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BRIEF COMMUNICATION

Ecological speciation in sympatric palms: 4. Demographic analyses support speciation of *Howea* in the face of high gene flow

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Running title (40 characters) Gene flow and speciation in *Howea* palms

Keywords Coalescence, sympatry, speciation, ddRAD

AUTHOR CONTRIBUTIONS

VS designed the research with contributions from ASTP, JI, and RKB. ASTP and TPS collected data. ASTP and JI analysed the data. IH and WB contributed to field collections. ASTP and VS wrote the manuscript. All authors commented on the manuscript.

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DATA ACCESSIBILITY

The sequence data are available at the Sequence Reads Archive under accession numbers PRJNA386480 and SRP063985.

1 BRIEF COMMUNICATION

2

3 Ecological speciation in sympatric palms: 4. Demographic analyses support speciation of 4 *Howea* in the face of high gene flow

5

6 Abstract

7 The idea that populations must be geographically isolated (allopatric) to evolve into separate
8 species has persisted for a long time. It is now clear that new species can also diverge despite
9 ongoing genetic exchange, but few accepted cases of speciation in sympatry have held up when
10 scrutinised using modern approaches. Here, we examined evidence for speciation of the *Howea*
11 palms of Lord Howe Island, Australia, in light of new genomic data. We used coalescence-based
12 demographic models combined with double digest restriction-site associated DNA sequencing of
13 multiple individuals and provide support for previous claims by Savolainen et al. (Nature 441: 210–
14 213, 2006) that speciation in *Howea* did occur in the face of gene flow.

15

16 Introduction

17 Sympatric speciation has re-emerged as a controversial topic, with recent analyses of genome-wide
18 data casting doubt on some of the most best-known examples, such as cichlid fish in Cameroonian
19 crater lakes (Martin *et al.* 2015). Due to a lack of confidence in ascertaining whether speciation has
20 taken place in the face of gene flow, our understanding of the genomic underpinning of such
21 processes has also remained piecemeal (Renaut *et al.* 2013; Cruickshank & Hahn 2014). Here, we
22 examined a case of speciation in *Howea* palms, a genus that comprises only two species, both
23 endemic to the subtropical Lord Howe Island (LHI; Savolainen *et al.* 2006). The island is isolated
24 (600 km from mainland Australia) and minute (<16 km²). Furthermore, modelling of the geological
25 history and sizes of LHI and nearby Ball's Pyramid rock showed that, for any pair of endemic sister
26 species that have diverged within the lifetime of the island, an allopatric phase in their divergence is
27 unlikely (Papadopoulos *et al.* 2011). This was critical in promoting *Howea* as a prime example of
28 sympatric speciation under a biogeographic definition (Savolainen *et al.* 2006; Mallet *et al.* 2009;
29 Coyne 2011; Papadopoulos *et al.* 2011). Marked flowering time differences between the species
30 indicate that prezygotic isolation is now strong and current levels of gene flow are likely to be low
31 (Savolainen *et al.* 2006; Babik *et al.* 2009; Dunning *et al.* 2016; Hipperson *et al.* 2016). Indirect
32 evidence of post-zygotic isolation due to selection against juvenile hybrids supports the hypothesis
33 that divergent selection has influenced the speciation process (Hipperson *et al.* 2016). Given that the
34 distributions of *Howea* palms overlap extensively and that they are wind pollinated, Savolainen *et*
35 *al.* (2006) argued that speciation is likely to have occurred in the face of high gene flow, which may
36 have reduced quickly as divergence progressed (Savolainen *et al.* 2006; Babik *et al.* 2009;

37 Papadopoulos *et al.* 2011, 2013b, 2014). However, previous efforts were based on a limited number
38 of markers (two gene sequences and amplified fragment length polymorphism; Savolainen et al.
39 2006, Babik et al. 2009), which leaves room for doubt about the precise timing of divergence and
40 did not allow for the quantification of the extent of the gene exchange between *H. forsteriana* and
41 *H. belmoreana*. Here, we characterise the demographic history of the *Howea* palms with a genome-
42 wide dataset to evaluate (i) whether genomic data support speciation with gene flow and (ii) how
43 genetic exchange progressed during speciation in sympatry.

45 **Material and Methods**

47 **DNA EXTRACTION AND DOUBLE DIGEST RAD-SEQUENCING**

48 Leaf tissue was collected and preserved in silica gel from 42 *H. belmoreana* and 54 *H. forsteriana*
49 individuals sampled at Far Flats on LHI where both species co-occur. Genomic DNA was extracted
50 from leaf tissue using DNeasy Plant Mini kits (Qiagen). Agarose gel electrophoresis was used to
51 assess the quality of each DNA extract and DNA quantification was performed with a Qubit 2.0
52 fluorometer (Life technologies). DNA samples were then processed following a modified version of
53 the double digest RADseq protocol of Peterson *et al.* (2012). Digestion of the DNA template of
54 each sample (250-1000ng) was performed by combining the sample with 0.1 µl *EcoRI*-HF (10 U;
55 NEB), 1µl *MspI* (10U; NEB), 5 µl NEB buffer 4, and nuclease-free water to a total volume of 50 µl.
56 Digestion reactions were incubated at 37°C for 3 hours. Digests were cleaned using the Agencourt
57 AMPure XP bead clean up (Beckman Coulter) and quantified using a Qubit 2.0 fluorometer.
58 Ligation of 6 base pairs (bp) - barcoded P1 (*EcoRI* overhang) and P2 (*MspI*) adaptors was
59 performed in individual reactions composed of ca. 400 ng of the digested DNA product, 2 µl of
60 each adaptor (4 nM), 2 µl T4 DNA ligase (4000 U; NEB), 4 µl 10x Ligase buffer (NEB) and
61 nuclease-free water to a total of volume 40 µl. Ligation reactions were incubated at 23°C for 30
62 min, followed by 65°C for 10 min, and then cooled to 18°C at a rate of 2°C / 90 seconds. Samples
63 were pooled into batches of 12 containing compatible sets of barcodes, and cleaned using
64 Agencourt AMPure XP beads. Each pool was size-selected between 344-408 bp using a Pippin Prep
65 electrophoresis system (Sage Science). Each size-selected pool was PCR amplified in 6-8 reactions
66 each composed of 6 µl of DNA template, 2 µl of each ddRAD primer, 0.2 µl Phusion *Taq*
67 polymerase (NEB), 0.4 µl dNTPs (10 mM; NEB), 4 µl 5x Phusion HF buffer (NEB) and 5.4 µl
68 nuclease-free water. PCR reactions were run on a Veriti 96-Well Fast Thermal Cycler (Applied
69 Biosystems) at 98°C for 30s, 12 cycles of 98°C for 15s, 65°C for 30s, 72°C for 30s, and final 72°C
70 for 7 min. PCR reactions for each pool were combined and cleaned using Agencourt AMPure XP
71 beads. Each cleaned pool was diluted to 4 nM and four pools with compatible barcodes were

combined to produce libraries of 48 uniquely barcoded samples. Two ddRAD sequence libraries were prepared in this way. All libraries were sequenced to 100 bp, paired-end on an Illumina HiSeq 2500 (one lane per library) at the MRC Clinical Sciences Centre, Imperial College London. This generated an average of 6,186,469 reads per sample. Genotyping of ddRAD data was performed using the *STACKS* pipeline, building upon the catalogue generated in Papadopoulos *et al.* (2019b). To expand the catalogue to encompass haplotypes present in both *Howea* species, samples were assembled into loci using *USTACKS* (-m20, -M3) and these stacks were merged into the existing catalogue allowing 3 mismatches between loci in different individuals. To genotype individuals, loci were assembled with lower coverage in *USTACKS* (-m5, -M3) and these stacks were mapped to the catalogue loci.

DEMOGRAPHIC MODELLING

For the demographic analysis, the Far Flats individuals were genotyped at 4,581 loci (23,109 SNPs) with fewer than 20 missing individuals (minor allele frequency = 0.05). To account for missing data, we projected the number of individuals down to 36/54 *H. forsteriana* and 23/42 *H. belmoreana* and calculated the joint folded site-frequency-spectrum using *δaδi* (Gutenkunst *et al.* 2009). We then inferred the demographic history of *Howea* from the site-frequency-spectrum using two methods: (i) the composite-likelihood framework implemented by *fastsimcoal2* (Excoffier *et al.* 2013) and (ii) the diffusion approximation approach implemented in a modified version of *δaδi* (Gutenkunst *et al.* 2009; Tine *et al.* 2014). This modified version of *δaδi* accounts for variation in the rate of gene flow across the genome by dividing the genome into two types of loci (in P and 1-P proportions) with potentially different migration rates.

For *fastsimcoal2*, we estimated parameters 60 times for each of 10 demographic models to determine the combination of parameters with the highest likelihood. These models are shown on Figure 1a and assume either no population growth, or population growth: model 1 - speciation without gene flow; model 2 - speciation with recent gene flow following secondary contact; model 3 - speciation with initial gene flow; model 4 - speciation with constant gene flow; and model 5 - speciation with two distinct periods of migration where gene flow may vary. Model fit was assessed using the Akaike Information Criterion (AIC). Non-parametric bootstrapping (100 simulated datasets under the best model with 10 rounds of parameter estimation for each simulation) was used to estimate 95% confidence intervals for each parameter for the best model.

For *δaδi*, we compared the fit of the same 10 models as above plus another eight, that is, models 2-5 with and without population growth, but also including heterogeneous rates of migration across the

107 genome. We ran two rounds of simulated annealing (one hot and one cold) followed by a final
108 round of Broyden–Fletcher–Goldfarb–Shanno optimisation. For each of the 18 models, we
109 performed a minimum of 30 runs to ensure thorough estimation of the maximum likelihood and
110 used AIC to perform model selection. For the best fitting model, we then ran 30 bootstrap replicates
111 (ensuring that each replicate had at least 10 runs) using the built-in *δaδi* procedure to get confidence
112 intervals around the parameter estimates.

113

114 To calibrate the demographic models in *fastsimcoal2* and *δaδi*, we estimated the substitution rate in
115 *Howea*. We first built a phylogenetic tree using genome wide data for all available palm species.
116 Transcriptome assemblies were obtained for *H. forsteriana* (Dunning et al 2016), *Elaeis guineensis*
117 and *E. oleifera* (African and American oil palms; Singh *et al.* 2013). Short read transcript data for
118 *Phoenix dactylifera* (date palm; Al-Mssallem *et al.* 2013) were obtained from the Short Read
119 Archive (Accession Number SRR341952) and a transcriptome assembly was produced using
120 *Trinity* (Grabherr et al. 2011) with default parameters and min_kmer_cov set to 2. *TransDecoder*
121 was used to predict open reading frames (ORF) with a minimum length of 100 amino acids, and the
122 longest ORF was selected. Reciprocal blast hits for all four palm species were established by
123 collating reciprocal best blast results from a pair of species with the remaining two species; this
124 rendered an initial set of orthologous alignments for the four species. An *M-coffee* (Wallace et al.
125 2006) pipeline was used to score these alignments. The nucleotide sequences were translated into
126 protein using *t-coffee*, then the protein sequences were aligned with *MAFFT* (Katoh & Toh 2008),
127 *Muscle* (Edgar 2004), *t-coffee* (Notredame et al. 2000) and *k-align* (Lassmann & Sonnhammer
128 2005) and translated back to DNA. Low quality positions with scores lower than 8 were trimmed.
129 Protein alignments were used to guide gap placement in the nucleotide alignments. Finally,
130 *maxalign* was used to check for any misaligned sequences in the alignments (Gouveia-Oliveira et
131 al. 2007). Only alignments where all the orthologous sequences were properly aligned and deemed
132 as high quality were retained and concatenated into a single file. *CODEML* (Yang 2007) was used
133 to calculate 4-fold degenerate sites, which are considered not to be subject to selection. Finally,
134 *MCMCtree* (Yang 2007) was used to build a tree. To calibrate this tree, we used secondary
135 calibrations from the most complete phylogenetic tree of the palm family (Baker & Couvreur 2013).
136 Independent substitution rates were inferred for each branch in the tree, and HKY85 (the most
137 complex substitution model available in *MCMCtree*) was selected. The MCMC chain was run to
138 gather 20,000 samples after convergence had been achieved. The first 10,000 iterations were
139 discarded as a burn-in. Assuming a generation time of 10 years for *Howea* palms (Lord Howe
140 Island Nursery, *pers. comm.*), we estimated the rate of substitution of the branch leading to *H.*
141 *forsteriana* to be 1.3×10^{-8} mutations per site per generation.

Results and discussion

The demographic analyses provide strong support that *Howea* palms on LHI diverged in the face of ongoing gene flow. We have categorised the demographic models into four groups: (i) no population growth; (ii) population growth; (iii) no growth but heterogeneous migration across the genome; (iv) growth with heterogeneous migration across the genome. Using *fastsimcoal2*, where heterogeneous migration across the genome does not apply, model 5 (divergence with two periods where gene flow may vary) was most likely when no growth was modelled (Table 1). When population growth was permitted, model 5 was within 10 AIC from models 2 and 3, and therefore these three models are undistinguishable (Burnham & Anderson 2002) (Table 1). Using *δaδi*, model 5 was always the most likely model. However, when growth and heterogeneous migration across the genome were permitted, model 5 was indistinguishable from model 3 (speciation with initial gene flow; Table 1). The more complex growth models had smaller AIC than the no growth counterparts (Table 1), pointing to a period of exponential growth following species divergence. – However, for both *δaδi* and *fastsimcoal2* the confidence intervals included the point estimates for all parameters only for the simplest scenario (i.e., without population size changes or heterogeneous rate of genome flow across the genome, Table S1). Other scenarios had greater levels of uncertainty indicating that our data were not sufficient to constrain such complex models and the point estimates for these models are likely to be unreliable. As the two demographic methods produced comparable results for the no growth scenarios, we focus our discussion on these simpler cases. In models including two distinct periods of migration, we did not constrain the earlier migration rate to be higher than the more recent migration rate. Nevertheless, in all of these models the earlier rate of gene flow is estimated to be higher than more recent rates, supporting a reduction in gene flow during speciation. A similar pattern was found during the divergence of *Senecio* on Mt. Etna (Filatov *et al.* 2016).

Figure 1b shows the detailed scenario for model 5 without growth. Confidence intervals for all parameters for *δaδi* were much wider than for *fastsimcoal2* (Table S1), so we mainly discuss results from the latter. To assess the fit of the data to the model, we calculated the likelihood ratio G-statistic (Composite likelihood ratio = 470.2) and compared this to the null distribution of simulated values (Excoffier *et al.* 2013). The observed value is in the tail of the simulated distribution (above the 99.9 percentile; range = 327.4 – 485.9). This is expected as these demographic models are a

176 simplification of the real history of the species, whereas the null distribution is based on data
 177 simulated under the simple model (Excoffier *et al.* 2013).

178 Migration was initially two orders of magnitude higher from the smaller *H. forsteriana* population
 179 into the larger *H. belmoreana* lineage (proportion of migrants received per generation = 4.00×10^{-4}
 180 vs 4.71×10^{-6} ; effective migrants per generation, $N_m = 13.01$ vs 0.27). This initial period was
 181 followed by a reduction in gene flow (proportion of migrants received per generation = 1.6×10^{-7} vs
 182 3.3×10^{-7} ; $N_m = 0.01$ vs 0.02). This is consistent with *H. forsteriana* being derived from a
 183 *belmoreana*-like ancestor that colonised a new habitat in which the *H. belmoreana* genetic
 184 background was selected against. The initially high N_m (mean = 6.64) is in the top 6% of values
 185 found in other examples speciation with gene flow (range = 0.00 - 25.22; Pinho & Hey 2010;
 186 Filatov *et al.* 2016) and the proportion of migrants is similar to that found in the sympatric
 187 Nicaraguan cater lake cichlids (7.48×10^{-5} - 8.51×10^{-5} ; Kautt *et al.* 2016). These migration estimates
 188 fall below those expected under population genetic definitions of sympatric speciation ($m = 0.5$;
 189 Fitzpatrick *et al.* 2008). However, it is important to note that our migration estimates are averages
 190 over long periods of time, forced by a model that has an abrupt transition from one population to
 191 two populations. If we had a model that allowed a progressive reduction in gene flow, we may have
 192 seen values close to 0.5 at the start and then a rapid reduction as the initial barriers were built.
 193 Unfortunately, we do not have the data to fit such a model. Of course, it would also have strong
 194 heterogeneity across the genome. Based on a generation time of 10 years, we found that the species
 195 initially diverged roughly 2.7 million years ago, which is older than previously estimated by
 196 phylogenetic analysis of two genes (1.92 +/- 0.52 million years ago; Savolainen *et al.* 2006).
 197 Allowing for different rates of migration across the genome (using $\delta a \delta i$) resulted in a more recent
 198 divergence time than other models at 1.41 Mya, but this fell outside the bootstrap confidence
 199 intervals. Our estimates predate the proposed mid-Pleistocene age of the current calcareous deposits
 200 of the island (Brooke 2003; Woodroffe *et al.* 2006; Papadopoulos *et al.* 2013a). It is possible that
 201 colonisation was on calcareous soils pre-dating those currently on the island, and which would have
 202 eroded since then. Alternatively, colonisation of mid-Pleistocene calcareous sites may have taken
 203 place after divergence with the initial selection pressures stemming from other sources, such as
 204 water availability or salinity (Papadopoulos *et al.* 2013b). The timing of the shift to a lower migration
 205 rate differs substantially between the methods; *fastsimcoal2* points to a large reduction in migration
 206 ~40,000 years after initial divergence, whereas a $\delta a \delta i$ indicates smaller change that took place much
 207 more recently (100,000 year after divergence). Estimates of current population sizes (*H. forsteriana*
 208 $N_e = 32,510$; *H. belmoreana* $N_e = 57,181$) are similar to the estimated census population size (all
 209 LHI *Howea* ~100,000, with 2.7 times as many *H. belmoreana* as *H. forsteriana*) (Savolainen *et al.*
 210 2006; Hipperson *et al.* 2016). The *fastsimcoal2* estimated ancestral N_e (424,288) is within the

211 bounds of possibility, but is likely to be an overestimate as variation in coalescence time due to
 212 selection in the ancestral species may cause *fastsimcoal2* to explain excess variance by inferring
 213 very large N_e . The *δaδi* ancestral N_e was an order of magnitude lower (12,570), and it is therefore
 214 unclear which estimate is best. It seems likely that small initial colonising group came from a large
 215 mainland population and then grew rapidly, but this not possible to determine with our analyses.
 216 The inconsistent results for the more complex models show that our analyses are limited by the
 217 data. Refinements could be made using whole genome re-sequencing data, although the large
 218 genome size and complexity makes this challenging. Additionally, more detailed data would allow
 219 the inclusion of other parameters in more complex frameworks that have emerged recently - such as
 220 that of Roux *et al* (2016), which models heterogeneity in coalescence times due to selection.
 221 Currently, reproductive isolation between the species is strong, although not complete given that
 222 occasional fertile hybrids are formed (Babik *et al.* 2009). Our results are consistent with the idea
 223 that initial local adaptation and post-zygotic isolation were supplemented by the rapid completion of
 224 pre-zygotic isolation through flowering time differences (Papadopoulos *et al.* 2013b, 2014).
 225 Furthermore, given the supporting evidence these results allow us to rule out continuous absence of
 226 gene flow, even though LHI was larger at the time of speciation (Papadopoulos *et al.* 2011, 2013a;
 227 Linklater *et al.* 2018). Unlike recent genomic reanalyses of classic cases of speciation in sympatry,
 228 our results support the proposition that *Howea* palms must have diverged with continuous gene
 229 flow. We note, however, that genomic data by themselves may only permit the rejection of the
 230 simplest form of allopatry (Yang *et al.* 2017). In this sense, our demographic analyses should be
 231 seen in concert with other lines of evidence such as the geological history of LHI, lack of
 232 population structure on LHI, and the finding of candidate reproductive isolation genes (Dunning *et al.*
 233 *et al.* 2016). Furthermore, our analyses indicate that divergence may have predated the origin of the
 234 calcarenite soils on LHI, and therefore identifying candidate ‘speciation genes’ with functions
 235 related to drought and salt tolerance may be more important than extremes of pH.
 236
 237

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Table 1. Summary statistics for model selection. The most likely models within 10 AIC are highlighted in bold.

Model	Method	Max. Ln (likelihood)	No. of parameter s	AIC	Delta AIC
1 (speciation without gene flow)	<i>fastsimcoal2</i>	-175730	4	351467	3092
2 (speciation with recent gene flow following secondary contact)	<i>fastsimcoal2</i>	-174992	7	349997	1622
3 (speciation with initial gene flow)	<i>fastsimcoal2</i>	-175522	7	350109	1734
4 (speciation with constant gene flow)	<i>fastsimcoal2</i>	-175049	6	351059	2683
5 (species divergence with two periods where gene flow may vary)	<i>fastsimcoal2</i>	-174955	9	349928	1552
1 + growth	<i>fastsimcoal2</i>	-174898	6	349808	1433
2 + growth	<i>fastsimcoal2</i>	-174179	9	348375	0
3 + growth	<i>fastsimcoal2</i>	-174181	9	348377	2
4 + growth	<i>fastsimcoal2</i>	-174265	8	348548	173
5 + growth	<i>fastsimcoal2</i>	-174180	11	348382	7
1	<i>δaδi</i>	-2602.12	4	5210	2705
2	<i>δaδi</i>	-2051.04	7	4114	1609
3	<i>δaδi</i>	-2055.46	7	4123	1617
4	<i>δaδi</i>	-2067.86	6	4146	1640
5	<i>δaδi</i>	-2012.61	9	4041	1536
1 + growth	<i>δaδi</i>	-1747.95	6	3506	1000
2 + growth	<i>δaδi</i>	-1384.78	9	2786	280
3 + growth	<i>δaδi</i>	-1427.55	9	2871	366
4 + growth	<i>δaδi</i>	-1431.38	8	2877	371
5 + growth	<i>δaδi</i>	-1377.74	11	2775	270
2 + heterogeneous M	<i>δaδi</i>	-1741.57	10	3501	996
3 + heterogeneous M	<i>δaδi</i>	-1786.64	10	3591	1086
4 + heterogeneous M	<i>δaδi</i>	-1785.31	9	3587	1081
5 + heterogeneous M	<i>δaδi</i>	-1730.07	14	3486	981
2 + growth + heterogeneous M	<i>δaδi</i>	-1255.63	12	2533	28
3 + growth + heterogeneous M	<i>δaδi</i>	-1241.78	12	2506	0
4 + growth + heterogeneous M	<i>δaδi</i>	-1256.03	11	2532	26

Fig. 1. Coalescence analyses of demography in *Howea*. (a) Five models were tested, either assuming constant population sizes, allowing for exponential population growth through time, or heterogeneous migration across the genome (18 scenarios in total). Model 1: speciation without gene flow; 2: speciation with recent gene flow following secondary contact; 3: speciation with initial gene flow; 4: speciation with constant gene flow; and 5: species divergence with two periods where gene flow may vary. (b) Parameter estimates for the best fitting model with no growth estimated in *fastsimcoal2* (model 5).

Table S1. Inferred parameters for the selected model 5

Parameter	<i>fastsimcoal2</i>			<i>δaδi</i>		
	Point estimation	Lower CI	Upper CI	Point estimation	Lower CI	Upper CI
Ancestral Ne	424288	125435	547787	12570	11589	19889
<i>H. forsteriana</i> Ne	32510	31192	34006	31806	29809	33687
<i>H. belmoreana</i> Ne	57181	54802	59951	49054	47272	52285
T ₀	2.66	2.56	4.52	2.71	0	40.34
T ₁	2.38	2.10	2.96	0.11	0	2.84
MIG _{F→B} (T ₀ -T ₁)	4.00 x 10 ⁻⁴	5.87 x 10 ⁻⁵	4.21 x 10 ⁻⁴	5.67 x 10 ⁻⁷	2.97 x 10 ⁻⁹⁹	1.45 x 10 ⁻⁵
MIG _{B→F} (T ₀ -T ₁)	4.71 x 10 ⁻⁶	1.32 x 10 ⁻⁵	5.94 x 10 ⁻⁵	7.06 x 10 ⁻¹¹	2.01 x 10 ⁻¹³⁸	1.27 x 10 ⁻⁵
MIG _{F→B} (T ₁ -Present)	1.57 x 10 ⁻⁷	1.57 x 10 ⁻⁷	2.27 x 10 ⁻⁷	1.49 x 10 ⁻⁷	3.50 x 10 ⁻⁷⁰	6.28 x 10 ⁻⁶
MIG _{B→F} (T ₁ -Present)	3.30 x 10 ⁻⁷	2.82 x 10 ⁻⁷	4.12 x 10 ⁻⁷	3.78 x 10 ⁻⁷	5.36 x 10 ⁻³¹	6.76 x 10 ⁻⁶

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