



Ocean warming increases the nitrogen demand and the uptake of organic nitrogen of the globally distributed seagrass *Zostera marina*

Alexandre, Ana; Quinta, Raquel; Hill, Paul; Jones, Davey L.; Santos, Rui

Functional Ecology

DOI:
[10.1111/1365-2435.13576](https://doi.org/10.1111/1365-2435.13576)

Published: 01/07/2020

Peer reviewed version

[Cyswllt i'r cyhoeddiad / Link to publication](#)

Dyfyniad o'r fersiwn a gyhoeddwyd / Citation for published version (APA):
Alexandre, A., Quinta, R., Hill, P., Jones, D. L., & Santos, R. (2020). Ocean warming increases the nitrogen demand and the uptake of organic nitrogen of the globally distributed seagrass *Zostera marina*. *Functional Ecology*, 34(7), 1325-1335. <https://doi.org/10.1111/1365-2435.13576>

Hawliau Cyffredinol / General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

1 Ocean warming increases the nitrogen demand and the uptake of organic nitrogen of the
2 globally distributed seagrass *Zostera marina*

3

4 Ana Alexandre^{1*}, Raquel Quintã², Paul W. Hill³, Davey L. Jones^{3,4}, Rui Santos¹

5

6 ¹Marine Plant Ecology Research Group, Centre of Marine Sciences (CCMAR),
7 University of Algarve, Gambelas, 8005-139 Faro, Portugal

8

9 ²Portuguese Institute of the Sea and Atmosphere (IPMA) Aquaculture Research Station
10 (EPPO), Av. do Parque Natural da Ria Formosa, 8700-194 Olhão, Portugal

11

12 ³School of Natural Sciences, Bangor University, Gwynedd, LL57 2UW, United
13 Kingdom

14

15 ⁴The Oceans Institute, The University of Western Australia, Perth, WA 6009, Australia

16

17 *Corresponding author: aalexandre@ualg.pt

18

19

20

21

22

23

24

25 Running head: Shifts in *Zostera marina* N uptake with temperature

26

27

28

29

30 Abstract

31

32 1. The impact of global warming on the metabolic state of a species may be examined
33 by either measuring physiological rates across a latitudinal gradient or by assessing
34 short-term responses under experimentally controlled temperature regimes. The
35 combination of the two approaches is seldom used but it provides valuable information
36 on an organism's responses to temperature at broader temporal and spatial scales while
37 allowing the isolation of temperature effects from other environmental variables.

38 2. Here we used both approaches to assess the warming effects on the total acquisition
39 of dissolved inorganic nitrogen (DIN; nitrate, ammonium) and organic N (DON; amino
40 acids, peptides) by the globally widespread seagrass *Zostera marina*. DIN and DON
41 uptake rates were measured in plants from three sites covering the species latitudinal
42 distribution in Europe (Iceland, United Kingdom and Portugal). The responses of DIN
43 and DON uptake rates of plants from the middle latitude (UK) to a latitudinal range of
44 temperatures (8, 12 and 17 °C) were also measured. We further examined the microbial
45 uptake of DON along the latitudinal distribution and whether temperature is the main
46 driver of that uptake.

47 3. Our results showed that warming greatly increased the total N uptake by *Z. marina*
48 and also the relative contribution of DON to total N acquisition. The microbial uptake
49 of DON increased towards warmer latitudes, and temperature was the main driver of
50 these observations.

51 4. Ocean warming will increase the nitrogen demand of *Z. marina* and this demand may
52 be met by an increasing uptake of organic nitrogen forms. This indicates that *Z. marina*,
53 and probably other seagrass species, can be winners under global change as nitrogen

54 uptake capacity will not limit growth driven by increased photosynthetic assimilation of
55 CO₂.

56

57 Keywords: DIN uptake, DON uptake, global warming, latitudinal distribution,
58 microbial uptake, seagrasses, temperature, *Zostera marina*

59

60 Introduction

61

62 Global warming (IPCC, 2018) is affecting the metabolic rates of organisms and the
63 distribution of species, shaping the structure and functioning of marine ecosystems and
64 their trajectories (Walther et al., 2002; Poloczanska et al., 2013). The potential impacts
65 of rising temperatures on organisms and ecosystems are often investigated by short-term
66 experimental temperature manipulation (see Shaver et al., 2000, Rustad et al., 2001 and
67 Aronson & McNulty, 2009 for a review of methods). Controlled temperature
68 experimentation allows separation of temperature effects from other confounding
69 environmental factors that covary with temperature in natural conditions. However, this
70 approach may fail to accurately predict both the magnitude and the direction of species
71 responses due to the transient nature (short-duration) of the experiments and the step
72 increases in experimental temperatures, which are often unrealistic (Rustad, 2008).

73 Testing a species response across a wide latitudinal range is an alternative to address the
74 effects of temperature on organisms because sites along the gradients integrate the
75 geographical climate variation over large temporal scales, thus providing empirical-
76 based predictions of consequences of temperature changes at broader temporal and
77 spatial scales (Fukami & Wardle, 2005; De Frenne, 2013). In the present study, we
78 combined latitudinal gradient analysis with laboratory responses to temperature

79 manipulation to address how climate warming may affect seagrasses' nutrient
80 acquisition, a key physiological process driving production.

81 Seagrasses represent one of the most heterogeneous landscape structures of shallow-
82 water marine ecosystems in the world, however, it is predicted that they will be highly
83 impacted by climate change (Bostrom et al., 2006; Chefaoui et al., 2018). Warming may
84 affect seagrasses directly by altering nutrient uptake rates (e.g. nitrogen (N) and
85 phosphorus (P)), or indirectly via a potential acceleration of N mineralisation that may
86 increase seagrass productivity by increasing inorganic nutrient availability in nutrient-
87 limited environments. This process has been well described in terrestrial ecosystems
88 (Rustad et al., 2001; Bai et al., 2013) but is poorly understood in seagrass communities
89 (Duarte et al., 2018).

90 Seagrasses can take up both dissolved inorganic nitrogen (DIN; nitrate, ammonium) and
91 dissolved organic nitrogen (DON; amino acids, peptides), with a general preference for
92 the uptake of ammonium (NH_4^+) (Alexandre et al., 2011 and references therein), but it
93 is not known how this preference may shift with temperature. In fact, there is little
94 information on whether, or how, seagrass nutrient uptake will respond to global
95 warming (Moore & Short, 2006). A recent study showed that nitrate (NO_3^-) uptake by
96 *Zostera marina* increased by 50% under warming, however, this was undertaken under
97 an unrealistic warming scenario (10 to 18/25 °C) and exposure to a high nitrate
98 concentration (100 μM ; Kaldy, 2014). In terrestrial habitats, it is expected that increased
99 soil temperature will increase plant uptake of N and P in species from warm habitats
100 more than in those from colder environments (BassiriRad, 2000), but warming is also
101 expected to increase plant N and P uptake in arctic species (Jonasson et al., 1999). The
102 few available data also indicate that increased soil temperature elicits a differential
103 effect on the uptake of ammonium versus nitrate, i.e. the ratio of ammonium to nitrate

104 uptake will consistently decrease with increasing temperature in various plant species
105 (BassiriRad, 2000). Potential future changes in the availability of the different N forms
106 in the sediment and water column alongside changes in seagrass N preference under
107 global warming might shift seagrasses' competitive interactions with other species.
108 N availability is one of the major factors limiting primary productivity of seagrasses,
109 particularly in oligotrophic environments where nutrient concentrations are very low
110 (e.g. Agawin et al., 1996; Alcoverro et al., 1997). Seagrasses are well adapted to these
111 conditions, being highly efficient at taking up ephemeral pulses of dissolved inorganic
112 nitrogen (DIN) (Alexandre et al., 2017). On the other hand, the uptake of dissolved
113 organic nitrogen (DON) by seagrasses has been largely overlooked, even though this
114 represents a large component of the total dissolved N pool in coastal waters and is a
115 precursor to DIN formation (Sharp, 1983, 2002; Bronk et al., 2007). In fact, seagrasses
116 can take up N from both simple (amino acids and urea) and complex (peptides) organic
117 substrates at ecologically relevant rates (Vonk et al., 2008; Van Engeland et al., 2011;
118 Alexandre et al., 2015), suggesting that DON may be an important N source for these
119 plants.

120 *Z. marina* is the most abundant seagrass species in the northern hemisphere spreading
121 over a wide latitudinal range, from warm subtropical to arctic regions (Cabello-Pasini et
122 al., 2003; Green & Short 2003). The use of DON by this species was studied at the
123 southern limit of its geographical distribution range, in Ria Formosa lagoon, south
124 Portugal (Alexandre et al., 2015). In this coastal lagoon, DON forms a significant pool
125 of bioavailable N in the water column and in sediments (~ 60 % of total soluble N) and
126 the uptake of DON (as amino acids and peptides) by *Z. marina* represented a significant
127 fraction (~ 30 %) of the total N taken up by the species. However, it is not known how
128 widespread is the use of DON by *Z. marina* and how it may vary along the species'

129 latitudinal range. Latitude may be relevant as the availability of DON is expected to
130 increase at higher and colder latitudes as DIN regeneration from the mineralisation of
131 organic matter decreases due to decreasing rates of microbial N mineralisation (Chapin
132 et al., 1993; Mozdzer et al., 2014). This trend has been reported in coastal saltmarshes
133 colonized by *Spartina alterniflora* along a latitudinal distribution range (30 - 44 °N)
134 (Mozdzer et al., 2014). In boreal forests, the low mineralisation rates result in one order
135 of magnitude higher concentrations of DON than those of DIN (Näsholm et al., 2009).
136 Further, in arctic ecosystems the ratios of primary production to N mineralisation are
137 high and consequently arctic vascular plants use amino acid and peptidic N to avoid N
138 limitation derived from low DIN regeneration (Hill et al., 2011). It is also energetically
139 more favourable to take up DON in comparison to DIN as the compounds taken up can
140 be directly used in cell metabolism.

141 This study assesses how global warming may affect the N uptake rates by the globally
142 widespread seagrass *Z. marina*. Specically we tested the following questions: 1) What is
143 the availability of DON relative to DIN in *Z. marina* meadows along a latitudinal
144 gradient?, 2) What is the latitudinal variation of the uptake rates of DIN and DON by *Z.*
145 *marina* plant parts (leaves and roots)?, 3) What is the latitudinal variation of the
146 contribution of DIN and DON to the total N budget of *Z. marina*?, 4) What is the
147 latitudinal variation of the microbial uptake of DON and 5) Is temperature the main
148 driver of the latitudinal variations? Laboratory experiments will allow to assess if the
149 latitudinal observations are mainly driven by temperature and consequently what will be
150 the response of *Z. marina* meadows N cycling to global warming.

151

152 Methods

153

154 Sampling sites

155 The latitudinal variation of *Zostera marina* uptake rates were assessed in the summer of
156 2014, when seagrass productivity is highest, in plants collected from meadows located
157 along the species latitudinal distribution range in Europe: a high-latitude meadow
158 located in Alftafjordur, Iceland (64°59'N; 22°36'W), a mid-latitude meadow located in
159 Porthdinllaen, United Kingdom (52°56'N, 4°33'W), and a lower-latitude meadow
160 located in Ria Formosa lagoon, Portugal (36°58'N, 8°02'W) (see Fig. S1 in Supporting
161 Information). Iceland (high-latitude) and Portugal (lower-latitude) represent the
162 northern and southernmost limits of the species distribution, respectively, and United
163 Kingdom (mid-latitude) an intermediate point. The seagrass uptake rates along the
164 latitudinal gradient was assessed by measuring uptake rates of each meadow at the local
165 mean annual temperature. These were combined with temperature manipulation
166 experiments with plants collected at the intermediate site within the distribution (UK),
167 to isolate the temperature effects from other environmental variables. Plant biomass and
168 N availability at each site were used to calculate the contribution of DIN and DON to
169 the total N budget of *Z. marina*.

170 Samples (n = 5) of seawater (20 mL each) and sediment porewater were collected
171 during low tide (approximate water height of 20 cm) at each meadow to determine the
172 concentration of inorganic (ammonium and nitrate) and organic nitrogen (free amino
173 acids and peptides), as well as total soluble nitrogen (TN). Seawater samples were
174 filtered (Whatman cellulose acetate filters, 0.45 µm pore size) and stored at -20 °C until
175 analysis. A total of four sediment cores (3 cm diameter, 5 cm depth) were pooled for
176 each porewater sample (~20 mL) to reduce the effect of nutrient heterogeneity. The four
177 pooled cores were randomly collected within the meadow at an approximate distance of
178 10 cm, and the different groups of pooled cores were collected at a distance of at least 2

179 m. The sediment samples were centrifuged (2300 g, 15 min at 4 °C) and the
180 supernatants were filtered (Whatman cellulose acetate filters, 0.45 µm pore size) and
181 frozen until analysis.
182 Shoot density (no. shoots m⁻²) and areal biomass (g dry weight m⁻²) were also
183 determined for each site by counting the number of shoots and weighing the dried plants
184 (48 h at 60 °C) collected within a sampling quadrat (20 cm x 20 cm, n = 3). The
185 quadrats were haphazardly tossed inside the meadow, ensuring distances higher than 20
186 cm between different tossings.

187

188 Plant collection and acclimation

189 *Z. marina* plants were collected during low tide from the meadows and cleaned of
190 adhering sediment, avoiding damage of the root hairs, and epiphytes removed from the
191 leaves by gentle hand scraping. The plants were transported in seawater collected at the
192 sampling site to the laboratory, where they were acclimated for 3 days at local summer
193 conditions of light and mean annual temperatures (Marine and Freshwater Research
194 Institute; climate-data.org; Global Sea Temperature database). In Iceland, plants were
195 acclimated at 8 °C with a 20:4 h light:dark cycle; in the UK, plants were acclimated at
196 12 °C with a 18:6 h light:dark cycle and in Portugal at 17 °C with a 16:8 h light:dark
197 cycle.

198

199 *Z. marina* nitrogen uptake experiments

200 Uptake rates of inorganic (ammonium and nitrate) and organic (alanine and trialanine)
201 N were determined by incubating whole plants fully immersed in 300 mL of N-free
202 artificial seawater (0.2 µm filtered water, salinity of 35, pH 8.2) containing either
203 ¹⁵NH₄Cl, ¹⁵KNO₃ or ¹³C₃H₇¹⁵NO₂ (L-alanine) at concentrations of 5, 25, 50 and 100 µM

204 or $^{13}\text{C}_9\text{H}_{17}^{15}\text{N}_3\text{O}_4$ (L-trialanine) at concentrations of 5, 15 and 50 μM (atom % = 98,
205 alanine obtained from Cambridge Isotope Laboratories; trialanine obtained from CK
206 Gas Products, Hook, UK). Above and belowground biomass of incubated tissues are
207 given in Table S1. Alanine was selected because it is one of the amino acids with higher
208 rates of occurrence in the proteins of all types of organisms and therefore is important in
209 N cycling. Trialanine was chosen because of its ecological significance as it represents a
210 common structure that is released by protein hydrolysis. All experiments were
211 performed at constant light ($\approx 100 \mu\text{mol quanta m}^{-2} \text{ s}^{-1}$) for 0.5 h and at each site-
212 specific acclimation temperature. Incubations with the four N sources at the different
213 nutrient concentrations were done simultaneously whereas replicates ($n = 3$) were done
214 sequentially to overcome feasibility constraints derived from the setup of a large number
215 of incubations chambers necessary to test all treatments simultaneously (N sources and
216 N concentrations, $n = 16$ chambers).

217 At the end of uptake experiments, the leaves were immediately separated from the
218 rhizomes and roots. Tissues were rinsed with deionised water to remove adhering salt
219 and isotopic labels before being dried (60°C , 48 h), and reduced to a fine powder for
220 analysis of the atom % of ^{15}N and total N content.

221

222 Microbial DON uptake experiments

223 At each meadow, seawater samples and sediment cores of 3 cm diameter and 5 cm
224 depth were collected ($n = 3$), placed in 50 mL polypropylene centrifuge tubes, sealed
225 immediately and transported on ice to the laboratory. Sediment samples were
226 acclimated for three days to the site-specific temperature (8, 12 and 17°C in Iceland,
227 UK and Portugal, respectively). Sediment sub-samples of 1 g of fresh weight were then
228 placed in 1.5 mL polypropylene tubes. The water content of natural sediments (~ 0.5

229 mL g⁻¹ fresh weight) was previously determined by drying at 105 °C for 24 h and used
230 to calculate the amount of seawater needed to add to the sediment in order to obtain a
231 final incubation volume of 1 mL after sediment centrifugation. Artificial seawater with
232 a salinity equal to the natural seawater of each site was purged with N₂ to remove
233 oxygen and added to the sediment tubes to reach the final incubation volume of 1 mL of
234 water content in each tube. Microbial uptake rates of amino acids and peptides were
235 determined by adding ¹⁴C-labelled alanine or trialanine solutions (20 µL; 2.8 kBq mL⁻¹
236 seawater or g⁻¹ soil; American Radiolabeled Chemicals, St Louis, MO, USA) to the
237 seawater and sediment sub-samples at a range of concentrations (5, 25, 50 and 100 µM
238 for alanine and 5, 15 and 50 µM for trialanine). Samples from Iceland, UK and Portugal
239 were incubated respectively at 8, 12 and 17 °C for 0.5 h under vigorous shaking. After
240 incubation, samples were centrifuged (18 000 g, 10 min) and the supernatant recovered.
241 Subsequently, 0.2 mL of HCl (1 M) was added to release any H¹⁴CO₃⁻ trapped in
242 solution (Brailsford et al., 2019) and the ¹⁴C activity in the supernatant (0.3 mL)
243 determined on a Wallac 1404 liquid scintillation counter with automatic quench
244 correction (Wallac EG&G, Milton Keynes, UK) after addition of 4 mL of HiSafe3
245 liquid scintillation cocktail (Perkin Elmer Corp., Waltham, MA). Unfortunately, it was
246 not possible to measure the microbial uptake of alanine and trialanine in the water in the
247 absence of sediment as the ¹⁴C activity measured at the end of the experiment was never
248 lower than the initial ¹⁴C activity, even when the incubation time was extended (i.e. no
249 measureable uptake).

250

251 Effects of temperature on N uptake

252 To evaluate the effect of seawater temperature on the uptake rates of *Z. marina* (DIN
253 and DON) and microbes (DON), plants and sediment cores (3 cm diameter, 5 cm depth)

254 were collected from the mid-latitude site (UK) and were incubated at the latitudinal
255 range of temperatures of 8, 12 and 17 °C. The rationale for this was that mid-latitude
256 plants are within the range of mean annual temperatures (8, 12 and 17 °C) of the three
257 latitudes studied, i.e. the minimum (8 °C) and maximum (17 °C) temperatures to which
258 mid-latitude plants are exposed coincide with the mean annual temperature of plants
259 from the high-latitude site in Iceland (8 °C) and from the lower-latitude site in Portugal
260 (17 °C). Plants and sediments were acclimated for three days to the experimental
261 conditions of light ($\sim 100 \mu\text{mol quanta m}^{-2} \text{ s}^{-1}$) and temperature under a 18:6 h
262 light:dark cycle. The uptake rates of inorganic (ammonium and nitrate) and organic
263 (alanine and trialanine) nitrogen by *Z. marina* and the microbial uptake of DON were
264 determined as described above. The average leaf biomass per incubated shoot was 0.24
265 g dry weight, whereas the average belowground biomass was 0.14 g dry weight.

266

267 Sample analysis

268 Concentrations of dissolved inorganic nitrogen (DIN) were determined colorimetrically
269 in a loop-flow analyzer ($\mu\text{MAC-1000}$ Systea, Agnani, Italy). Samples for determination
270 of total free amino acids and amino acids bound in short peptides were passed through a
271 1 kDa ultrafiltration membrane (Millipore, Billerica, Massachusetts, USA). Amino acid
272 N was determined fluorometrically before and after hydrolysis in 6 M HCl at 105 °C for
273 16 h under N_2 using glycine as a standard (Jones et al., 2002). Glycine was used because
274 it is the most predominant amino acid and its relative fluorescent intensity is similar to
275 that of other dominant amino acids (Parsons et al. 1984). Peptide N was determined as
276 the difference between the free and total amino acid pools, determined before and after
277 hydrolysis, respectively. Total nitrogen (TN) was determined after combustion to
278 nitrogen monoxide by chemiluminescence using a TOC-V-TN analyser (Shimadzu,

279 Kyoto, Japan). TN content and atom % ^{15}N of dried tissues were determined using a
280 PDZ Europa ANCA-GSL elemental analyzer interfaced to a PDZ Europa 20-20 isotope
281 ratio mass spectrometer (Sercon Ltd., Cheshire, UK). Precision of the $\delta^{15}\text{N}$ analysis was
282 higher than 0.03 ‰. Leaf and root ^{15}N background level was measured on five replicate
283 samples.

284

285 Data analysis

286 The N uptake rates of *Z. marina* were expressed in $\mu\text{mol g}^{-1}$ dry weight h^{-1} . ^{15}N
287 enrichment (%) of tissues after incubation was calculated by subtracting the post-
288 incubation ^{15}N levels from the initial background levels, multiplied by the total N
289 content (g) in the tissue and divided by its dry weight (g dry weight). Uptake rates were
290 plotted against substrate concentration (μM) and the uptake kinetic parameters were
291 derived using the Michaelis-Menten kinetic model

$$292 \quad V = (V_{\max} \times S) / (K_m + S)$$

293 where V is uptake rate ($\mu\text{mol g}^{-1}$ dry weight h^{-1}), V_{\max} is maximum uptake rate ($\mu\text{mol g}^{-1}$
294 DW h^{-1}), S is substrate concentration (μM) and K_m is the half-saturation constant (μM).
295 Data not displaying saturation kinetics were fitted to a linear regression model ($V =$
296 uptake rate, $S =$ substrate concentration).

297 Whole-plant nitrogen budgets per ground area ($\mu\text{mol m}^{-2} \text{h}^{-1}$) of *Z. marina* along the
298 latitudinal gradient and at each experimental temperature were estimated by multiplying
299 the N uptake rates ($\mu\text{mol g}^{-1}$ dry weight h^{-1}) of the plants at the ambient nutrient
300 concentrations (μM) measured in each site by the areal biomass (g^{-1} dry weight m^{-2}).

301 The DIN and DON contribution to the total N acquisition by the leaves and roots were
302 calculated by dividing the sum of the acquisition of ammonium and nitrate, or amino
303 acids and peptides, of each plant part by the total plant N acquisition.

304 Microbial DON uptake rates were expressed in $\text{nmol mL}^{-1} \text{h}^{-1}$. ^{14}C microbial uptake of
305 alanine and trialanine was calculated based on the percentage of ^{14}C -labelled solution
306 taken up by the microbes, obtained from the proportion of initial ^{14}C activity remaining
307 in solution after incubation.

308 Differences in ambient concentrations between study sites were tested for each N source
309 separately using one-way analysis of variance (ANOVA). The effects of latitude /
310 nutrient concentration and temperature / nutrient concentration on N uptake rates by *Z.*
311 *marina* were tested using two-way ANOVAs for each N source and plant part. All other
312 differences in *Z. marina* uptake rates were tested using the non-parametric Kruskal-
313 Wallis test because the assumption of data normality was not met even after data
314 transformation.

315 The effects of latitude / trialanine concentration and of temperature / alanine
316 concentration on microbial uptake rates were tested using two-way ANOVA. The
317 effects of latitude / alanine concentration and of temperature / trialanine concentration
318 on microbial uptake rates were tested with the non-parametric Kruskal-Wallis test
319 because the ANOVA assumptions of data normality and equal variance were not met
320 even after data transformation. Data were square root transformed when normality was
321 not verified. All tests were performed at a level of significance of $p < 0.05$.

322

323 Results

324

325 Concentration of inorganic and organic N

326 The concentrations of the different forms of soluble N were generally much higher in
327 the sediment porewater than in the water column at all sites (Table 1). The nutrient
328 concentrations in the water column did not vary significantly among sites except for

329 ammonium and amino acids, which were higher in Portugal and in the UK, respectively.
330 The concentration of ammonium in the sediment porewater was not significantly
331 different among the study sites, whereas the concentration of all other N sources varied
332 significantly but with no consistent latitudinal pattern.
333 Amino acids and peptides, a fraction of the dissolved organic nitrogen (DON) pool,
334 constituted an important component of the total nitrogen available in both the sediment
335 and water column of the studied sites, representing 53%, 29% and 43% of the total
336 nitrogen in the sediment and 52%, 13% and 41% in the water, respectively in Iceland,
337 UK and Portugal. No latitudinal consistency was found.

338

339 Plant density and biomass

340 Shoot density in Iceland (1483 ± 379 shoots m^{-2}) was much higher than in the UK (533
341 ± 113 shoots m^{-2}) and Portugal (408 ± 52 shoots m^{-2}) but the aboveground biomass was
342 lower at the northern limit of the species distribution (44 ± 11 g dry weight m^{-2})
343 compared to the UK (185 ± 12 g dry weight m^{-2}) and Portugal (130 ± 9 g DW m^{-2}) due
344 to the smaller size of the leaves (leaf length: 15 ± 4 cm in Iceland, 37 ± 10 cm in the UK
345 and 32 ± 9 cm in Portugal). Below-ground biomass was conservative: in Iceland it was
346 55 ± 5 g dry weight m^{-2} , in the UK it was 56 ± 12 g dry weight m^{-2} and in Portugal it
347 was 55 ± 5 g dry weight m^{-2} . *Z. marina* from Iceland showed higher concentrations of
348 nitrogen in the leaf tissues (2.35 ± 0.2 % dry weight) compared to plants from the UK
349 and Portugal (1.65 ± 0.4 and 1.83 ± 0.2 % DW, respectively).

350

351 N uptake by *Zostera marina*

352 Overall, the DIN and DON uptake rates by *Z. marina* was higher towards the equator,
353 and reduced in the poleward direction, particularly at higher substrate concentrations

354 (Fig. 1, Table S2). *Z. marina* growing at the southern limit of its distribution (Portugal)
355 showed significantly higher uptake rates compared to plants growing further north
356 except for the uptake of ammonium through the roots, which was similar between
357 latitudes at all nutrient concentrations (Fig. 1e, Table S2). Differences in the uptake
358 rates between latitudes at the lowest N concentration (5 μ M) were only detected for
359 organic N sources (alanine and trialanine). Maximum nitrogen (inorganic and alanine)
360 uptake rates (V_{\max}) consistently increased with decreasing latitude, with a 2 to 4-fold
361 decrease from Portugal to Iceland (Table S3). The latitudinal effects on nutrient affinity
362 (α) and on the half-saturation constant (K_m) were not as clear. The increase of V_{\max} with
363 decreasing latitude is mostly caused by temperature, as indicated by the temperature
364 experiment, where V_{\max} increased significantly with temperature (Table S4). The results
365 of the two-way analysis of variance examining the effects of temperature (T) and
366 nutrient concentration (N) on the nitrogen uptake rates of *Zostera marina* support that
367 the latitudinal pattern observed is caused by temperature as higher temperatures resulted
368 in higher N uptake (Fig. 2, Table S5).

369

370 Whole-plant N budget

371 The leaf, root and whole-plant N budgets of *Z. marina* increased with decreasing
372 latitude both for DIN and DON (Fig. 3a, Table S6). Root uptake was more important for
373 *Z. marina* nitrogen budget than leaf uptake as it contributed 93%, 72% and 70% to the
374 whole-plant budget, respectively in Iceland, UK and Portugal (Fig. 3b, Table S6). These
375 values also show that root contribution decreases with decreasing latitude. Overall, the
376 uptake of DON by *Z. marina* contributed about one third to the whole-plant nitrogen
377 budget, showing that this N form is ecologically relevant throughout the latitudinal
378 gradient. Ammonium represented the largest contributor to the total DIN budget of the

379 species at all sites, particularly via root uptake from the sediment (66% in Iceland, 44%
380 in UK and 41% in Portugal, Table S6). The contribution of amino acids and peptides
381 via root uptake (26 % in Iceland, 27% in UK and 24% in Portugal) was smaller
382 compared to ammonium but much higher than that of nitrate (2, 0.5 and 5%,
383 respectively).

384 The temperature experiments showed a general pattern of increasing DIN and DON
385 budgets towards warmer sites, except for the DIN and DON of leaves (Fig. 3c, Table
386 S7). The relative contributions of DIN and DON to the whole-plant budgets did not
387 vary with latitude, but the relative contribution of DON to the whole plant N budget
388 increased with temperature, as opposed to DIN (Fig. 3d, Table S7).

389

390 Microbial DON uptake

391 The sediment microbial uptake of alanine did not change significantly among sites ($H =$
392 4.491, $p = 0.106$), as opposed to the uptake of trialanine that increased with decreasing
393 latitude (Fig. 4a and b). Despite the lack of a significant latitudinal variation of alanine,
394 the temperature experiments showed significant effects on the microbial uptake rates of
395 alanine at the highest nutrient concentration (100 μM) ($F = 20.46$, $P < 0.001$) (Fig. 4c).
396 The microbial uptake of trialanine increased significantly from 8 °C to 12 °C and 17 °C
397 ($H = 6.423$, $P = 0.04$) (Fig. 4d).

398

399 Discussion

400

401 Our study showed that the uptake rates of inorganic and organic N by both leaves and
402 roots and the total meadow uptake under local conditions (N budget) of the globally
403 widespread seagrass *Zostera marina* increased from northern to southern sites. The

404 hypothesis that temperature is the main driver of this latitudinal trend was supported by
405 the temperature experiments.

406

407 N uptake rates and N budget of *Z. marina*

408 For all N sources, the uptake rates and the uptake kinetics of *Z. marina* increased with
409 temperature as reported for the uptake rates of ammonium and glycine by the Arctic
410 saltmarsh grass *Puccinellia phryganodes* (Henry & Jefferies, 2003). Kaldy (2014) is, to
411 our knowledge, the only other report that has experimentally tested the effects of
412 temperature on the acquisition of nitrogen by *Z. marina*. This study showed a 50 %
413 increase in nitrate uptake as the temperature increased from 10 to 18/25 °C in plants
414 exposed to elevated nitrate concentrations (100 µM). Indirect indications of temperature
415 effects on the acquisition of nitrogen by seagrasses have been presented in studies
416 evaluating the seasonal variation of N uptake rates (Lee & Dunton, 1999; Hasegawa et
417 al., 2005) or the influence of season and temperature on the activity of enzymes
418 involved in the nitrogen metabolism, such as glutamine synthetase (Kraemer &
419 Mazzella, 1999) and nitrate reductase (Alexandre et al., 2004). In these studies, the
420 uptake rates and the enzymatic activities generally increased with increasing
421 temperature. Despite the greater amount of bioavailable DON relative to DIN in all
422 studied sites, the uptake of DIN represented the largest fraction of the total N budget of
423 *Z. marina* across its latitudinal distribution. This was due to the high uptake of
424 ammonium (54 - 68 % of the total N uptake), which is the preferential nitrogen source
425 of *Z. marina* (Short & McRoy, 1984; Hemminga et al., 1994; Alexandre et al., 2015).
426 Even though the rate of nitrogen uptake through the leaves was generally higher than
427 through the roots at similar nutrient concentrations, the overall contribution of root
428 uptake to the total N budget of *Z. marina* (70 - 94 %) was much higher than that of

429 leaves (7 - 30 %) because the concentrations of the different nitrogen sources in the
430 sediment porewater were several-fold higher than in the water column. This relative
431 contribution of the different plant parts contrasts with other reports showing an equal
432 contribution of leaves and roots to the total annual N acquisition by *Z. marina* (Pedersen
433 & Borum, 1993; Lee & Dunton, 1999), but their calculations accounted only for
434 inorganic nitrogen. Amino acids and oligopeptides are a relevant source of nitrogen for
435 *Z. marina*, particularly through root uptake. The average contribution of amino acids
436 (12 %) and peptides (14 %) via root uptake exceeded that of nitrate (2 %) by several-
437 fold at the extremes of the latitudinal gradient. This result supports the relevant role of
438 organic nitrogen as a complementary N source for *Z. marina*, previously reported by
439 Alexandre et al. (2015).

440 In contrast to our initial hypothesis, the pool of bioavailable DON did not increase with
441 latitude driven by lower microbial N mineralisation and this was not reflected in higher
442 DON uptake by *Z. marina*, as observed for arctic and antarctic vascular plants (Chapin
443 et al., 1993, Hill et al., 2011). Both the uptake of DON by *Z. marina* and the DON
444 contribution to the whole-plant N budget increased towards lower latitude and this trend
445 was driven by higher temperature. At high latitude, the lower contribution of DON to
446 the total N budget of *Z. marina* is determined by the plants' lower DON uptake rates
447 rather than by a lower availability of DON sources.

448

449 Microbial DON uptake

450 The microbial uptake of DON in *Z. marina* sediments increased towards southern
451 latitudes and this was driven by temperature. DON uptake is ecologically relevant as
452 amino acids and peptides accounted for a significant pool of bioavailable nitrogen in *Z.*
453 *marina* meadows, particularly in the sediments (30 to 50 % of the total N). The

454 observed latitudinal pattern contrasts with that reported by Mozdzer et al. (2014), who
455 found significant increases in sediment DON availability with increasing latitude in
456 *Spartina alterniflora* saltmarshes, which coincided with a significant decrease in
457 microbial DON uptake.

458 To the best of our knowledge, this was the first report of the latitudinal effects of
459 temperature on the DON uptake of both microbes and seagrasses, which is relevant to
460 understand their competition for these nitrogen sources. Dissolved organic nitrogen
461 serves both as a direct source of nitrogen for *Z. marina* and as a mineralisation substrate
462 for its associated microbial community, and both of their uptake rates increase with
463 temperature. Seagrasses in general and *Z. marina* in particular may have a competitive
464 advantage over microbes for DON uptake because they also acquire nutrients through
465 the leaves from the water column, where microbial uptake of alanine and trialanine was
466 very low, according to the lower ^{14}C uptake when compared to the sediment.

467 The northern distribution limits of *Z. marina* are warming twice as fast as the rest of the
468 globe (IPCC, 2018) creating the conditions for its expansion towards the pole. This
469 warming trend will increase the nitrogen demand of *Z. marina* that may be met by an
470 increasing uptake of organic nitrogen forms. The competition with microbes for this
471 nutrient source is also expected to increase as warming will also enhance the microbial
472 uptake of DON. Our results highlight that *Z. marina*, and probably other seagrass
473 species, can be winners under global change conditions of increased temperature and
474 CO_2 . Many reports have shown that higher CO_2 levels will increase the photosynthetic
475 production of seagrasses (Jiang et al., 2010, Alexandre et al., 2012, Ow et al., 2015),
476 which may result in higher growth that will not be limited by the plants' nitrogen uptake
477 capacity.

478

479 Acknowledgements

480

481 We thank J. Svavarsson and E. Olafsson for their support at the University of Iceland,
482 C.B. de los Santos for sharing the geographical coordinates of the site in Iceland, and S.
483 Chesworth for the analysis of total nitrogen at Bangor University. We also thank two
484 anonymous reviewers for their constructive comments on the manuscript. A.A. was
485 supported by a postdoctoral fellowship from the Portuguese Foundation for Science and
486 Technology (FCT, SFRH/BPD/91629/2012). This study received Portuguese national
487 funds from FCT - Foundation for Science and Technology through project
488 UIDB/04326/2020, and in the ambit of the contract program DL
489 57/2016/CP1361/CT0001.

490

491 Authors contribution

492

493 A.A. and R.S. conceived and designed the study. A.A., R.Q. and P.W.H. carried out the
494 experimental work. A.A., P.W.H., D.J. and R.S. analysed the data. A.A. wrote the
495 manuscript. All authors contributed critically to the drafts and gave final approval for
496 publication.

497

498 Data availability statement

499

500 Data are available from the Mendeley Data under doi: 10.17632/r4m88s8m8w.1
501 (Alexandre and Santos 2020 Zm N uptake latitude and temperature).

502

503 References

504

505 Agawin, N. S. R., Duarte, C. M. & Fortes, M. D. (1996). Nutrient limitation of
506 Philippine seagrasses (Cape Bolinao, NW Philippines): in situ experimental evidence.
507 Marine Ecology Progress Series, 138, 233–243. doi:<http://hdl.handle.net/10261/149956>

508

509 Alcoverro, T., Romero, J., Duarte, C. M. & López, N.I. (1997). Spatial and temporal
510 variations in nutrient limitation of seagrass *Posidonia oceanica* growth in the NW
511 Mediterranean. Marine Ecology Progress Series, 146, 155-161. doi:

512 [10.3354/meps146155](https://doi.org/10.3354/meps146155)

513

514 Alexandre, A., Silva, J. & Santos, R. (2004). The maximum nitrate reductase activity of
515 the seagrass *Zostera noltii* (Hornem.) varies along its vertical distribution. Journal of
516 Experimental Marine Biology and Ecology, 307, 127–135. doi:

517 <https://doi.org/10.1016/j.jembe.2004.02.002>

518

519 Alexandre, A., Silva, J. & Santos, R. (2010). Inorganic nitrogen uptake and related
520 enzymatic activity in the seagrass *Zostera noltii*. Marine Ecology, 31, 539-545. doi:

521 [10.1111/j.1439-0485.2010.00378.x](https://doi.org/10.1111/j.1439-0485.2010.00378.x)

522

523 Alexandre, A., Silva, J., Bouma, T. J. & Santos, R. (2011). Inorganic nitrogen uptake
524 kinetics and whole-plant nitrogen budget in the seagrass *Zostera noltii*. Journal of
525 Experimental Marine Biology and Ecology, 401, 7-12. doi:

526 [10.1016/j.jembe.2011.03.008](https://doi.org/10.1016/j.jembe.2011.03.008)

527

- 528 Alexandre, A., Silva, J., Buapet, P., Björk, M. & Santos, R. (2012). Effects of CO₂
529 enrichment on photosynthesis, growth, and nitrogen metabolism of the seagrass *Zostera*
530 *noltii*. *Ecology and Evolution*, 2, 2625–2635. doi: 10.1002/ece3.333
531
- 532 Alexandre, A., Hill, P. W., Jones, D. L. & Santos, R. (2015). Dissolved organic
533 nitrogen: a relevant, complementary source of nitrogen for the seagrass *Zostera marina*.
534 *Limnology and Oceanography*, 60, 1477-1483. doi:10.1002/lno.10084
535
- 536 Alexandre, A., Baeta, A., Engelen, A. H. & Santos, R. (2017) Interactions between
537 seagrasses and seaweeds during surge nitrogen acquisition determine interspecific
538 competition. *Scientific Reports*, 7, 13651. doi:10.1038/s41598-017-13962-4
539
- 540 Aronson, E. L. & McNulty, S. G. (2009). Appropriate experimental ecosystem warming
541 methods by ecosystem, objective, and practicality. *Agricultural and Forest Meteorology*,
542 149, 1791–1799. doi: <https://doi.org/10.1016/j.agrformet.2009.06.007>
543
- 544 Bai, E., Li, S., Xu, W., Li, W., Dai, W. & Jiang, P. (2013). A meta-analysis of
545 experimental warming effects on terrestrial nitrogen pools and dynamics. *New*
546 *Phytologist*, 199, 441–445. doi: 10.1111/nph.12252
547
- 548 BassiriRad, H. (2000). Kinetics of nutrient uptake by roots: responses to global change.
549 *New Phytologist*, 147, 155-169. doi: <https://doi.org/10.1046/j.1469-8137.2000.00682.x>
550
- 551 Bauwe, H., Hagemann, M. & Fernie, A. R. (2010). Photorespiration: players, partners
552 and origin. *Trends in Plant Science*, 15, 330-336. doi:10.1016/j.tplants.2010.03.006

553

554 Bostrom, C., Jackson, E. L. & Simenstad, C. A. (2006). Seagrass landscapes and their
555 effects on associated fauna: A review. *Estuarine Coastal and Shelf Science*, 68, 383-
556 403. doi: <https://doi.org/10.1016/j.ecss.2006.01.026>

557

558 Brailsford, F. L., Glanville, H. C., Golyshin, P. N., Johnes, P. J., Yates, C. A. & Jones,
559 D. L. (2019). Microbial uptake kinetics of dissolved organic carbon (DOC) compound
560 groups from river water and sediments. *Scientific Reports*, 9, 11229.
561 doi:<https://doi.org/10.1038/s41598-019-47749-6>

562

563 Bronk, D. A., See, J. H., Bradley, P. & Killberg, L. (2007). DON as a source of
564 bioavailable nitrogen for phytoplankton. *Biogeosciences*, 4, 283–296.
565 doi:<https://doi.org/10.5194/bg-4-283-2007>

566

567 Cabello-Pasini, A., Muniz-Salazar, R. & Ward, D. H. (2003). Annual variations of
568 biomass and photosynthesis in *Zostera marina* at its southern end of distribution in the
569 North Pacific. *Aquatic Botany*, 76, 31–47. doi:[https://doi.org/10.1016/S0304-](https://doi.org/10.1016/S0304-3770(03)00012-3)
570 [3770\(03\)00012-3](https://doi.org/10.1016/S0304-3770(03)00012-3)

571

572 Chapin, F. S. III, Moilanen, L. & Kielland, K. (1993). Preferential use of organic
573 nitrogen for growth by a non-mycorrhizal arctic sedge. *Nature*, 361, 150-153.
574 doi:<https://doi.org/10.1038/361150a0>

575

- 576 Chefaoui, R. M., Duarte, C. M. & Serrao, E. A. (2018). Dramatic loss of seagrass
577 habitat under projected climate change in the Mediterranean Sea. *Global Change*
578 *Biology*, 24, 4919-4928. doi:<https://doi.org/10.1111/gcb.14401>
579
580 Climate-data.org, <https://en.climate-data.org/continent/europe/>
581
- 582 De Frenne, P., Graae, B. J., Rodríguez-Sánchez, R., Kolb, A., Chabrierie, O., Decocq,
583 G., ... Verheyen, K. (2013). Latitudinal gradients as natural laboratories to infer
584 species' responses to temperature. *Journal of Ecology*, 101, 784-795.
585 doi:<https://doi.org/10.1111/1365-2745.12074>
586
- 587 Duarte, B., Martins, I., Rosa, R., Matos, A. R., Roleda, M. Y., Reusch, T. B. H., ...
588 Jueterbock, A. (2018). Climate change impacts on seagrass meadows and macroalgal
589 forests: an integrative perspective on acclimation and adaptation potential. *Frontiers in*
590 *Marine Science*, 5, UNSP 190. doi:<https://doi.org/10.3389/fmars.2018.00190>
591
- 592 Fukami, T. & Wardle, D. A. (2005). Long-term ecological dynamics: reciprocal
593 insights from natural and anthropogenic gradients. *Proceedings of the Royal Society B*,
594 272, 2105-2115. doi:10.1098/rspb.2005.3277
595
- 596 Global Sea Temperature, [https://www.seatemperature.org/europe/united-](https://www.seatemperature.org/europe/united-kingdom/llandudno.htm)
597 [kingdom/llandudno.htm](https://www.seatemperature.org/europe/united-kingdom/llandudno.htm)
598
- 599 Green, E. P. & Short, F.T. (2003). *World atlas of seagrasses*. University of California
600 Press.

601

602 Hasegawa, N., Iizumi, H. & Mukai, H. (2005). Nitrogen dynamics of the surfgrass

603 *Phyllospadix iwatensis*. Marine Ecology Progress Series, 293, 59–68.

604 doi:10.3354/meps293059

605

606 Hemminga, M. A., Koutstaal, B. P., Van Soelen, J. A. & Merks, G. A. (1994). The

607 nitrogen supply to intertidal eelgrass (*Zostera marina*). Marine Biology, 118, 223-227.

608 doi:<https://doi.org/10.1007/BF00349788>

609

610 Henry, H. A. L. & Jefferies, R. L. (2003). Interactions in the uptake of amino acids,

611 ammonium and nitrate ions in the Arctic salt-marsh grass, *Puccinellia phryganodes*.

612 Plant Cell & Environment, 26, 419-428. doi:10.1046/j.1365-3040.2003.00973.x

613

614 Hill, P. W., Farrar, J., Roberts, P., Farrel, M., Grant, H., Newsham, K. K., ... Jones, D.

615 L. (2011). Vascular plant success in a warming Antarctic may be due to efficient

616 nitrogen acquisition. Nature Climate Change, 1, 50-53. doi:10.1038/nclimate1060

617

618 IPCC (Intergovernmental Panel on Climate Change) (2018). Global warming of 1.5 °C

619 Summary for Policymakers, 33 pp. http://report.ipcc.ch/sr15/pdf/sr15_spm_final.pdf

620

621 Jiang, Z. J., Huang, X. P. & Zhang, J. P. (2010). Effects of CO₂ enrichment on

622 photosynthesis, growth and biochemical composition of seagrass *Thalassia hemprichii*

623 (Ehrenb.) Aschers. Journal of Integrative Plant Biology, 52, 904–913.

624 <https://doi.org/10.1111/j.1744-7909.2010.00991.x>

625

- 626 Jonasson, S., Michelsen, A., Schmidt, I. K. & Nielsen, E. V. (1999). Responses in
627 microbes and plants to changed temperature, nutrient, and light regimes in the arctic.
628 Ecology, 80, 1828–1843. doi:10.2307/176661
629
- 630 Jones, D. L., Owen, A. G. & Farrar, J. F. (2002). Simple method to enable the high
631 resolution determination of total free amino acids in soil solutions and soil extracts. Soil
632 Biology & Biochemistry, 34, 1893–1902. doi:https://doi.org/10.1016/S0038-
633 0717(02)00203-1
634
- 635 Kaldy, J. E. (2014). Effect of temperature and nutrient manipulations on eelgrass
636 *Zostera marina* L. from the Pacific Northwest, USA. Journal of Experimental Marine
637 Biology and Ecology, 453, 108-115. doi:https://doi.org/10.1016/j.jembe.2013.12.020
638
- 639 Kraemer, G. P., Mazzella, L. (1999). Nitrogen acquisition, storage, and use by the co-
640 occurring Mediterranean seagrasses *Cymodocea nodosa* and *Zostera noltii*. Marine
641 Ecology Progress Series, 183, 95-103. doi:10.3354/meps183095
642
- 643 Lee, K. S., Dunton, K. H. (1999). Inorganic nitrogen acquisition in the seagrass
644 *Thalassia testudinum*: development of a whole-plant nitrogen budget. Limnology &
645 Oceanography, 44, 1204-1215. doi:https://doi.org/10.4319/lo.1999.44.5.1204
646
- 647 Moore, K. A. & Short, F. T. (2006). Comparative biology of *Zostera*. In: A. W. D.
648 Larkum, R. J. Orth & C. M. Duarte (Eds.) Seagrasses: biology, ecology and
649 conservation (pp. 361-386). The Netherlands, Springer.
650

- 651 Marine and Freshwater Research Institute
652 (http://www.hafro.is/undir_eng.php?REFID=11&ID=174&REF=2)
653
- 654 Mozdzer, T. J., McGlathery, K. J., Mills, A. L. & Zieman, J. C. (2014). Latitudinal
655 variation in the availability and use of dissolved organic nitrogen in Atlantic coast salt
656 marshes. *Ecology*, 95, 3293–3303. doi:<https://doi.org/10.1890/13-1823.1>
657
- 658 Näsholm, T., Kielland, K. & Ganeteg, U. (2009). Uptake of organic nitrogen by plants.
659 *New Phytologist*, 182, 31–48. doi:10.1111/j.1469-8137.2008.02751.x
660
- 661 Ow, Y. X., Collier, C. J. & Uthicke, S. (2015). Responses of three tropical seagrass
662 species to CO₂ enrichment. *Marine Biology*, 162, 1005–1017. doi: 10.1007/s00227-
663 015-2644-6
664
- 665 Parsons, T., Maita & Y., Lalli, C. (1984). A manual of chemical and biological methods
666 for seawater analysis. Oxford. Pergamon Press.
667
- 668 Pedersen, M. F. & Borum, J. (1993). An annual nitrogen budget for a seagrass *Zostera*
669 *marina* population. *Marine Ecology Progress Series*, 101, 169-177.
670 doi:10.3354/meps101169
671
- 672 Poloczanska, E. S., Brown, C. J., Sydeman, W. J., Kiessling, W., Schoeman, D. S.,
673 Moore, P. J., ... Richardson, A. J. (2013). Global imprint of climate change on marine
674 life. *Nature Climate Change*, 3, 919–925. doi:<https://doi.org/10.1038/nclimate1958>
675

- 676 Rustad, L. E., Campbell, J. L., Marion, G. M., Norby, R. J., Mitchell, M. J., Hartley, A.
677 E., ... Gurevitch, J. (2001). A meta-analysis of the response of soil respiration, net
678 nitrogen mineralization, and aboveground plant growth to experimental ecosystem
679 warming. *Oecologia*, 126, 543–562. doi:<https://doi.org/10.1007/s004420000544>
680
- 681 Rustad, L. E. (2008). The response of terrestrial ecosystems to global climate change:
682 towards an integrated approach. *Science of the Total Environment*, 404, 222-235.
683 doi:<https://doi.org/10.1016/j.scitotenv.2008.04.050>
684
- 685 Sharkey, T. D. (2007). Estimating the rate of photorespiration in leaves. *Physiologia*
686 *Plantarum*, 73, 147-152. doi:10.1111/j.1399-3054.1988.tb09205.x
687
- 688 Sharp, J. H. (1983). The distributions of inorganic nitrogen and dissolved and
689 particulate organic nitrogen in the sea. In: E. J. Carpenter & D. G. Capone (Eds.),
690 *Nitrogen in the Marine Environment* (pp. 1-35). New York, NY: Academic Press.
691
- 692 Sharp, J. H., Rinker, K. R., Savidge, K. B., Abell, J., Benaim, J. Y., Bronk, D., ...
693 Wong, C. S. (2002). A preliminary methods comparison for measurement of dissolved
694 organic nitrogen in seawater. *Marine Chemistry*, 78, 171– 184.
695 doi:[https://doi.org/10.1016/S0304-4203\(02\)00020-8](https://doi.org/10.1016/S0304-4203(02)00020-8)
696
- 697 Shaver, G. R., Canadell, J., Chapin III, F. S., Gurevich, J., Harte, J., Henry, G., ...
698 Rustad, L. (2000). Global warming and terrestrial ecosystems: a conceptual framework
699 for analysis. *Bioscience*, 50, 871-882. doi:[https://doi.org/10.1641/0006-3568\(2000\)050\[0871:GWATEA\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2000)050[0871:GWATEA]2.0.CO;2)
700

701

702 Short, F.T. & McRoy, C. P. (1984). Nitrogen uptake by leaves and roots of the seagrass

703 *Zostera marina* L. *Botanica Marina*, 27, 547–555. doi:10.1515/botm.1984.27.12.547

704

705 Van Engeland, T., Bouma, T. J., Morris, E. P., Brun, F. G., Peralta, G., Lara, M., ...

706 Middelburg, J. J. (2011). Potential uptake of dissolved organic matter by seagrasses and

707 macroalgae. *Marine Ecology Progress Series*, 427, 71-81. doi:10.3354/meps09054

708

709 Vonk, J. A., Middelburg, J. J., Stapel, J. & Bouma, T. J. (2008). Dissolved organic

710 nitrogen uptake by seagrasses. *Limnology & Oceanography*, 53, 542-548.

711 doi:10.2307/40006439

712

713 Walther, G. R., Post, E., Convey, P., Menzel, A., Parmesan, C., Beebee, T. J. C., ...

714 Bairlein, F. (2002). Ecological responses to recent climate change. *Nature*, 416, 389-715 395. doi:<https://doi.org/10.1038/416389a>

716

717 SUPPORTING INFORMATION

718 Additional supporting information may be found in the online version of this article.

719

720 Table S1. Average aboveground and belowground biomass of *Zostera marina* incubated

721 in each site along the latitudinal distribution gradient.

722 Table S2. Nitrogen uptake kinetic parameters of *Zostera marina* along the species'

723 latitudinal distribution gradient.

724 Table S3. Results of the two-way ANOVA examining the effects of latitude and nutrient

725 concentration on the nitrogen uptake rates of *Zostera marina*.

726 Table S4. Nitrogen uptake kinetic parameters of *Zostera marina* at three different
727 temperatures.

728 Table S5. Results of the two-way ANOVA examining the effects of temperature and
729 nutrient concentration on the nitrogen uptake rates of *Zostera marina*.

730 Table S6. Whole-plant nitrogen budget of *Zostera marina* along the species latitudinal
731 distribution range in Europe.

732 Table S7. Whole-plant nitrogen budget of *Zostera marina* at three different
733 temperatures.

734 Figure S1. Aspect of the *Zostera marina* meadows in Iceland, United Kingdom and
735 Portugal.

736

737

738

739

740

741

742

743 Table 1. Concentration (μM) of the different nitrogen sources (ammonium, nitrate, free
744 amino acids, peptides and total nitrogen) in the water column and sediment porewater in
745 the study sites (Iceland, United Kingdom and Portugal). Values are mean \pm S.D. (n = 5).
746 Levels of significance of the statistical tests are indicated as (*) $P < 0.05$, (**) $P < 0.01$,
747 (***) $P < 0.001$, (^{ns}) not significant. Different letters indicate significant differences.

748 Peptide values in *italic* are not reliable as they are higher than total nitrogen and higher
749 than sediment values (see text in Supporting Information).

750

	Iceland	United Kingdom	Portugal	Test statistic
<i>Water column</i>				
Ammonium	0.18 ± 0.19 ^a	0.77 ± 0.36 ^a	1.35 ± 1.06 ^b	H = 7.322*
Nitrate	0.43 ± 0.68	0.14 ± 0.06	3.12 ± 1.89	H = 4.842 ^{ns}
Amino acids	0.29 ± 0.10 ^a	0.67 ± 0.30 ^b	0.29 ± 0.11 ^a	F = 6.483*
Peptides (estimated) [§]	2.50 ± 1.83	1.03 ± 0.49	4.82 ± 4.04	H = 5.700 ^{ns}
Total nitrogen	5.33 ± 3.90	12.86 ± 6.14	12.36 ± 10.35	F = 2.784 ^{ns}
<i>Sediment</i>				
Ammonium	20.87 ± 8.49	20.03 ± 8.98	30.17 ± 6.75	F = 1.769 ^{ns}
Nitrate	0.66 ± 0.55 ^a	0.54 ± 0.16 ^a	15.08 ± 5.09 ^b	F = 26.201***
Amino acids	7.83 ± 4.25 ^a	62.25 ± 20.06 ^b	6.72 ± 2.12 ^a	H = 7.385*
Peptides	42.23 ± 16.43 ^{ab}	17.70 ± 13.33 ^a	50.59 ± 12.59 ^b	F = 5.081*
Total nitrogen	93.93 ± 29.35 ^a	278.73 ± 102.57 ^b	134.72 ± 33.49 ^{ab}	H = 9.118***

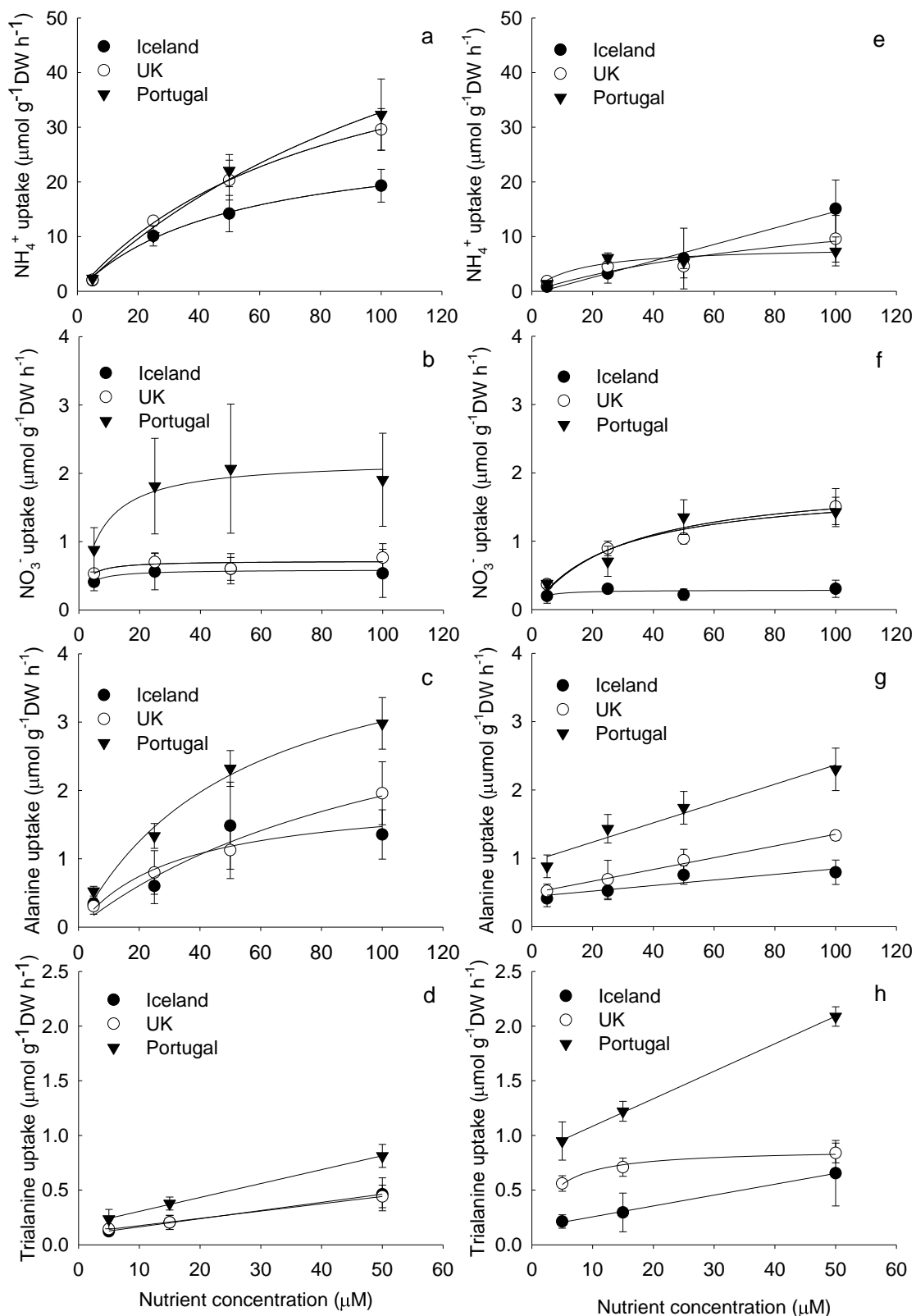
751 [§]Concentrations estimated from the percentage of peptides relative to the total soluble

752 nitrogen in the sediment porewater (see text).

753

754 Figure 1. Uptake rates ($\mu\text{mol g}^{-1}$ dry weight h^{-1}) of the different nitrogen sources by
755 leaves and roots of *Zostera marina* as a function of concentration (μM) along the
756 species latitudinal distribution gradient (Iceland, United Kingdom and Portugal):
757 ammonium (a), nitrate (b), alanine (c) and trialanine (d) by the leaves; ammonium (e),
758 nitrate (f), alanine (g) and trialanine (h) by the roots. Values are mean \pm S.D. (n = 3).
759 DW = dry weight.

760



761

762

763

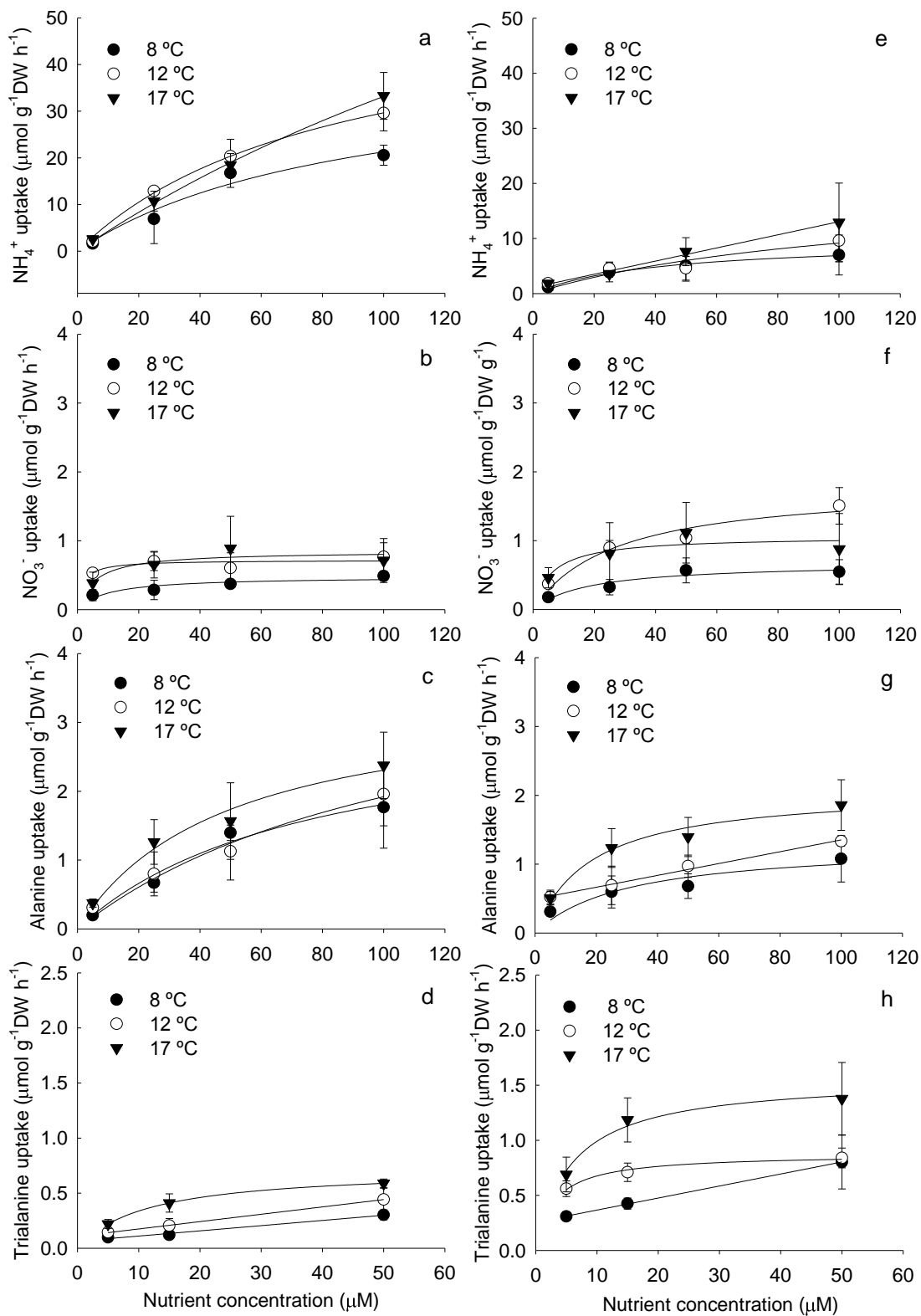
764

765

766

767

768 Figure 2. Uptake rates ($\mu\text{mol g}^{-1}$ dry weight h^{-1}) of the different nitrogen sources by
769 leaves and roots of *Zostera marina* as a function of concentration (μM) at three
770 incubation temperatures (8, 12 and 17 °C): ammonium (a), nitrate (b), alanine (c) and
771 trialanine (d) by the leaves; ammonium (e), nitrate (f), alanine (g) and trialanine (h) by
772 the roots. Values are mean \pm S.D. (n = 3). DW = dry weight.



773

774

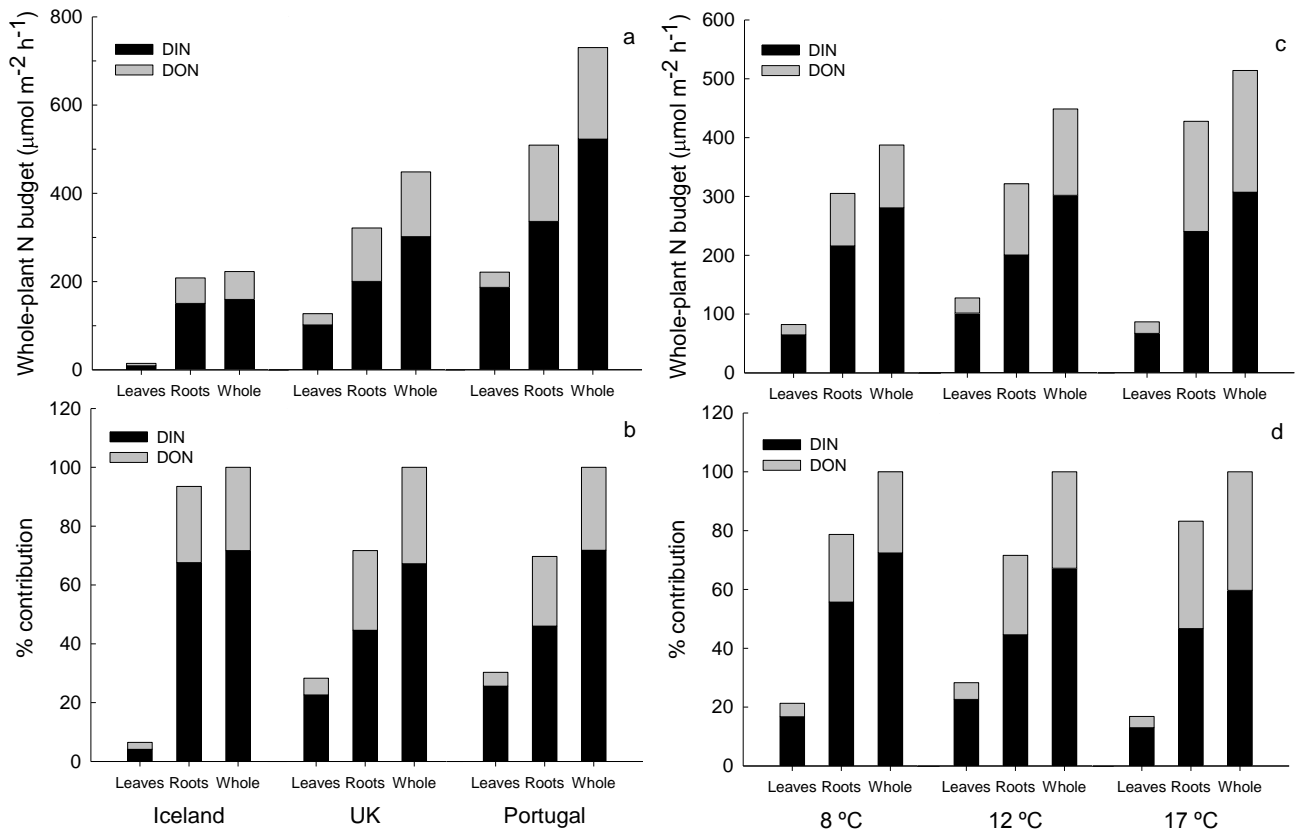
775

776

777

778

779 Figure 3. Whole-plant nitrogen budget of *Zostera marina* along the species latitudinal
 780 distribution range in Europe (Iceland, United Kingdom and Portugal) as $\mu\text{mol m}^{-2} \text{h}^{-1}$ (a)
 781 and % contribution (b), and the experimental effects of temperature on the whole-plant
 782 nitrogen budget as $\mu\text{mol m}^{-2} \text{h}^{-1}$ (c) and % contribution (d).
 783



784

785

786

787

788

789

790

791

792

793

794

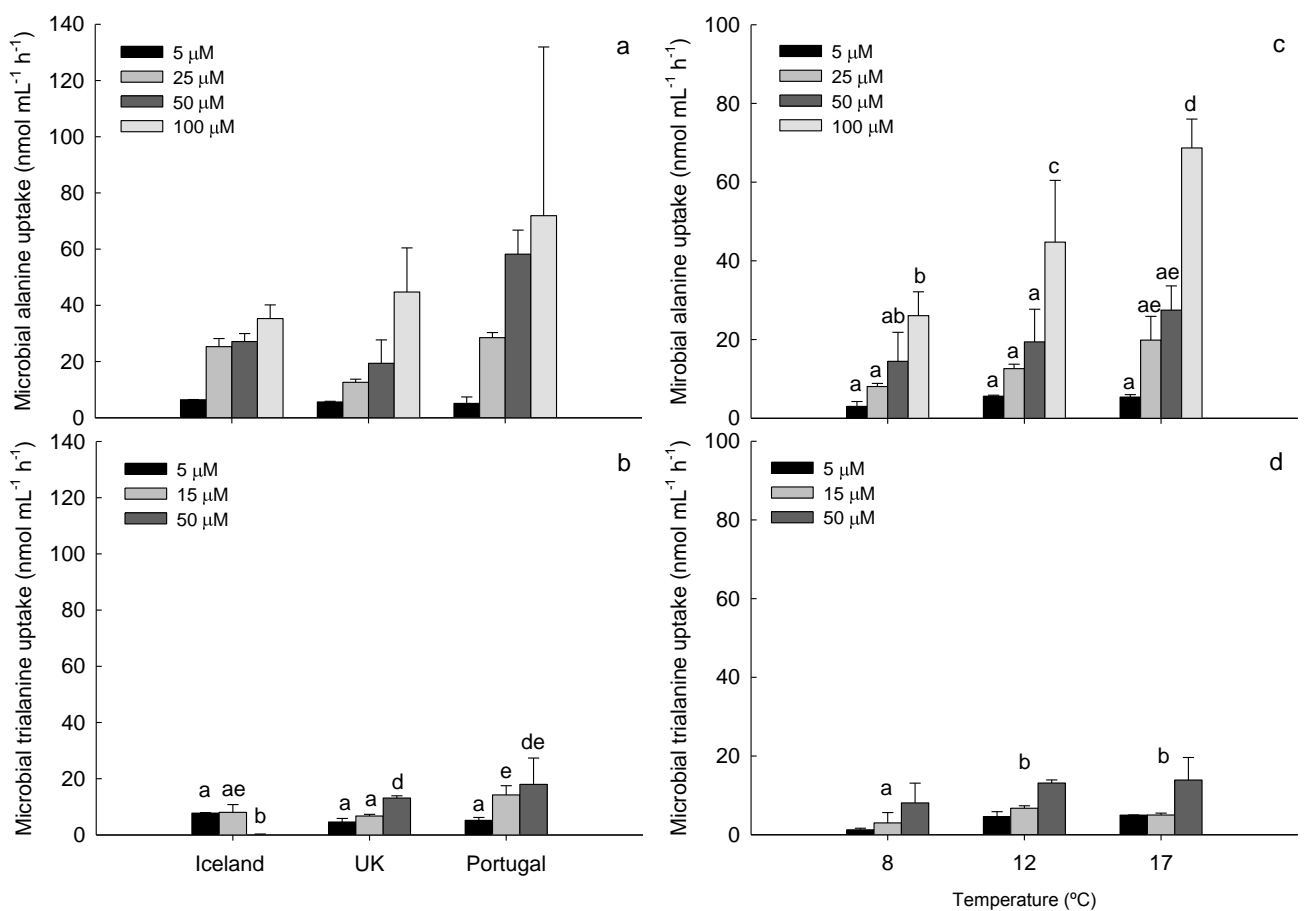
795

796

797

798 Figure 4. Microbial uptake ($\text{nmol mL}^{-1} \text{h}^{-1}$) of alanine (a) and trialanine (b) as a function
 799 of substrate concentration (μM) in *Zostera marina* meadows at three sites (Iceland,
 800 United Kingdom and Portugal) along the species latitudinal distribution gradient, and
 801 the experimental effects of temperature on the microbial uptake of alanine (c) and
 802 trialanine (d). Values are mean \pm SD ($n = 3$). Significant differences are indicated by
 803 different letters. ($P < 0.05$).

804



805

806

807

808

809

810

811 SUPPORTING INFORMATION

812

813 Supplementary Methods

814

815 *Z. marina* nitrogen uptake experiments

816 During incubations, plant shoots were loose in the incubation chambers and naturally
817 gained a relatively vertical alignment, with their roots pointing towards the bottom of
818 the chamber and the leaves to the surface. The media were constantly stirred during
819 incubation to decrease the thickness of the boundary layer at the leaf/root surface and to
820 ensure a homogeneous distribution of the isotopic labels and that leaf surfaces received
821 similar amounts of photosynthetic active radiation. The authors acknowledge that, in
822 this experiment, belowground plant parts were incubated in oxygenated conditions,
823 which is in contrast with the anoxic natural environment. However, preliminary
824 experiments showed no effect of rhizosphere oxygenation on the ammonium uptake
825 rates of leaves and roots, as was also reported for the sister species *Z. noltei* (Alexandre
826 et al., 2010; Alexandre et al., 2011).

827 Because no significant internal translocation of incorporated ^{15}N between leaves and
828 roots is expected within the chosen experimental incubation time (Vonk et al., 2008;
829 Alexandre et al., 2011), we assumed that the ^{15}N content determined in leaf and root
830 tissues after incubation represents only the amount of N that was taken up by the
831 respective plant parts. The removal of a substantial part of the microbial community
832 from the incubation media (artificial seawater filtered through 0.2 μm) and the short
833 incubation time (0.5 h) restricted exogenous DON mineralisation during the incubations
834 (Sharkey, 2007; Bauwe et al., 2010). Therefore based on these many previous studies,

835 we assume that most, if not all, of the amino acids and peptides were taken up by the
836 leaves and roots of *Z. marina* without prior cleavage.

837

838 Sample analysis

839 The concentration of ammonium was determined using the hypochlorite/indophenol
840 method (detection and quantification limits = 0.03 and 0.07 μM), whereas nitrate was
841 determined by the cadmium reduction method (detection and quantification limits =
842 0.06 and 0.11 μM). Amino acid N was determined according to Jones et al. (2002).
843 Briefly, 20 μL of sample or standard and 200 μL of working reagent (5 mL of OPAME
844 concentrate + 200 mL of borate buffer 0.02 M pH 9.5) were combined and read after 1
845 min on a LS-5B Fluorometer (Perkin Elmer Corp., Boston, MA) with an excitation
846 wavelength of 340 nm and an emission wavelength of 450 nm. The OPAME reagent
847 was obtained by dissolving 50 mg of *o*-phthaldialdehyde in 5 mL of methanol and
848 adding 100 μL of β -mercaptoethanol.

849

850 Supplementary Results

851

852 Concentration of inorganic and organic N

853 The concentration of peptides in the water column was higher than in the sediment, and
854 exceeded the total N by 3 to 10-fold, an incongruity that was not observed in the
855 sediment porewater (Table 1). We concluded that these values are not reliable,
856 suggesting an unresolved technical issue (e.g. contamination or unknown interference).
857 To overcome this, water column peptide values were estimated from the percentage of
858 peptides relative to the total soluble nitrogen observed in the sediment porewater (50 %
859 in Iceland, 8 % in the UK and 40 % in Portugal).

860

861 N uptake by *Zostera marina*

862 The uptake of dissolved inorganic nitrogen (DIN) by *Z. marina* generally displayed
 863 saturation kinetics whereas the uptake of DON was in most cases best described by a
 864 linear model, indicating that saturation was not reached within the range of nutrient
 865 concentrations used in the experiment (Fig. 1, Table S2).

866 The uptake of trialanine from the sediments increased only slightly with nutrient
 867 concentration ($F = 4.083$, $p = 0.035$) except for trialanine in Iceland, where values at 50
 868 μM ($0.12 \text{ nmol mL}^{-1} \text{ h}^{-1}$) were much smaller than at 5 μM and 15 μM (7.75 and 8.05
 869 $\text{nmol mL}^{-1} \text{ h}^{-1}$, respectively) (Fig. 4).

870

871 Supplementary Tables

872

873 Table S1. Average aboveground (leaves) and belowground (rhizomes + roots) biomass
 874 (g dry weight) of *Zostera marina* incubated in the high-, mid- and lower-latitude sites
 875 along the latitudinal distribution gradient.

876

	High-latitude	Mid-latitude	Lower-latitude
Aboveground	0.05	0.24	0.29
Belowground	0.03	0.13	0.22

877

878

879

880

881

882

883

884

885

886 Table S2. Uptake kinetic parameters of nitrogen uptake by *Zostera marina* plant parts at
 887 the three sites (Iceland, United Kingdom and Portugal) along the species' latitudinal
 888 distribution gradient. V_{\max} = maximum uptake rate ($\mu\text{mol N g}^{-1}$ dry weight h^{-1}); K_m =
 889 half-saturation constant (μM); and α = affinity constant (V_{\max}/K_m). The coefficient of
 890 determination (r^2), level of significance (P) and the standard error of the estimate
 891 (values in brackets) are given. Data not displaying saturation kinetics were fitted with a
 892 linear regression model (V = uptake rate; S = substrate concentration).
 893

	V_{\max}	K_m	α	r^2	P
<i>Ammonium (leaves)</i>					
Iceland	28.91 (2.30)	49.92 (8.73)	0.58	0.996	0.002
United Kingdom	54.63 (5.18)	84.24 (14.42)	0.65	0.997	0.001
Portugal	82.08 (24.48)	150.62 (67.20)	0.55	0.990	0.005
<i>Nitrate (leaves)</i>					
Iceland	0.59 (0.03)	2.05 (0.86)	0.29	0.83	0.087
United Kingdom	0.72 (0.06)	1.75 (1.27)	0.41	0.60	0.224
Portugal	2.19 (0.16)	6.56 (2.30)	0.33	0.94	0.029
<i>Alanine (leaves)</i>					
Iceland	1.95 (0.78)	32.60 (34.34)	0.06	0.82	0.096
United Kingdom	4.19 (1.50)	118.18 (67.95)	0.04	0.97	0.014
Portugal	4.62 (0.63)	53.68 (15.66)	0.09	0.99	0.006
<i>Trialanine (leaves)</i>					
Iceland	$V = 0.089 + 0.007 S$	-	-	1.00	0.012
United Kingdom	$V = 0.108 + 0.007 S$	-	-	1.00	0.012
Portugal	$V = 0.178 + 0.013 S$	-	-	0.99	0.016
	V_{\max}	K_m	α	r^2	P
<i>Ammonium (roots)</i>					
Iceland	$V = -0.486 + 0.151 S$	-	-	0.99	0.007
United Kingdom	19.39 (14.37)	111.61 (135.66)	0.17	0.87	0.069
Portugal	8.26 (1.58)	15.39 (10.65)	0.54	0.89	0.054
<i>Nitrate (roots)</i>					
Iceland	0.29 (0.04)	2.02 (2.19)	0.14	0.43	0.346
United Kingdom	1.81 (0.30)	27.00 (2.70)	0.07	0.95	0.027
Portugal	1.93 (0.45)	30.37 (19.23)	0.06	0.92	0.042
<i>Alanine (roots)</i>					
Iceland	$V = 0.439 + 0.004 S$	-	-	0.82	0.093
United Kingdom	$V = 0.493 + 0.009 S$	-	-	0.99	0.004
Portugal	$V = 0.954 + 0.014 S$	-	-	0.96	0.022
<i>Trialanine (roots)</i>					
Iceland	$V = 0.156 + 0.010 S$	-	-	0.99	0.023
United Kingdom	0.88 (0.03)	2.96 (0.56)	0.30	0.98	0.09
Portugal	$V = 0.832 + 0.025 S$	-	-	1.00	0.011

894 Table S3. Results of the two-way ANOVA examining the effects of latitude (L) and
 895 nutrient concentration (N) on the nitrogen uptake rates of *Zostera marina*. Test statistic
 896 (F) and levels of significance are indicated as (*) $P < 0.05$, (**) $P < 0.01$, (***) $P <$
 897 0.001 , (^{ns}) not significant.
 898

	Latitude (L)	Nutrient concentration (N)	L x N
<i>Leaves</i>			
Ammonium	F = 11.87***	F = 118.2***	F = 3.73**
Nitrate	F = 24.48***	F = 2.45 ^{ns}	F = 1.15 ^{ns}
Alanine	F = 22.51***	F = 45.70***	F = 3.16*
Trialanine	F = 18.31***	F = 55.05***	F = 2.49 ^{ns}
<i>Roots</i>			
Ammonium	F = 0.02 ^{ns}	F = 24.33***	F = 2.12 ^{ns}
Nitrate	F = 101.73**	F = 38.47***	F = 7.86***
Alanine	F = 84.83***	F = 35.18***	F = 3.96**
Trialanine	F = 118.12***	F = 43.46***	F = 8.45***

899
 900
 901
 902
 903
 904
 905
 906
 907
 908
 909
 910
 911
 912
 913
 914
 915
 916
 917

918 Table S4. Uptake kinetic parameters of nitrogen uptake by *Zostera marina* plant parts at
 919 three different temperatures (8, 12 and 17 °C). V_{\max} = maximum uptake rate ($\mu\text{mol N g}^{-1}$
 920 dry weight h^{-1}); K_m = half-saturation constant (μM); and α = affinity constant (V_{\max}/K_m).
 921 The coefficient of determination (r^2), level of significance (P) and the standard error of
 922 the estimate (values in brackets) are given. Data not displaying saturation kinetics were
 923 fitted with a linear regression model (V = uptake rate; S = substrate concentration).
 924

	V_{\max}	K_m	α	r^2	P
<i>Ammonium (leaves)</i>					
8 °C	40.48 (16.53)	90.20 (64.87)	0.45	0.96	0.021
12 °C	54.63 (5.18)	84.24 (14.42)	0.65	0.99	0.001
17 °C	130.76 (25.89)	294.41 (74.16)	0.44	0.99	0.001
<i>Nitrate (leaves)</i>					
8 °C	0.48 (0.08)	9.96 (7.12)	0.05	0.79	0.113
12 °C	0.72 (0.06)	1.75 (1.27)	0.41	0.60	0.224
17 °C	0.85 (0.11)	5.77 (3.77)	0.15	0.80	0.105
<i>Alanine (leaves)</i>					
8 °C	3.20 (0.81)	76.95 (36.09)	0.04	0.98	0.011
12 °C	4.19 (1.50)	118.18 (67.95)	0.04	0.97	0.014
17 °C	3.43 (0.59)	48.69 (18.49)	0.07	0.98	0.012
<i>Trialanine (leaves)</i>					
8 °C	$V = 0.063 + 0.005 S$	-	-	0.99	0.073
12 °C	$V = 0.108 + 0.007 S$	-	-	1.00	0.012
17 °C	0.72 (0.00)	11.44 (0.03)	0.06	1.00	0.001
	V_{\max}	K_m	α	r^2	P
<i>Ammonium (roots)</i>					
8 °C	9.66 (0.81)	40.38 (8.15)	0.24	0.99	0.004
12 °C	19.39 (14.37)	111.61 (135.66)	0.17	0.87	0.069
17 °C	$V = 1.113 + 0.119 S$	-	-	0.99	0.004
<i>Nitrate (roots)</i>					
8 °C	0.68 (0.13)	18.20 (11.77)	0.04	0.89	0.055
12 °C	1.81 (0.30)	27.00 (2.70)	0.07	0.95	0.027
17 °C	1.06 (0.14)	6.11 (4.20)	0.17	0.79	0.110
<i>Alanine (roots)</i>					
8 °C	1.30 (0.34)	29.78 (21.59)	0.04	0.87	0.066
12 °C	$V = 0.493 + 0.009 S$	-	-	0.99	0.004
17 °C	2.09 (0.22)	18.26 (6.32)	0.12	0.97	0.017
<i>Trialanine (roots)</i>					
8 °C	$V = 0.257 + 0.011 S$	-	-	1.00	0.010
12 °C	0.88 (0.03)	2.96 (0.56)	0.3	0.98	0.089
17 °C	1.57 (0.10)	5.82 (1.42)	0.27	0.98	0.090

925

926 Table S5. Results of the two-way ANOVA examining the effects of temperature (T) and
 927 nutrient concentration (N) on the nitrogen uptake rates of *Zostera marina*. Test statistic
 928 (F) and levels of significance are indicated as (*) $P < 0.05$, (**) $P < 0.01$, (***) $P <$
 929 0.001 , (^{ns}) not significant.
 930

	Temperature (T)	Nutrient concentration (N)	T x N
<i>Leaves</i>			
Ammonium	F = 10.36 ^{***}	F = 124.00 ^{***}	F = 2.82 [*]
Nitrate	F = 9.26 ^{**}	F = 3.21 [*]	F = 0.76 ^{ns}
Alanine	F = 4.25 [*]	F = 37.84 ^{***}	F = 0.55 ^{ns}
Trialanine	F = 38.70 ^{***}	F = 61.99 ^{***}	F = 2.44 ^{ns}
<i>Roots</i>			
Ammonium	F = 12.30 ^{***}	F = 45.36 ^{***}	F = 12.23 ^{***}
Nitrate	F = 6.27 ^{**}	F = 4.37 [*]	F = 0.96 ^{ns}
Alanine	F = 20.03 ^{***}	F = 28.42 ^{***}	F = 1.40 ^{ns}
Trialanine	F = 26.91 ^{***}	F = 18.84 ^{***}	F = 1.80 ^{ns}

931
 932
 933
 934
 935
 936
 937
 938
 939
 940
 941
 942
 943
 944
 945
 946
 947
 948
 949

950 Table S6. Whole-plant nitrogen budget ($\mu\text{mol m}^{-2} \text{h}^{-1}$) of *Zostera marina* along the
 951 species latitudinal distribution range in Europe, calculated using the nitrogen uptake
 952 rates ($\mu\text{mol g}^{-1} \text{dry weight h}^{-1}$), areal biomass ($\text{g}^{-1} \text{dry weight m}^{-2}$) and ambient nutrient
 953 concentrations (μM) determined at each site (Iceland, United Kingdom and Portugal).
 954 Values in brackets show the percentage contribution of each nitrogen source to the
 955 plant's total N acquisition. DIN = dissolved inorganic nitrogen, DON = dissolved
 956 organic nitrogen.
 957

	Iceland	United Kingdom	Portugal
<i>Leaves</i>			
Ammonium	4.6 (2.1 %)	91.6 (20.4 %)	94.8 (13 %)
Nitrate	4.5 (2.0 %)	9.9 (2.2 %)	91.8 (12.6 %)
Amino acids	0.8 (0.3 %)	4.4 (1.0 %)	3.2 (0.4 %)
Peptides	4.7 (2.1 %)	21.3 (4.8 %)	31.3 (4.3 %)
DIN	9.1 (4.1 %)	101.4 (22.6 %)	186.6 (25.6 %)
DON	5.4 (2.4 %)	25.7 (5.7 %)	34.5 (4.7 %)
<i>Roots</i>			
Ammonium	146.6 (65.8 %)	197.7 (44.1 %)	300.8 (41.2 %)
Nitrate	3.9 (1.8 %)	2.4 (0.5 %)	35.2 (4.8 %)
Amino acids	25.9 (11.6 %)	70.6 (15.7 %)	57.6 (7.9 %)
Peptides	31.8 (14.3%)	50.5 (11.3 %)	115.3 (15.8 %)
DIN	150.5 (67.6 %)	200.1 (44.6 %)	336.1 (46.0 %)
DON	57.7 (25.9 %)	121.1 (27.1 %)	173.0 (23.7 %)
<i>Whole-plant</i>			
Total DIN	159.6 (71.7 %)	301.5 (67.2 %)	522.7 (71.8 %)
Total DON	63.1 (28.3 %)	146.8 (32.8 %)	207.5 (28.2 %)

958
 959
 960
 961
 962
 963
 964
 965
 966
 967
 968
 969
 970

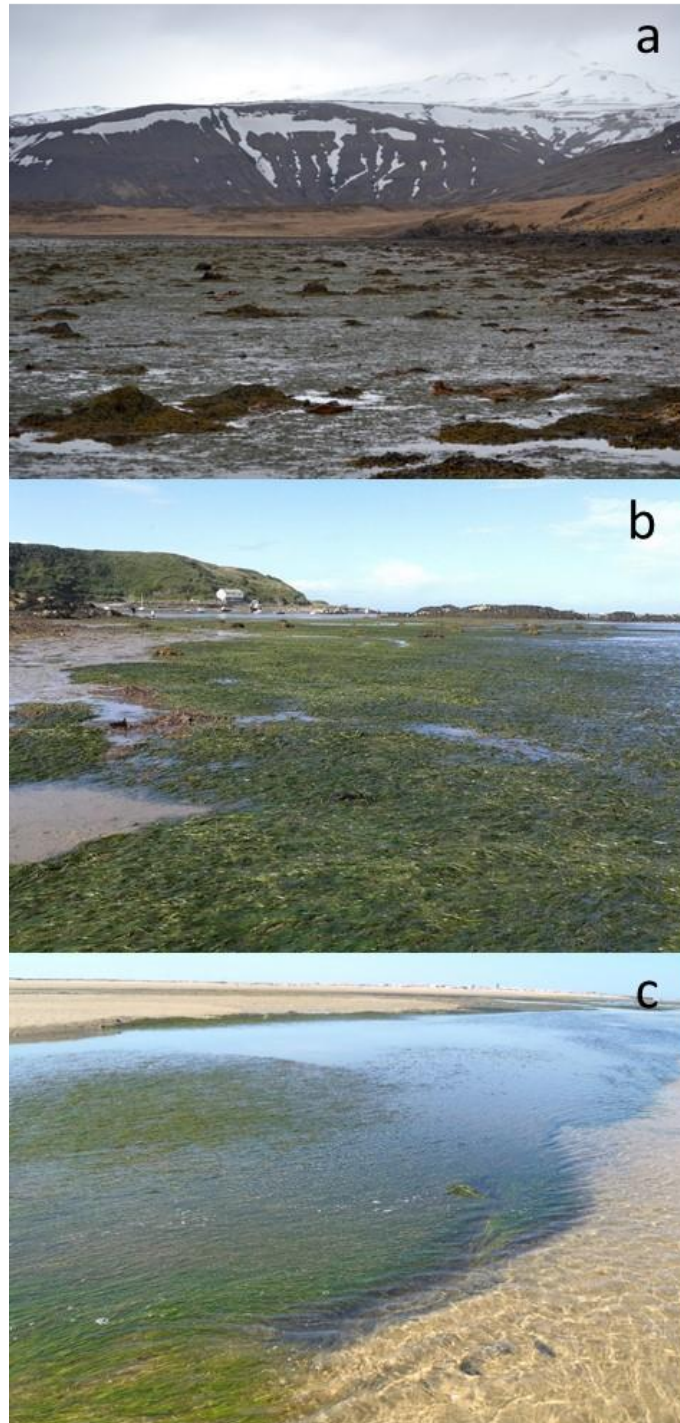
971 Table S7. Whole-plant nitrogen budget ($\mu\text{mol m}^{-2} \text{h}^{-1}$) of *Zostera marina* at three
 972 different temperatures (8, 12 and 17 °C), calculated using the nitrogen uptake rates
 973 ($\mu\text{mol g}^{-1}$ dry weight h^{-1}) obtained at each temperature, areal biomass (g^{-1} dry weight m^{-2})
 974 and ambient nutrient concentrations (μM) at the intermediate site (United Kingdom).
 975 Values in brackets show the percentage contribution of each nitrogen source to the
 976 plant's total N acquisition. DIN = dissolved inorganic nitrogen, DON = dissolved
 977 organic nitrogen.
 978

	8 °C	12 °C	17 °C
<i>Leaves</i>			
Ammonium	63.4 (16.4 %)	91.6 (20.4 %)	63.1 (12.3 %)
Nitrate	1.2 (0.3 %)	9.9 (2.2 %)	3.7 (0.7 %)
Amino acids	5.1 (1.3 %)	4.4 (1.0 %)	8.6 (1.7 %)
Peptides	12.6 (3.3 %)	21.3 (4.8 %)	11.0 (2.1 %)
DIN	64.6 (16.7 %)	101.4 (22.6 %)	66.8 (13.0 %)
DON	17.7 (4.6 %)	25.7 (5.7 %)	19.6 (3.8 %)
<i>Roots</i>			
Ammonium	214.6 (55.4 %)	197.7 (44.1 %)	243.3 (45.6 %)
Nitrate	1.3 (0.3 %)	2.4 (0.5 %)	5.7 (1.1 %)
Amino acids	58.9 (15.2 %)	70.6 (15.7 %)	108.3 (21.1 %)
Peptides	30.3 (7.8 %)	50.5 (11.3 %)	79.2 (15.4 %)
DIN	215.9 (55.7 %)	200.1 (44.6 %)	240.0 (46.7 %)
DON	89.2 (23.0 %)	121.1 (27.0 %)	187.4 (36.5 %)
<i>Whole-plant</i>			
Total DIN	280.5 (72.4 %)	301.5 (67.2 %)	306.8 (59.7 %)
Total DON	106.9 (27.6 %)	146.8 (32.8 %)	207.0 (40.3 %)

979
 980
 981
 982
 983
 984
 985
 986
 987
 988
 989
 990
 991

992 Figure S1. Aspect of the *Zostera marina* meadows in (a) Iceland (high-latitude), (b)
993 United Kingdom (mid-latitude), and (c) Portugal (lower-latitude) during summer at low
994 tide.

995



996