

Sympatric speciation in the genomic era

Foote, Andrew

Trends in Ecology and Evolution

DOI: 10.1016/j.tree.2017.11.003

Published: 01/02/2018

Cyswllt i'r cyhoeddiad / Link to publication

Dyfyniad o'r fersiwn a gyhoeddwyd / Citation for published version (APA): Foote, A. (2018). Sympatric speciation in the genomic era. *Trends in Ecology and Evolution*, *33*(2), 85-95. https://doi.org/10.1016/j.tree.2017.11.003

Hawliau Cyffredinol / General rights Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

· Users may download and print one copy of any publication from the public portal for the purpose of private study or research.

- You may not further distribute the material or use it for any profit-making activity or commercial gain
 You may freely distribute the URL identifying the publication in the public portal ?

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

±

Opinion Submission for Trends in Ecology and Evolution

1 Sympatric Speciation in the Genomic Era

2 Andrew D. Foote

3 Molecular Ecology and Fisheries Genetics Laboratory, School of Biological Sciences, Bangor University,

4 Bangor, Gwynedd, LL57 2UW, UK

5

6 Keywords: sympatric speciation; genomics; divergence-with-gene-flow

7

8 Sympatric speciation has been of key interest to biologists investigating how natural and 9 sexual selection drive speciation without the confounding variable of geographic isolation. 10 The advent of the genomic era has provided a more nuanced and quantitative 11 understanding of the different and often complex modes of speciation by which sympatric 12 sister taxa arose, and a re-assessment of some of the most compelling empirical case studies of sympatric speciation. However, I argue that genomic studies based on 13 contemporary populations may never be able to provide unequivocal evidence of true 14 15 primary sympatric speciation, and there is a need to incorporate palaeogenomic studies 16 in to this field. This inability to robustly distinguish cases of primary and secondary divergence-with-gene-flow may be inconsequential, as both are useful for understanding 17 18 the role of large effect barrier loci in the progression from localised genic isolation to 19 genome-wide reproductive isolation. I argue that they can be of equivalent interest due 20 to shared underlying mechanisms driving divergence and potentially leaving similar 21 coalescent patterns.

- 22
- 23

24 A Century of Contention Over Sympatric Speciation

25 Primary sympatric speciation is the evolution of reproductive isolation without geographic 26 barriers, in which new species arise from a single ancestral population [1-5]. As these criteria 27 do not allow for any physical separation between the incipient species, the potential for inter-28 breeding and gene flow remains throughout the speciation process, from inception to 29 completion. Recombination can therefore break up linkage between alleles beneficially 30 associated with environmental variation, and alleles associated with incompatibilities and 31 reproductive isolation [6]. As such, it is the most extreme, restrictive and arguably the most 32 controversial scenario of divergence-with-gene-flow [7-11]. Thus, the existence and relevance 33 of this mode of speciation in nature has been hotly debated for over a century [1-11]. The 34 continued great interest for evolutionary biologists in sympatric speciation is understanding the 35 seemingly rare conditions and processes under which natural and sexual selection can drive 36 ecological divergence and reproductive isolation in a continuously distributed population [4,7], 37 as compared with allopatric speciation, in which geographic barriers initiate reproductive 38 isolation and population divergence follows [2-8]. Under the latter scenario, it can be difficult 39 to establish the extent of the role of selection due to ecological variation relative to intrinsic 40 barriers developed during geographic isolation in promoting reproductive isolation [12].

41

42 After over a century of debate, and despite its theoretical plausibility and some apparently 43 compelling empirical examples, many facets of sympatric speciation remain controversial. 44 Given this, a recent review on speciation argued that the debate over allopatric versus sympatric 45 speciation was unproductive and should not be a significant part of the future research agenda 46 [13]. However, as per the oft-quoted prediction by Mayr: 'Sympatric speciation is like the 47 Lernaean hydra which grew two new heads whenever one of its old heads was cut off.....the 48 issue will be raised again at regular intervals' [8]. The advent of high-throughput sequencing, 49 coupled with the development and application of population genomic methods that allow the 50 inference of complex evolutionary histories, have led to a resurgent interest in sympatric 51 speciation and a re-assessment of some of the most compelling empirical case studies 52 [14,15,16].

53

In the genomic era, we can now quantify the genetic contribution of one or more ancestral populations to contemporaneously sampled sympatric daughter species. These advances have led to some of the most compelling examples of primary sympatric speciation being reconsidered as a product of multiple colonisations and secondary contact. Other examples

appear to be robust. However, here I argue that such backward-in-time approaches have limited 58 59 ability to distinguish between periods of spatial overlap, but the absence of gene flow (i.e., 60 when no coalescence events take place between the ancestral incipient species), and the absence of gene flow during periods of spatial separation. I propose that a forward-in-time 61 approach, utilising palaeogenomics may be a complementary approach that could leverage 62 63 additional information in some contexts. Lastly, I consider whether primary and secondary 64 sympatric speciation represent a mechanistic dichotomy. I suggest that primary and secondary 65 contact can leave a similar genomic signature, when speciation is driven by tightly clustered or large effect loci. Arguably, the advent of affordable population genomic studies should place 66 less focus on whether study systems result from primary or secondary contact and instead focus 67 Thereby, facilitating on the mechanistic aspects of the genomic architecture and making progress in identifying the 68 69 conditions and processes under which natural and sexual selection can drive speciation, without 70 extrinsic barriers to gene flow [13].

71

72 Genomic insights into the ancestral context of sympatric speciation

73 A compelling empirical case study of primary sympatric speciation requires the robust 74 inference of past biogeography; specifically, that the present day sympatric daughter species arose from a common ancestral population, with no period of geographic isolation (see Box 1). 75 76 Prior to the genomic era, empiricists used phylogenetics and assumed that the geographic 77 distribution of the ancestral population was the same as the present-day daughter species, if 78 they formed **monophyletic** species pairs or flocks in geographically isolated 'island' habitats 79 [17-24]. However, a major limitation of the inference of sympatric speciation from the monophyletic relationship among sympatric species is that monophyly may result from several 80 processes, other than true sympatric speciation (Figure 1). Modelling speciation as a bifurcating 81 82 tree presents a point estimate of this evolutionary process [14] and does not consider the 83 possibility that species derived ancestry from multiple source populations [25-27]. This is a 84 key flaw with the criteria of Covne and Orr [4]: monophyly of sympatric sister species is 85 consistent with, but not exclusive to a scenario of sympatric speciation. It does not provide conclusive evidence that present day sympatric sister species emerged from a single 86 colonisation, nor does it reject the alternative scenario of multiple colonisations in which 87 88 monophyly results from introgression upon secondary contact [7,28].

90 However, these different scenarios do typically generate different patterns of genome-wide 91 ancestry that can be used to distinguish between them. Under a scenario of sympatric speciation 92 from a single source population, the daughter species will share a common ancestry, with 93 segregating alleles being mainly those that are recently derived or were at low frequency in the 94 ancestral population [14,15,16]. Alternatively, if sympatric sister species are the result of 95 multiple colonisations and gene flow upon secondary contact, then each species should share differing proportions of ancestry with source outgroups (Figure 1). We can consider this as a 96 97 continuum, from a single **panmictic** colonising population (Figure 1A); to colonisation by a 98 hybrid swarm (Figure 1B); and lastly multiple colonisations and secondary contact following 99 periods of geographic isolation (Figure 1C). This is a representative, but not an exhaustive list 100 of possible scenarios that could generate the same consensus phylogentic pattern as sympatric 101 speciation. Recently developed genomics methods can provide robust evidence of admixture 102 and estimate ancestry proportions, even if gene flow events occurred hundreds of generations 103 ago and under scenarios of incomplete lineage sorting and demographic change [29-32]. For 104 example, the closely related *D*-statistic (ABBA-BABA) and *f*-statistic tests identify taxa that 105 share an excess of ancestry (measured as derived alleles and allele frequencies respectively) 106 with an outgroup [29,30]. The tract length of genomic regions inferred to have introgressed 107 during secondary contact can provide further information on the timing of gene flow, and 108 whether introgression pre- or post-dated sympatric diversification [33,34].

109

110 The application of such a population genomics approach has reassessed the sympatric origins of arguably some of the most compelling empirical examples of sympatric speciation: 111 monophyletic species pairs and flocks of cichlids found in small uniform crater lakes in 112 113 Cameroon, Nicaragua and Tanzania [14,15,16]. The lakes were argued to be sufficiently small-114 in-size; ecologically monotonous with no microgeographical barriers; and isolated from outside riverine populations by the crater rim, that sympatric speciation appeared to be the most 115 116 likely biogeographical scenario under which these sister species had diverged [17,18]. In each 117 case, cichlid species within the lakes have diverged in ecologically-associated morphological traits, and show evidence of reproductive isolation and monophyly, consistent with sympatric 118 119 origins [15,17-21,35]. However, analyses of genome-wide ancestry have revealed varying 120 complexity in the evolutionary history of cichlids within each study area. These range from 121 genomic ancestries that are best explained by multiple colonisations of Cameroon crater lakes 122 and secondary gene flow following periods of allopatry [14]; to divergence in sympatry in 123 Nicaraguan crater lakes, but following secondary colonisation events and admixture prior to

the radiations within each lake [16]; to what appears to be speciation following a single colonisation in a Tanzanian crater lake, albeit with some gene flow from the lake to nearby outgroup populations [15].

127

128 These descriptive results can then be developed into demographic models, allowing the 129 estimation of ancestral divergence times, effective population sizes and migration rates, and 130 the testing of alternative evolutionary scenarios (e.g. [15,36,37]). However, modelling whether 131 sympatric populations diverged with gene flow, or whether migration took place sometime 132 after the populations had diverged, consistent with secondary contact, requires the estimation 133 of the timing and the number of migration events [38-40]. These parameters can be intractable, 134 as genomic data from present day populations can be consistent with many migration and admixture scenarios, which result in the same coalescent times [39,40]. More general caveats 135 136 also apply, for example, most models are oversimplified representations of biological reality, 137 and only inputted models are tested. Model-based approaches are therefore best accompanied 138 with model-free methods to identify a range of estimates for parameters, and scenarios to test. 139 Additionally, there is a need to exclude non-neutral loci and account for genome-wide variation 140 in effective migration and recombination [37,41].

141

142 The biological realism and relevance of the classification of the mode of speciation into the discrete geographic categories such as sympatric, parapatric and allopatric has been 143 144 questioned. Almost all candidate case studies of sympatric speciation have some degree of 145 spatio-temporal differentiation between sister taxa, for example due to the patchy distribution of preferred habitat [14,42-44]. To countenance this, some have suggested that the relationship 146 between taxa during the speciation process may be better quantified in a population genetics 147 148 framework that quantifies key parameters such as migration rate [42]. This approach, and 149 modelling sympatric speciation in general, relies on assuming a starting point of panmixia in 150 the ancestral population [5]. Yet this assumption of ancestral panmixia has been difficult or 151 impossible to prove or reject in empirical case studies prior to the genomic era [42]. Others 152 have argued for retaining a spatial component of sympatric speciation, in accordance with 153 Mayr's definition [8]: that speciating sister taxa should be in '*cruising range*' of each other 154 throughout the speciation process [44]. However, in each case, the geographic context of 155 speciation is divided into artificially discrete categories, whether they be based on spatial or 156 genetic measures of separation [11]. Instead, the geographic context of speciation is perhaps 157 best viewed as a graded continuum [10,11]. The genomic approaches outlined above estimate

158 the contribution of the shared ancestral population and any other contributing outgroup 159 populations to the ancestry of the daughter species. Thereby providing a continuous and 160 quantitative measure of the context and mode of speciation. This still does not fully resolve the uncertainty in the geographic context of divergence. For example, even among sympatric taxa 161 162 with no detectable contribution from ancestral outgroups, as in Figure 1A, there may have been 163 periods of spatial segregation among currently sympatric sister taxa. Ultimately, our ability to 164 reconstruct the evolutionary history of sympatric sister taxa back to the shared ancestral 165 population using backward-in-time genomic approaches, is constrained to being able to 166 identify periods of gene flow through **coalescent** events, but is not able to distinguish periods of spatial overlap without gene flow from periods of spatial isolation. 167

168

169 Due to the timescales over which evolutionary processes such as adaptation and speciation take 170 place, forward-in-time approaches are rarely utilised due to the limitations on the number of 171 generations that can be sampled. However, the advent of palaeogenomics is expanding the 172 scope of timescales over which we can sample genomes and look at genetic change from an ancestral population going forward in time to daughter species, and can complement 173 174 hindcasting from contemporaneously sampled genomes. For example, sediment cores from post-glacial lakes can be used to sample lineages from the time the glaciers retreated to the 175 176 present day (Figure 2). Such an approach has recently been applied to extract DNA from 177 sediment of two lakes in Sweden, spanning the past 10,000 years, to reconstruct the colonisation and connectivity between whitefish (Coregonus lavaretus) ecotypes [45]. Whilst 178 179 only very low concentrations of DNA are found in sediments, the sequencing of hard parts 180 within the different layers of the sediment core, for example bones or spines, can yield genomic 181 sequences that allow the tracking of genomic changes at QTL forwards in time.

182

183 The genomic architecture of sympatric speciation

The genomic architecture of a trait can be summarised as the number of underlying loci, their effect size and additivity, and their physical spacing across the genome. In addition to being shaped by recent and ongoing selection, this genomic architecture can be influenced by processes that include demographic history, linked selection in the ancestral population, recent and ongoing selection, and recombination rate [46].

189

190 Key questions in the study of sympatric speciation are how a genomic architecture shaped by 191 gradual, incremental changes that occur under natural selection can account for rapid bursts of

192 adaptive divergence; how localised genomic changes result in genome-wide reproductive 193 isolation; and how they can overcome the homogenising effect of ongoing gene flow [47-49]. 194 Over the past decade genomic studies of adaptation have progressed from investigating single 195 or a few candidate genes to genome-wide studies, and have highlighted how divergence linked 196 to adaption can be widespread across the genome. Yet the chronology of genic change during 197 speciation, and how this progresses from individual 'barrier loci', through to genome-wide 198 differentiation (and how to study these processes), is still contentious and widely debated (see 199 reference [49] and associated commentaries).

200

201 One of the primary approaches to exploring these questions has been to compare genome-wide 202 variation in differentiation (F_{ST}) of allele frequencies across the 'speciation continuum'; *i.e.*, 203 between multiple pairs of sympatric and allopatric sister taxa that are at different stages of 204 divergence [47,48]. This approach has been applied to multiple taxa, with varied results. While, 205 most such studies to date have shown a progressive increase in the build-up of mean genome-206 wide differentiation across the speciation continuum [50-53], and some have highlighted 207 important barrier loci that reduce localised effective migration within some genomic regions 208 due to being associated with adaptation and/or reproductive isolation [54,55]; many of these 209 studies have identified alternative underlying causes of heterogeneity in the landscape of 210 genomic differentiation [50-52]. These include reduced diversity from linked selection in the 211 ancestral population, for example due to background selection (BGS) removing deleterious 212 variants [56]; BGS is in-turn associated with variation in recombination rate and gene density 213 in regions such as centromeres [57,58]; and selection on genome-wide smaller effect loci 214 underlying polygenic traits. The genomic background of these different processes can then 215 mask any potential signal from barrier loci associated with adaptation or reproductive isolation. 216 However, young examples of sympatric speciation may generate rare exemplar study systems, in which there are clear 'genomic islands' which contain barrier loci associated with 217 218 reproductive isolation and ecological diversification.

219

The effect size of a locus on a phenotypic trait has a positive correlative relationship with pleiotropy and deleterious effects [59], therefore adaptation is predicted to typically progress due to small changes in frequency across many alleles, each with a small additive phenotypic effect [60]. However, as noted above, in scenarios of ongoing gene flow during sympatry, recombination would be expected to break up linkage between loci associated with ecological adaptation and those associated with mate preference, thus counteracting ecologically driven

speciation [6-9]. Additionally, the strength of selection on a locus is not just a function of its 226 227 effect size and its interaction with the environment, it is also a function of effective population 228 size (Ne). The more robust examples of primary sympatric speciation are typically those that 229 have colonised a remote, or closed, ecosystem prior to diverging, e.g. Lord Howe Island flora [23,24] and crater lake cichlids [17-19]. Thus, it seems realistic that only a small number of 230 231 initial colonisers founded these island or closed ecosystems. This founder effect is expected to 232 greatly lower selection coefficients at loci of small effect that act additively on traits. Therefore, 233 traits associated with ecological variation or mate choice that diverge during sympatric 234 speciation, are more likely to be determined by loci tightly linked to each other in genomic 235 regions of low recombination such as inversions [46,61], or be synergistically pleiotropic, *i.e.* 236 so-called 'magic traits', which have a role in both ecological adaptation and assortative mating 237 [62]. Therefore, these study systems are those that we expect barrier loci of large effect to be 238 differentiated against a homogenous genome-wide background.

239

240 Recent genomic studies investigating quantitative trait loci (QTL) in model systems for 241 speciation-with-gene-flow, have largely validated these predictions. For example, in Midas 242 cichlids in Nicaraguan crater lakes, the highest effect size QTL for body shape and pharyngeal jaw morphology, both traits which show ecological-associated variation [20,21] are tightly 243 244 clustered on a single chromosome and allele frequencies at these loci segregate in sympatric sister species [63]. Comparison of the genomes of benthic and littoral ecomorphs of 245 Astatotilapia cichlids from a Tanzanian crater lake found regions of high differentiation and 246 high divergence clustered mainly in five linkage groups harbouring genes associated with 247 248 morphology and optical sensitivity, and therefore ecological variation and mate choice [15]. A recent study on sympatric populations of monkey flower species Minulus laciatus and M. 249 250 guttatus found that a few large effect size QTL explained much of the variance in flowering time and flower size traits [64]. Differences in flowering time are thought to be locally 251 252 adaptive: M. laciatus, is found on dry exposed rocky outcrops and flowers earlier than M. 253 guttatus to avoid the seasonal drought; and act as a prezygotic barrier to gene flow, therefore 254 qualifying as a 'magic trait' [64]. Allochrony also plays a role in reproductive isolation between 255 sympatric hawthorn and apple-infesting host races of the *Rhagoletis pomonella* fly, which 256 differ in the intensity and timing of diapause [65]. SNP loci associated with the timing of 257 diapause onset and diapause intensity were in several tightly linked clusters, thought to be 258 within inversions [66].

The findings of these empirical studies are highly concordant with the predictions of most 260 261 theoretical models of sympatric speciation, which require linkage between loci associated with 262 reproductive isolation and loci associated with ecological adaptation, or pleiotropy in which ecological adaptation and reproductive isolation evolve simultaneously [67-69]. This contrasts 263 264 with empirical examples in which a period of allopatry was important in segregating alleles 265 associated with ecological variation. In examples of the latter scenario, intrinsic barriers can 266 build up in many widespread genomic regions without recombination breaking them up during 267 this allopatric phase. Thus, in many examples of sympatric speciation we anticipate large 268 changes in allele frequencies at single or a few loci, while the rest of the genome is 269 homogenised, until complete genome-wide isolation is established. Therefore, the coalescent 270 times of the barrier loci are expected to pre-date the genome-wide time-to-most-recent-271 common-ancestor (TMRCA) [70] (Figure 3). In contrast, if genome-wide polygenic adaptation 272 and reproductive incompatibilities have evolved in allopatry, prior to secondary contact, then 273 the TMRCA of the loci associated with reproductive isolation will be within the genome-wide 274 range and need not be associated with large changes in allele frequencies, making them cryptic 275 to genome-wide scan methods.

276

277 Strict primary divergence-with-gene-flow may not be needed for studying the evolution of 278 large effect barrier loci against a homogenous genomic background. In theory, this pattern, 279 could also be expected even if the genetic underpinning of divergent ecological adaptation and 280 reproductive isolation develops during allopatry, and then segregates again after an initial 281 period of mixing upon secondary contact, provided there is genome-wide homogenisation upon 282 secondary contact (Figure 3). An allopatric phase and/or introgression events can facilitate 283 speciation by intensifying disruptive selection and introducing new genomic variation that can 284 act as a substrate for segregating polymorphisms under natural and sexual selection. Guerrero & Hahn [71] recently suggested that balanced polymorphisms in the ancestral population, 285 could sort upon splitting into daughter species, either due to ecological variation selecting for 286 287 alternate alleles, or through selectively neutral sorting. They highlighted that such a process could explain the high absolute genetic divergence (D_{XY}) , suggestive of an ancient divergence, 288 289 in the few genomic islands found when comparing the littoral and benthic ecomorphs of the 290 Tanzanian crater lake Massoko. The two ecomorphs are estimated to have diverged only 500-291 1,000 years, having diverged from the putative source population 10,000 years ago in a crater 292 lake that formed ~50,000 years ago [15]. Guerrero & Hahn [71] highlight that these regions 293 containing putative balanced polymorphisms would form 'genomic islands' even without

background F_{ST} and D_{XY} being lowered due to genome-wide homogenisation from gene flow. 294 295 However, it is not hard to imagine that these two forms could have arisen and collapsed 296 multiple times since colonising the crater, for example, due to episodic changes in water depth. 297 If negative frequency dependent selection maintained ecologically adaptive polymorphisms 298 even when the two forms collapse into an otherwise homogenous population, such a process 299 of repeated collapse and vicariance could mask any genomic signature of divergent origins in 300 the present-day populations, with the exception of balanced polymorphisms, which would 301 coalesce much further back in time than the genome-wide mean TMRCA (Figure 3).

302

303 Lineage sorting and high genomic differentiation are also found at loci of large effect in the 304 partially sympatric benthic-limnetic species pairs of threespine sticklebacks found in several 305 lakes in British Columbia, Canada and hypothesised to have originated from a secondary 306 invasion [72]. A PCA analysis of genome-wide neutrally evolving SNPs found a pattern of 307 clustering by lake [73], which would be consistent with independent divergence of the benthic 308 and limnetic forms of stickleback within each lake. However, SNPs evolving under natural 309 selection grouped individuals by ecological niche, with further clustering of the older benthic 310 form with geographically proximate single-form freshwater populations, whilst the younger 311 limnetic form clustered more closely with marine populations [73]. These results are consistent 312 with re-use of standing genetic variation from a second marine-to-freshwater colonisation, 313 which then provided the raw genetic material for divergence within each lake driven by 314 disruptive selection. Thus, the adaptation and speciation loci coalesce much further back in 315 time, than the mean TMRCA of unlinked neutral loci. A further example is the sympatric 316 hawthorn and apple-infesting races of *Rhagoletis pomonella* fruit fly, in which the inversion 317 polymorphism influencing diapause traits evolved during an allopatric phase greater than a 318 million years ago [74].

- 319
- 320

321 Concluding remarks

In the genomic era, sympatric speciation continues to be a controversial and much-debated phenomenon. The exemplar study systems, such as crater lake cichlids of Cameroon, which had convinced even the most hardened sceptics [4], have been called into question. Genome sequences provide the unprecedented means to reconstruct the ancestry of contemporary

instances

populations; for example, identifying where sympatric sister taxa that were thought to represent 326 originating from a common ancestral population a monophyletic group, are instead derived from multiple ancestral source populations [14]. 327 However, there remains a bias towards being able to disprove primary sympatric speciation, 328 329 whilst generating conclusive evidence in support of primary sympatric speciation based on 330 hindcasting using modern genomes remain elusive. I suggest that palaeogenomics may have a complementary role to play in future studies; for example, the sequencing of DNA from 331 332 sediment cores can identify the temporal patterns of spatial overlap between two speciating 333 lineages, even in the absence of gene flow. Lastly, the great interest of biologists in sympatric UNDERSTANDING speciation has been how two lineages can diverge and become reproductively isolated in the 334 335 absence of extrinsic barriers. In the genomic era, we can study this process at the genic level. 336 In this review, I have highlighted several characteristics of the genomic underpinning of sympatric speciation, and that these can be found in examples of primary and secondary 337 338 sympatric speciation. I therefore contend that it is the investigation of the process of sympatric 339 speciation, rather than a dogmatic search for true primary sympatric speciation that will be 340 most valuable to our understanding of speciation and adaptation at the genomic level.

341

342 Acknowledgements

I would like to thank the editor, Paul Craze, and Jeff Feder and one anonymous reviewers for their constructive feedback, and Alex Papadopulos for useful discussions on this topic which greatly improved this manuscript. Financial support was provided by the Welsh Government and Higher Education Funding Council for Wales through the Sêr Cymru National Research Network for Low Carbon, Energy and Environment, and from the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement No 663830.

| 351 | Re | ferences |
|-----|-----|--|
| 352 | 1. | Poulton, E.B. (1904) What is a species? Trans. Entomol. Soc. Lond. 1903, 77-116 |
| 353 | 2. | Jordan, D.S. (1905) The origin of species through isolation. Science 22, 545-62 |
| 354 | 3. | Mayr, E. (1942). Systematics and Origin of Species, Columbia University Press |
| 355 | 4. | Coyne, J. and Orr, H. (2004) Speciation, Sinauer Associates |
| 356 | 5. | Gavrilets, S. (2003) Models of speciation: what have we learned in 40 years? |
| 357 | | Evolution 57, 2197–2215 |
| 358 | 6. | Slatkin, M. (1987) Gene flow and the geographic structure of natural populations. |
| 359 | | Science 236, 787–792 |
| 360 | 7. | Bolnick, D.I. and Fitzpatrick, B.M. (2007) Sympatric speciation: models and |
| 361 | | empirical evidence. Annu. Rev. Ecol. Evol. Syst. 38, 459-487 |
| 362 | 8. | Mayr, E. (1963) Animal Species and Evolution, Belknap |
| 363 | 9. | Felsenstein, J. (1981) Skepticism towards Santa Rosalia, or why are there so few |
| 364 | | kinds of animals? Evolution 35, 124–138 |
| 365 | 10. | Jiggins C.D. (2006) Sympatric Speciation: Why the Controversy? Curr. Biol. 16, |
| 366 | | R333–R334 |
| 367 | 11. | Butlin, R.K. et al. (2008) Sympatric, parapatric or allopatric: the most important way |
| 368 | | to classify speciation? Philos. Trans. R. Soc. Lond. B Biol. Sci. 363, 2997-3007 |
| 369 | 12. | Bierne, N. et al. (2011) The coupling hypothesis: why genome scans may fail to map |
| 370 | | local adaptation genes. Mol. Ecol. 20, 2044–2072 |
| 371 | 13. | Marie Curie Speciation Network (2012) What do we need to know about speciation? |
| 372 | | Trends Ecol. Evol. 27, 27–39 |
| 373 | 14. | Martin, C.H. et al. (2015). Complex histories of repeated gene flow in Cameroon |
| 374 | | crater lake cichlids cast doubt on one of the clearest examples of sympatric speciation. |
| 375 | | Evolution 69, 1406–1422 |
| 376 | 15. | Malinsky, M. et al. (2015) Genomic islands of speciation separate cichlid ecomorphs |
| 377 | | in an East African crater lake. Science 350, 1493–1498 |
| 378 | 16 | Kautt, A.F. et al. (2016) Multispecies outcomes of sympatric speciation after |
| 379 | | admixture with the source population in two radiations of Nicaraguan crater lake |
| 380 | | cichlids. PLoS Genet. 12: e1006157 |
| 381 | 17. | Schliewen, U.K. et al. (1994) Sympatric speciation suggested by monophyly of crater |
| 382 | | lake cichlids. Nature 368, 629-632 |

| 383 | 18. | Schliewen, U.K. and Klee, B. (2004) Reticulate sympatric speciation in Cameroonian |
|-----|-----|---|
| 384 | | crater lake cichlids. Frontiers Zool. 1:5 |
| 385 | 19. | Barluenga M. et al. (2006) Sympatric speciation in Nicaraguan crater lake cichlid |
| 386 | | fish. Nature 439, 719–723 |
| 387 | 20. | Elmer, K.R. et al. (2010) Rapid sympatric ecological differentiation of crater lake |
| 388 | | cichlid fishes within historic times. BMC Biology 8:60 |
| 389 | 21. | Elmer, K.R. et al. (2010) Local variation and parallel evolution: morphological and |
| 390 | | genetic diversity across a species complex of neotropical crater lake cichlid fishes. |
| 391 | | Phil. Trans. R. Soc. B 365, 1763–1782 |
| 392 | 22. | Filchak, K.E. et al. (2000) Natural selection and sympatric divergence in the apple |
| 393 | | maggot Rhagoletis pomonella. Nature 407, 739–742 |
| 394 | 23. | Savolainen, V. et al. (2006) Sympatric speciation in palms on an oceanic island. |
| 395 | | Nature 441, 213–213 |
| 396 | 24. | Papadopulos, A.S.T. et al. (2011) Speciation with gene flow on Lord Howe Island. |
| 397 | | Proc. Natl. Acad. Sci. 108, 13188–13193 |
| 398 | 25. | Cavalli-Sforza, L.L. (1973) Analytic review: some current problems of human |
| 399 | | population genetics. Am. J. Hum. Genet. 25, 82-104 |
| 400 | 26. | Cavalli-Sforza, L.L. and Piazza, A. (1975) Analysis of evolution: evolutionary rates, |
| 401 | | independence and treeness. Theor. Popul. Biol. 8, 127-165 |
| 402 | 27. | Felsenstein, J. (1982) How can we infer geography and history from gene |
| 403 | | frequencies? J Theor. Biol. 96, 9-20 |
| 404 | 28. | Schliewen, U.K. et al. (2006) Evolutionary biology-Evidence for sympatric |
| 405 | | speciation? Nature 444, E12-E13 |
| 406 | 29. | Green, R.E. et al. (2010) A draft sequence of the Neandertal genome. Science 328, |
| 407 | | 710–722 |
| 408 | 30. | Durand, E.Y. et al. (2011) Testing for ancient admixture between closely related |
| 409 | | populations. Mol. Biol. Evol. 28, 2239–2252 |
| 410 | 31. | Patterson, N. et al. (2012) Ancient admixture in human history. Genetics 192, 1065- |
| 411 | | 1093 |
| 412 | 32. | Peter, B.M. (2016) Admixture, population structure and F-statistics. Genetics 202, |
| 413 | | 1485–1501 |
| 414 | 33. | Harris, K. and Nielsen, R. (2013) Inferring demographic history from a spectrum of |
| 415 | | shared haplotype lengths. PLoS Genet 9:e1003521 |

| 416 | 34. | Lawson, D.J. et al. (2012) Inference of population structure using dense haplotype |
|-----|-----|---|
| 417 | | data. PLoS Genetics 8, e1002453 |
| 418 | 35. | Schliewen, U.K. et al. (2001) Genetic and ecological divergence of a monophyletic |
| 419 | | cichlid species pair under fully sympatric conditions in Lake Ejagham, Cameroon. |
| 420 | | Mol. Ecol. 10, 1471-1488 |
| 421 | 36. | Meier, J.I. et al. (2016) Demographic modelling with whole-genome data reveals |
| 422 | | parallel origin of similar Pundamilia cichlid species after hybridization. Mol. Ecol. |
| 423 | | doi: 10.1111/mec.13838 |
| 424 | 37. | Rougeaux, C. et al. (2017) Modeling the multiple facets of speciation-with-gene-flow |
| 425 | | towards inferring the divergence history of Lake Whitefish species pairs (Coregonus |
| 426 | | clupeaformis). Genome Biol. Evol. https://doi.org/10.1093/gbe/evx150 |
| 427 | 38. | Strasburg, J. and Rieseberg, L. (2011) Interpreting the estimated timing of migration |
| 428 | | events between hybridizing species. Mol. Ecol. 20, 2353-2366 |
| 429 | 39. | Sousa, V. C. et al. (2011) On the nonidentifiability of migration time estimates in |
| 430 | | isolation with migration models. Mol. Ecol. 20, 3956-3962 |
| 431 | 40. | Juric, I. et al. (2016) The strength of selection against Neanderthal Introgression. |
| 432 | | PLoS Genetics 12: e1006340. |
| 433 | 41. | Sousa, V. C. et al. (2013) Identifying loci under selection against gene flow in |
| 434 | | isolation-with-migration models. Genetics 194, 211-233 |
| 435 | 42. | Fitzpatrick, B.M. et al. (2008) What, if anything, is sympatric speciation? J. Evol. |
| 436 | | <i>Biol.</i> 21, 1452–1459 |
| 437 | 43. | Babik, W. et al. (2009) How sympatric is speciation in the Howea palms of Lord |
| 438 | | Howe Island? Mol. Ecol. 18, 3629–3638 |
| 439 | 44. | Mallet, J. et al. (2009) Space, sympatry and speciation. J. Evol. Biol. 22, 2332-2341 |
| 440 | 45. | Olajos, F. et al. (2017) Estimating species colonization dates using DNA in lake |
| 441 | | sediment. Methods Ecol. Evol. DOI: 10.1111/2041-210X.12890 |
| 442 | 46. | Lynch, M. and Walsh, B. (2007) The origins of genome architecture, Sinauer |
| 443 | | Associates |
| 444 | 47. | Feder, J.L. et al. (2012) The genomics of speciation-with-gene-flow. Trends in |
| 445 | | <i>Genetics</i> 28, 342–350 |
| 446 | 48. | Seehausen, O. et al. (2014) Genomics and the origin of species. Nat. Rev. Genet. 15, |
| 447 | | 176–192 |
| 448 | 49. | Ravinet, M. et al. (2017) Interpreting the genomic landscape of speciation: a road |
| 449 | | map for finding barriers to gene flow. J. Evol. Biol. 30, 1450-1477 |

| 450 | 50. | Renaut, S. et al. (2013) Genomic islands of divergence are not affected by geography |
|-----|-----|--|
| 451 | | of speciation in sunflowers. Nat. Comm. 4, 1827 |
| 452 | 51. | Martin, S.H. et al. (2013) Genome-wide evidence for speciation with gene flow in |
| 453 | | Heliconius butterflies. Genome Res. 23, 1817–1828 |
| 454 | 52. | Foote, A.D. et al. (2016). Genome-culture coevolution promotes rapid divergence of |
| 455 | | killer whale ecotypes. Nature Comm. 7, 11693 |
| 456 | 53. | Vijay, N. et al. (2016) Evolution of heterogeneous genome differentiation across |
| 457 | | multiple contact zones in a crow species complex. Nat. Comm. 7, 13195 |
| 458 | 54. | Nadeau, N.J. et al. (2012) Genomic islands of divergence in hybridizing Heliconius |
| 459 | | butterflies identified by large-scale targeted sequencing. Phil. Trans. R. Soc. B. 367 |
| 460 | | 343–353 |
| 461 | 55. | Jones, F.C. et al. (2012) The genomic basis of adaptive evolution in threespine |
| 462 | | sticklebacks. Nature 484, 55–61 |
| 463 | 56. | Cruickshank, T.E. and Hahn, M.W. (2014) Reanalysis suggests that genomic islands |
| 464 | | of speciation are due to reduced diversity, not reduced gene flow. Mol. Ecol. 23, |
| 465 | | 3133–3157 |
| 466 | 57. | Ellegren, H. et al. (2012) The genomic landscape of species divergence in Ficedula |
| 467 | | flycatchers. Nature 491, 756–760 |
| 468 | 58. | Burri, R. et al. (2015) Linked selection and recombination rate variation drive the |
| 469 | | evolution of the genomic landscape of differentiation across the speciation continuum |
| 470 | | of Ficedula flycatchers. Genome Res. 25, 1656–1665 |
| 471 | 59. | Wagner, G.P. et al. (2008) Pleiotropic scaling of gene effects and the 'cost of |
| 472 | | complexity'. Nature 452, 470-472 |
| 473 | 60. | Fisher, R.A. (1930) The Genetical Theory of Natural Selection. Oxford University |
| 474 | | Press |
| 475 | 61. | Yeaman, S. and Whitlock, M.C. (2011) The genetic architecture of adaptation under |
| 476 | | migration-selection balance. Evolution 65, 1897-1911 |
| 477 | 62. | Servedio, M.R. et al. (2011) Magic traits in speciation: 'magic' but not rare? Trends |
| 478 | | <i>Ecol. Evol.</i> 26, 389–397 |
| 479 | 63. | Fruciano, C. et al. (2016) Genetic linkage of distinct adaptive traits in sympatrically |
| 480 | | speciating crater lake cichlid fish. Nature Comm. 7:12736 |
| 481 | 64. | Ferris, K.G. et al. (2017) The genetic architecture of local adaptation and reproductive |
| 482 | | isolation in sympatry within the Minulus guttatus species complex. Mol. Ecol. 26, |
| 483 | | 208–224 |

| 484 | 65. Bush, G.L. (1969) Sympatric host race formation and speciation in frugivorous flies |
|------------|--|
| 485 | of the genus Rhagoletis (Diptera Tephritidae). Evolution 23, 237-251 |
| 486 | 66. Ragland, G.J. et al. (2017) A test of genomic modularity among life-history |
| 487 | adaptations promoting speciation with gene flow. Mol. Ecol. 26, 3926-3942 |
| 488 | 67. Dieckmann, U. and Doebeli, M. (1999) On the origin of species by sympatric |
| 489 | speciation. Nature 400, 354-357 |
| 490 | 68. Fry, J.D. (2003) Multilocus models of sympatric speciation: Bush vs Rice vs |
| 491 | Felsenstein. Evolution 57, 1735–1746 |
| 492 493 | 69. Gavrilets, S. (2004) Fitness Landscapes and the Origin of Species, Princeton |
| 493 494 | 70 Vang M <i>et al.</i> (2017) Can genomic data alone tell us whether speciation happened |
| 495 | with gene flow? <i>Mol. Ecol.</i> 26, 2845–2849 |
| 496 | 71. Guerrero, R.F. and Hahn, M.W. (2017) Speciation as a sieve for ancestral |
| 497 | polymorphism. Mol. Ecol. doi: 10.1111/mec.14290 |
| 498 | 72. Taylor, E.B. and McPhail, J.D. (2000) Historical contingency and ecological |
| 499 | determinism interact to prime speciation in sticklebacks, Gasterosteus. Proc. R. Sci. H |
| 500 | 267, 2375–2384 |
| 501 | 73. Jones, F.C. et al. (2012) A genome-wide SNP genotyping array reveals patterns of |
| 502 | global and repeated species-pair divergence in sticklebacks. Curr. Biol. 83-90 |
| 503 | 74. Feder, J.L. et al. (2003) Allopatric genetic origins for sympatric host-plant shifts and |
| 504 | race formation in Rhagoletis. Proc. Natl. Acad. Sci. 100, 10314–10319 |
| 505 | |
| 506 | |
| 507 | |
| 508 | |
| 509 | |
| 510 | |
| 511 | |
| 512 | |
| 513 | |
| 514 | Glossary |

515 Barrier loci: genetic loci that cause reduced gene flow between speciating taxa at a localised 516 region of the genome.

517 **Coalescent:** when two lineages sampled from different populations merge back in time in a 518 commonly shared ancestral lineage.

519 **Disruptive selection:** selection that favours extreme phenotypes over intermediate

520 phenotypes within a population.

521 Divergence-with-gene-flow: the build-up of genetic and phenotypic differences, despite on-

522 going exchange of genes. This differentiation is typically driven by disruptive natural

523 selection. The term has been used inclusive of scenarios of divergence under ongoing gene

flow upon secondary contact, and is thus does not exclusively refer to sympatric speciation.

525 **Ecomorph:** a population which has distinctive ecological and morphological features.

526 Genomic islands: a region of the genome that is highly differentiated (estimated using F_{ST})

527 between taxa compared with the genome-wide mean level of differentiation.

528 Magic trait: a trait subject to divergent selection and a trait contributing to mate choice

529 which are pleiotropic expressions of the same gene(s).

530 **Monophyletic:** belonging to a clade containing all the descendants of a single ancestor.

531 **Panmixia:** random mating within a population.

532 **Parapatric speciation:** the evolution of reproductive isolation in the absence of geographical

533 barriers to gene flow, in which the diverging populations have adjacent ranges.

534 **Pleiotropic:** an allele that has an effect on more than one trait.

535 **Polymorphisms:** genetic loci that have more than one allele.

536 **Quantitative trait loci:** genetic markers that are correlated with phenotype. These markers

537 contain, or are linked to, genes and regulatory regions associated with quantitative

538 phenotypic variation.

- **Recombination:** the process by which genomic regions are exchanged and broken up,
- 540 producing new combinations of alleles at different loci. Recombination occurs during meiosis
- 541 in eukaryotic cells.

Box 1. Pre-Genomic Era Criteria for Identifying Sympatric Speciation

In their classic review of speciation, Coyne & Orr [4] proposed four criteria that would need to be met in order for compelling case studies of sympatric speciation to be established. Given the restrictive conditions under which sympatric speciation is theoretically possible, these criteria for assessing empirical examples are equally stringent. Following, the argument of Mayr [8], they place the burden of proof on sympatric speciation and assume allopatric speciation as the null hypothesis. The four criteria can arguably be split into two components, one specifying the biogeographic conditions, and the other component specifying the genetic criteria under which an empirical case study would make a compelling example of sympatric speciation (Figure I).

Biogeographic Component

- 1. Species must have largely or completely over-lapping geographic range (Figure IA).
- 2. The biogeographic and evolutionary history of the groups must make the existence of an allopatric phase *very unlikely* (Figure IB).

Genetic Component

- 3. Speciation must show substantial reproductive isolation (Figure IC).
- 4. Sympatric species must be endemic sister species or an endemic monophyletic species flock (Figure ID).

As with most aspects of the study of sympatric speciation, these criteria have been a point of contention. See Bolnick and Fitzpatrick [7] for an in-depth discussion and review of these conditions.



Figure I. Biogeographic and Genetic Criteria for Sympatric Speciation. Empirical case studies on crater lake cichlids were among the first to be considered as compelling examples of primary sympatric speciation [16-18]. (A) Cichlid species in these studies had distributions that overlapped and different species were in 'cruising range' *sensu* Mayr [7]. (8) The high rim of the caldera of these craters isolates the lake from neighbouring rivers, and the conical shape of the lake bottom prevents separate basins forming during periods of low water-level [16]. Thus, there are no geographical barriers to gene flow within the crater lake. (C) Analyses of nuclear DNA markers suggest that gene flow occurs predominantly within rather than between species (illustrated here with an admixture plot) [18]. (D) Phylogenetic analyses show that cichlid species within each lake form a monophyletic clade with respect to outgroups from neighbouring river systems, suggesting that they radiated *in situ* from a single shared ancestral population [16-18].

545

547 Figure 1 Evolutionary histories that could result in a monophyletic relationship among 548 sympatric sister species. Schematic tree figures (top) are coloured to indicate changes in 549 allele frequencies during divergence and introgression (indicated by horizontal arrows). 550 Schematic ancestry palettes (bottom) are coloured to indicate the differences in ancestry proportions shared between the sympatric sister species and outgroups under each scenario. 551 552 (A) Speciation follows a single colonisation of an isolated 'island' habitat and divergence during sympatry. Under this scenario, the three sympatric sister species would share a similar 553 554 proportion of their ancestry with outgroups. (B) Colonisation of an isolated 'island' habitat is 555 preceded by admixture with the outgroups followed by a period of panmixia could also result 556 in the three sympatric sister species sharing a similar proportion of their ancestry with 557 outgroups; however, colonisation by a structured meta-population or hybrid swarm could result in the amount of shared ancestry with outgroups differing among ecotypes. (C) Multiple 558 559 independent colonisations of an isolated 'island' habitat over time, and episodic admixture upon secondary contact would result in the introgressed species sharing more of their ancestry 560 561 with the outgroups most closely related to the source population of this secondary colonisation. These three examples are not meant to be exhaustive, but simply illustrative of how different 562 563 evolutionary histories can result in the same majority-rule topology if evolutionary history is modelled as a single bifurcating tree. This figure is adapted from reference [14]. 564

565

567 Figure 2 Palaeogenomic sampling of divergent speciating lineages from sediment cores. (A) An isolated lake is founded by a single lineage (grey). During a period of spatial 568 569 separation within the lake, two daughter lineages are derived (red and blue) and are adapted to 570 local ecological conditions and associated mate choice. Upon secondary contact, mate choice 571 maintains this segregation of the two lineages. Sampling the contemporary lineages from the lake, one would reconstruct an ancestral history similar to that portrayed in Figure 1A. and 572 573 would be unable to distinguish whether reproductive isolation had become established despite 574 lineages having remained spatially overlapped throughout their post-colonisation history, or, 575 as in this case, whether reproductive isolation had developed during a period of spatial 576 isolation. (B) Sampling sediment cores of lakes and sequencing the sediment layers, or hard 577 body parts within them, provides a time series of genomic data that can elucidate the temporal patterns of spatial overlap, in addition to the chronology and tempo of genomic changes 578 579 associated with adaptation and speciation, i.e. the onset of selection. 580

In the example shown, the sediment core has been drilled in the area used exclusively by the blue
 lineage during the allopatric phase. Sampling multiple cores would establish the approximate
 distribution of both lineages through space and time.

Figure 3 Patterns of genomic differentiation due to sympatric and allopatric 582 583 divergence. (A) Schematic tree figures (top) are coloured to indicate changes in allele frequencies at a large effect barrier locus during divergence and introgression (indicated by red 584 horizontal arrow). During divergence-with-gene-flow in sympatry, there is genome-wide 585 homogenisation due to ongoing gene-flow (indicated by black horizontal arrows). The 586 587 segregation of alleles in different incipient species at large effect barrier loci associated with 588 ecological adaptation and reproductive isolation will predate the mean genome-wide coalescent time. This should be true whether the segregating alleles in barrier loci result from de novo 589 mutations (indicated by red star) during sympatry, standing variation that was present prior to 590 the sympatric phase, including from balanced polymorphisms, introgression and secondary 591 contact. Thus, such loci should stand out against a background of homogenised loci in genome-592 593 wide scans. (B) In many scenarios where genome-wide incompatibilities have evolved during 594 allopatry, which preclude gene-flow upon secondary contact, then TMRCA of alleles at 595 incompatibility loci will fall within the range of the genome-wide mean TMRCA, and both will predate secondary contact. This may not be ubiquitous. For example, balanced 596 polymorphisms which segregated upon speciation would still have a TMRCA that predated the 597 598 genome-wide mean.







(B)

Depth

Ľ

sediment

