and acoustic index
     methods for assessing reef-fish populations. ICES J Mar Sci
     53:483–485
- 621 Guillard J, Lebourges-Dhaussy A, Brehmer P (2004) Simultaneous Sv
   622 and TS measurements on young-of-the-year (YOY) freshwater
   623 fish using three frequencies. ICES J Mar Sci 61:267–273
- Heppell SA, Semmens BX, Archer SK, Pattengill-Semmens CV,
  Bush PG, McCoy CM, Heppell SS, Johnson BC (2012)
  Documenting recovery of a spawning aggregation through size
  frequency analysis from underwater laser calipers measurements. Biol Conserv 155:119–127
- Johannes RE (1978) Reproductive strategies of coastal marine fishesin the tropics. Environ Biol Fish 3:65–84
- Johannes RE, Squire L, Granam T, Sadovy Y, Renguul H (1999)
  Spawning aggregations of groupers (Serranidae) in Palau.
  Marine Conservation Research Series Publication 1, The Nature
  Conservancy, 144 pp
- Johnston SV, Rivera JA, Rosario A, Timko MA, Nealson PA, Kumagai KK (2006) Hydroacoustic evaluation of spawning red hind (*Epinephelus guttatus*) aggregations along the coast of Puerto Rico in 2002 and 2003. Emerging technologies for reef fisheries research and management, National Marine Fisheries Service Professional Paper 5. NOAA, Seattle, WA, pp 10–17
- Jones DT, Wilson CD, Robertis AD, Rooper CN, Weber TC, Butler
  JL (2012) Evaluation of rockfish abundance in untrawlable
  habitat: combining acoustic and complementary sampling tools.
  Fish Buletinl 110:332–343
- Kadison E, Nemeth RS, Blondeau J, Smith T, Calnan J (2010) Nassau grouper (*Epinephelus striatus*) in St. Thomas, US Virgin Islands, with evidence for a spawning aggregation site recovery.
  Proceedings of the Gulf and Caribbean Fisheries Institute 62:273–279
- Kellner JB, Tetreault I, Gaines SD, Nisbet RM (2007) Fishing the line
   near marine reserves in single and multispecies fisheries. Ecol
   Appl 17:1039–1054
- Knudsen FR, Hawkins AD, McAllen R, Sand O (2009) Diel
  interactions between sprat and mackerel in a marine lough and
  their effects upon acoustic measurements of fish abundance. Fish
  Res 100:140–147

- Kobara S, Heyman WD (2008) Geomorphometric patterns of Nassau<br/>grouper (*Epinephelus striatus*) spawning aggregation sites in the<br/>Cayman Islands. Marine Geodesy 31:231–245659Kracker L (2007) Hydroacoustic surveys: a non-destructive approach660
- Kracker L (2007) Hydroacoustic surveys: a non-destructive approach to monitoring fish distributions at National Marine Sanctuaries.
   Technical Memorandum 66, National Ocean Service, National Centers for Coastal Ocean Science, NOAA, Charlston, SC

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706

707

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710

711

712

713

714

715

716

- Lilja J, Marjomäki TJ, Jurvelius J, Rossi T, Heikkola E (2004) Simulation and experimental measurement of side-aspect target strength of Atlantic salmon (*Salmo salar*) at high frequency. Can J Fish Aquat Sci 61:2227–2236
- Løland A, Aldrin M, Ona E, Hjellvik V, Holst JC (2007) Estimating and decomposing total uncertainty for survey based abundance estimates of Norwegian spring spawning herring. ICES J Mar Sci 64:1302–1312
- Love RH (1971) Measurements of fish target strength: a review. Fishery Bulletin 69:703–715
- Machias A, Tsimenidis N (1995) Biological factors affecting the swimbladder volume of sardine (*Sardina pilchardus*). Mar Biol 23:859–867
- Murphy HM, Jenkins GP (2010) Observational methods used in marine spatial monitoring of fishes and associated habitats: a review. Mar Freshw Res 61:236–252
- Nemeth RS (2005) Population characteristics of a recovering US Virgin Islands red hind spawning aggregation following protection. Mar Ecol Prog Ser 286:81–97
- Nielsen JR, Lundgren B (1999) Hydroacoustic ex situ target strength measurements on juvenile cod (*Gadus morhua* L.). ICES J Mar Sci 56:627–639
- Ona E, Mitson RB (1996) Acoustic sampling and signal processing near the seabed: the deadzone revisited. ICES J Mar Sci 53:677–690
- Parker-Stetter SL, Rudstam LG, Sullivan PJ, Warner DM (2009) Standard operating procedures for fisheries acoustic surveys in the Great Lakes. Special Publication 09-01, Great Lakes Fisheries Commission, Ann Arbor, MI
- Reid DG (2000) Report on echo trace classification. Cooperative Research Report No. 238, International Council for the Exploration of the Sea, Copenhagen, Denmark, 107 pp
- Rivera J, Kellison T, Appeldoorn RS, Schärer M, Nemeth M, Rowell T, Mateos D, Nealson P (2010) Detection of Mona Island and Abrir La Sierra, Puerto Rico red hind (*Epinephelus guttatus*) 1 m off the bottom with hydroacoustic techniques. Proceedings of the Gulf and Caribbean Fisheries Institute 63:143–148
- Rose GA (2009) Variations in the target strength of Atlantic cod during vertical migration. ICES J Mar Sci 66:1205–1211
- Rudstam LG, Parker SL, Einhouse DW, Witzel LD, Warner DM, Stritzel JL, Parrish DL, Sullivan PJ (2003) Application of in situ target-strength estimations in lakes: examples from rainbowsmelt surveys in Lakes Erie and Champlain. ICES J Mar Sci 60:500–507
- Ryan TE, Kloser RJ, Macaulay GJ (2009) Measurement and visual verification of fish target strength using an acoustic-optical system attached to a trawlnet. ICES J Mar Sci 66:238–1244
- Sadovy de Mitcheson Y (2016) Mainstreaming fish spawning aggregations into fishery management calls for a precautionary approach. BioScience 66:295–306
- Sadovy Y, Domeier M (2005) Are aggregation fisheries sustainable? Reef fish fisheries as a case study. Coral Reefs 24:254–262
- Sadovy Y, Colin PL (eds) (2012) Reef fish spawning aggregations: biology, research and management. Springer, Berlin
- Sadovy de Mitcheson Y, Erisman B (2012) Fishery and biological implications of fishing spawning aggregations, and the social and economic importance of aggregating fishes. In: Sadovy Y, Colin PL (eds) Reef fish spawning aggregations: biology, research and management. Springer, Berlin, pp 225–284
  718
  718
  719
  720
  721
  722
  723

Sadovy de Mitcheson Y, Cornish A, Domeier M, Colin P, Russell M, 724 Lindeman K (2008) A global baseline for spawning aggregations 725 of reef fishes. Conserv Biol 22:1233-1244 726

723

727

740

741

- Sala E, Ballesteros E, Starr RM (2001) Rapid decline of Nassau grouper spawning aggregations in Belize: fishery management and conservation needs. Fisheries 26:23-30
- 728 729 Semmens BX, Bush P, Heppell S, Johnson B, McCoy C, Pattengill-730 Semmens C, Whaylen L (2007) Charting a course for Nassau 731 grouper recovery in the Caribbean: what we've learned and what 732 we still need to know. Proceedings of the Gulf and Caribbean 733 Fisheries Institute 60:607-609
- 734 Simmonds EJ, MacLennan DN (2005) Fisheries acoustics: theory and 735 practice, 2nd ed. Fish and Fisheries Series, Blackwell, Oxford, 736 UK
- 737 Smith CL (1972) A spawning aggregation of Nassau grouper, 738 739 Epinephelus striatus (Bloch). Trans Am Fish Soc 101:257-261
  - Soule M, Hampton I, Barange M (1995) Evidence of bias in estimates of target strength obtained with a split-beam echo-sounder. ICES J Mar Sci 52:139-144
- 742 Stallings CD (2008) Indirect effects of an exploited predator on 743 recruitment of coral reef fishes. Ecology 89:2090-2095
- 744 Stallings CD (2009) Predator identity and recruitment of coral-reef 745 fishes: indirect effects of fishing. Mar Ecol Prog Ser 746 383:251-259
- 747 Starr RM, Sala E, Ballesteros E, Zabala M (2007) Spatial dynamics of 748 the Nassau grouper Epinephelus striatus in a Caribbean atoll. 749 Mar Ecol Prog Ser 343:239-249
- 750 Starr RM, Fox DS, Hixon MA, Tissot BN, Johnson GE, Barss WH 751 (1995) Comparison of submersible-survey and hydroacoustic-752 survey estimates of fish density on a rocky bank. Fishery Bulletin 753 94:113-123
- 754 Taylor CJ, Eggleston DB, Rand PS (2006) Nassau grouper 755 (Epinephelus striatus) spawning aggregations: hydroacoustic 756 surveys and geostatistical analysis. Emerging technologies for

reef fisheries research and management, National Marine Fisheries Service Professional Paper 5. NOAA, Seattle, WA, pp 18-25

- Thomas GL, Thorne RE (2003) Acoustical-optical assessment of Pacific herring and their predator assemblage in Prince William Sound, Alaska. Aquat Living Resour 16:247-253
- Trenkel VM, Ressler PH, Jech M, Giannoulaki M, Taylor C (2011) Underwater acoustics for ecosystem-based management: state of the science and proposals for ecosystem indicators. Mar Ecol Prog Ser 442:285-301
- Utne KR, Ona E (2006) Acoustic extinction in dense herring layers, measured from a bottom-mounted transducer. Document CM/ 2006/I: 10, International Council for the Exploration of the Sea, Copenhagen, Denmark
- Whaylen L, Pattengill-Semmens CV, Semmens BX, Bush PG, Boardman MR (2004) Observations of a Nassau grouper (Epinephelus striatus) spawning aggregation site in Little Cayman, including multi-species spawning information. Environ Biol Fish 70:305-313
- Whaylen L, Bush PG, Johnson BC, Luke KE, McCoy CMR, Heppell S, Semmens BX, Boardman M (2006) Aggregation dynamics and lessons learned from five years of monitoring at a Nassau grouper (Epinephelus striatus) spawning aggregation in Little Cayman, Cayman Islands, BWI. Proceedings of the Gulf and Caribbean Fisheries Institute 57:1-14
- Winfield IJ, Emmrich M, Guillard J, Mehner T, Rustadbakken A (2011) Guidelines for standardisation of hydroacoustic methods. 784 Centre for Ecology and Hydrology Project Number C03630, 785 European Commission, 30 pp 786
- Zhao X, Ona E (2003) Estimation and compensation models for the shadowing effect in dense fish aggregations. ICES J Mar Sci 60:155-163

787

788