

In situ novel environment assay reveals acoustic exploration as a repeatable behavioral response in migratory bats

Schabacker, Theresa; Lindecke, Oliver; Rizzi, Sofia; Marggraf, Lara; Pētersons, Gunārs; Voigt, Christian C.; Snijders, Lysanne

Scientific Reports

DOI:
[10.1038/s41598-021-87588-y](https://doi.org/10.1038/s41598-021-87588-y)

Published: 14/04/2021

Publisher's PDF, also known as Version of record

[Cyswllt i'r cyhoeddiad / Link to publication](#)

Dyfyniad o'r fersiwn a gyhoeddwyd / Citation for published version (APA):
Schabacker, T., Lindecke, O., Rizzi, S., Marggraf, L., Pētersons, G., Voigt, C. C., & Snijders, L. (2021). In situ novel environment assay reveals acoustic exploration as a repeatable behavioral response in migratory bats. *Scientific Reports*, 11(1), Article 8174.
<https://doi.org/10.1038/s41598-021-87588-y>

Hawliau Cyffredinol / General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.



OPEN

In situ novel environment assay reveals acoustic exploration as a repeatable behavioral response in migratory bats

Theresa Schabacker¹, Oliver Lindecke^{1,2}, Sofia Rizzi^{1,3}, Lara Marggraf¹, Gunārs Pētersons⁴, Christian C. Voigt^{1,5} & Lysanne Snijders^{1,6}✉

Integrating information on species-specific sensory perception with spatial activity provides a high-resolution understanding of how animals explore environments, yet frequently used exploration assays commonly ignore sensory acquisition as a measure for exploration. Echolocation is an active sensing system used by hundreds of mammal species, primarily bats. As echolocation call activity can be reliably quantified, bats present an excellent model system to investigate intraspecific variation in environmental cue sampling. Here, we developed an in situ roost-like novel environment assay for tree-roosting bats. We repeatedly tested 52 individuals of the migratory bat species, *Pipistrellus nathusii*, across 24 h, to examine the role of echolocation when crawling through a maze-type arena and test for consistent intraspecific variation in sensory-based exploration. We reveal a strong correlation between echolocation call activity and spatial activity. Moreover, we show that during the exploration of the maze, individuals consistently differed in spatial activity as well as echolocation call activity, given their spatial activity, a behavioral response we term ‘acoustic exploration’. Acoustic exploration was correlated with other exploratory behaviors, but not with emergence latency. We here present a relevant new measure for exploration behavior and provide evidence for consistent (short-term) intra-specific variation in the level at which wild bats collect information from a novel environment.

Sensory systems are fundamental to survival and reproduction, acting at the interface between an organism’s central nervous system and its environment^{1,2}. Sensory systems allow organisms to collect, process, and respond adequately to environmental information, thereby reducing uncertainties about aspects of their environment³. Animals make decisions that are strongly influenced by the information they have of their environment⁴, which in turn is based on the information that was transferred through their sensory systems^{2,5}. Sensory systems can include ‘active’ and ‘passive’ sensing⁶. Passive sensing (e.g., vision) relies on sources that generate or reflect energy (e.g., electromagnetic energy from the sun), extrinsic to the sensing organism. Active sensing relies on the use of self-generated energy to receive information from the environment (e.g., echolocation), enabling animals to control characteristics of the probe energy, such as intensity, direction, and timing⁶.

The collection of environmental cues, stemming from active or passive sensing, is reflected by a behavior commonly referred to as ‘exploration’⁷. Through exploration, animals gain vital information about the availability of resources, the distribution of conspecifics, and the presence of predators. Animals frequently get challenged by changes in their environment, which have the potential to impact an individual’s fitness^{8,9}. Currently, anthropogenic change, and particularly climate change, is impacting the distribution and presence of mates, predators, prey, and suitable habitat in both space and time^{10–13}. Gathering detailed and up-to-date environmental information via exploration can help animals anticipate and mitigate such disruptive changes in their environment.

Former studies often assessed exploration levels in wild animals by measuring spatial activity as a proxy for exploration, by considering the speed of exploration in a novel environment or the proportion of a new

¹Department of Evolutionary Ecology, Leibniz Institute for Zoo and Wildlife Research, Berlin, Germany. ²School of Natural Sciences, Bangor University, Bangor LL57 2UW, Gwynedd, UK. ³Faculty of Life Sciences, Humboldt Universität Zu Berlin, Berlin, Germany. ⁴Faculty of Veterinary Medicine, Latvia University of Life Sciences and Technologies, Jelgava, Latvia. ⁵Institute of Biology, Freie Universität Berlin, Berlin, Germany. ⁶Behavioural Ecology Group, Wageningen University, Wageningen, The Netherlands. ✉email: lysanne.snijders@wur.nl

environment explored (as discussed by^{14,15}). Interestingly, many of these investigations revealed consistent differences in exploration levels among individuals of the same species (reviewed in^{16,17}), which were also found to be heritable in some species, such as the great tit, *Parus major*^{18,19}. Individual differences in exploration are not trivial¹⁷, as they may correlate with foraging strategies^{20,21}, dispersal²² or competitive ability²³. Yet, exploration, when defined purely by spatial measures, may primarily represent activity-level and not necessarily reflect the number or quality of environmental cues being collected by the individual¹⁴, hampering our mechanistic understanding of how intraspecific variation in exploration and environmental change are likely to interact and shape natural selection of a species.

Measuring exploration, including a measure for environmental cue sampling, is challenging in species that primarily use passive sensing, such as birds, (but see¹⁵). Bats, however, are a suitable study model to circumvent these limitations. Except for members of the family of Pteroporidae, bats have adapted to their nocturnal niche by using laryngeal echolocation²⁴. Echolocation is an active acoustic sensing system, enabling bats to individually control the incoming information about the environment, by actively manipulating the frequency, direction, and timing of echolocation pulses. Modern technology now allows us to reliably quantify echolocation calls in the field. Echolocation call activity thus provides us with the exciting opportunity to investigate intraspecific variation in how animals sample cues when exploring novel environments.

Bats use echolocation to orchestrate a variety of tasks, starting with the assessment of environments (i.e., orientation) in the complete darkness of their roosts²⁵ and under nightly low light conditions in flight. Echolocation supports hunting of prey on the wing²⁶, communication²⁷, and navigation during wayfinding using landmarks^{28,29}. The Nathusius' pipistrelle (*Pipistrellus nathusii*), a tree-roosting insectivorous bat, is widely distributed in Europe and migrates every year between breeding grounds in northeastern Europe and hibernation areas in southwestern Europe^{30–33}. Migratory bats are particularly vulnerable to environmental changes as they depend on specific conditions on a global geographical scale rather than in just one specific habitat³⁴. Moreover, Nathusius' pipistrelles suffer from extensive mortalities at wind turbines^{35,36}. For constructive conservation plans, it is, therefore, necessary to gain a deeper understanding of how bats perceive and react to environmental cues and to uncover potential variation among individuals.

Here, we developed an emergence and novel environment assay for tree-roosting bats with laryngeal echolocation to be used in the field. At a migratory corridor at the coastline of Latvia, we repeatedly successfully assayed (24-h time-interval) 52 male and female Nathusius' pipistrelles, quantifying behavior for the first two minutes following emergence into a roost-like novel environment, consisting of a maze-type arena with nine chambers through which the bat could crawl. We (1) tested whether echolocation call activity related to spatial activity in the novel environment. If echolocation call activity is relevant to exploration, we would expect the acoustic and spatial responses to be positively correlated. To examine whether echolocation call activity provided information about exploration behavior in addition to spatial activity, we quantified the number of echolocation calls while controlling for spatial activity and (2) tested whether this 'acoustic exploration' behavior was individually repeatable, i.e., whether certain individuals consistently under- or oversampled a novel environment. Lastly, we (3) tested whether acoustic exploration was related to emergence latency, as a proxy for boldness, to evaluate the potential presence of a behavioral syndrome³⁷.

Results

Variation in behavioral responses while exploring a novel environment. Bats fully entered the maze in 91 out of 111 assays (82%), representing 59 unique individuals (Fig. 1a). In addition, four bats emerged with their head but never with their full body. Fully emerged bats showed substantial variation in all analyzed behavioral responses (Supplementary Table 1), including emergence behavior, e.g., latency of body emergence (1 to 162 s, Fig. 1b) and duration between the head and full-body emergence (1 to 114 s, Fig. 1c), acoustic behavior, e.g., number of echolocation calls during emergence (4 to 335, Fig. 1d) and the number of echolocation calls during the first 2 min after emergence (170 to 2127, Fig. 1e), and spatial behavior, e.g., number of unique chambers (1 to 9, Fig. 1f) and the total number of chambers (1 to 25, Fig. 1g) visited during the first 2 min in the novel environment. Furthermore, the number of times a bat poked its head in an adjacent chamber ranged from 0 to 7, the number of emitted 'air puffs', i.e., brief vocal outbursts audible to the human ear, ranged from 0 to 22 and in 23 out of 91 assays a bat (also) crawled through the novel environment upside down (Supplementary Table 1).

Echolocation call activity positively correlates to spatial activity in a novel environment. Different response measures of the same behavioral category were moderately to strongly correlated to each other ($r_s = 0.25–0.93$ for the emergence measures, $r_s = 0.22–0.98$ for the acoustic measures, and $r_s = 0.89$ for the spatial measures, Supplementary Table 2). The acoustic and spatial responses to a novel environment were strongly correlated to each other, with bats that visited more chambers also emitting more echolocation calls ($r = 0.65$, $p < 0.001$, Fig. 2a). Nevertheless, there was substantial variation in echolocation among bats visiting a similar number of chambers (Fig. 2a), a behavioral response we termed 'acoustic exploration' (Fig. 2b).

Individuals consistently under or oversample a novel environment. We were able to conduct two valid novel environment assays, approximately 24 h apart, with 52 of the 59 earlier-mentioned individuals, resulting in 104 valid tests for the consistency analyses. Bats fully entered the maze in 85 out of these 104 tests (82%). Acoustic exploration was significantly reduced during the second assay (Estimate (Est.) ± Standard Error (SE): -344.60 ± 52.43 , Cohen's $d = -1.51 \pm 0.23$, $p < 0.001$, Fig. 3a, Supplementary Table 3) and males showed more acoustic exploration than females (Est. ± SE = 144.71 ± 69.44 , Cohen's $d \pm SE = 0.63 \pm 0.31$, $p = 0.04$, Fig. 3b, Supplementary Table 3). Bats showed more acoustic exploration when the set-up faced more South compared to North and more East compared to West (North–South: Est. ± SE = -93.99 ± 43.81 , $p = 0.03$; East–West:

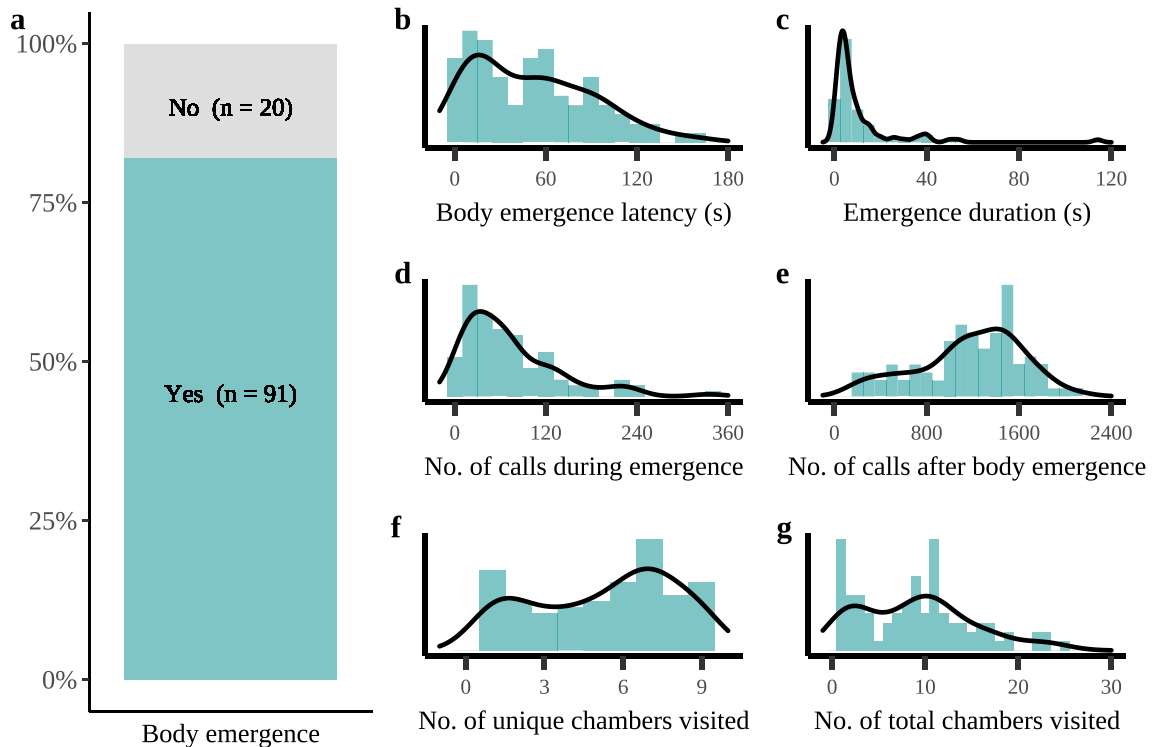


Figure 1. Variance in behavioral responses in *Nathusius' pipistrelles* exploring a novel environment. (a) In 82% of the tests, the bat fully emerged out of the starting tube and entered the novel environment (a maze-like arena). The substantial variation in the behavioral response to the novel environment is visualized for (b) latency to enter the novel environment (s), (c) duration between head and full-body emergence (s), (d) the number of echolocation calls during emergence (i.e., time between head and body emergence), (e) number of calls after full body emergence, (f) number of unique chambers visited and (g) the total number of chambers visited during the first two minutes in the novel environment. The lines in panels b–g show the density distribution while the bars show the frequency distribution. Note that although the density distribution extrapolates to negative values, the study data (as shown by the frequency distribution) only included positive values.

Est. \pm SE = 85.95 ± 42.36 , $p = 0.04$, Supplementary Fig. 1, Supplementary Table 3). Interestingly, individuals consistently differed in how thoroughly they acoustically explored the novel environment, i.e., the relative number of echolocation calls they emitted per chamber ($R_{\text{adj}} = 0.32$, SE = 0.15, CI = 0.07–0.62, $p = 0.03$, Fig. 3c).

Bats also tended to visit fewer chambers during the second test (Est. \pm SE = -0.28 ± 0.14 , $p = 0.053$, Supplementary Table 4) and individuals consistently differed in how many chambers they visited ($R_{\text{adj}} = 0.29$, SE = 0.16, CI = 0–0.58, $p = 0.048$). Test number did not significantly influence emergence latency (Est. \pm SE = -0.34 ± 0.23 , $p = 0.13$, Supplementary Table 5) and individuals that emerged quicker during the first test tended to also emerge quicker during the second test ($R_{\text{adj}} = 0.23$, SE = 0.13, CI = 0–0.50, $p = 0.054$). Yet, it should be noted that no repeatable individual differences in emergence latency could be detected when the minimum latencies for non-emergers were completely excluded (181 s, 18% of tests).

Acoustic exploration correlates to other exploratory behaviors but not to emergence latency.

Bats that emerged quicker (or slower) did not exhibit more acoustic exploration (Est. \pm SE = 5.00 ± 27.48 , $p = 0.86$, Fig. 4a, Supplementary Table 6). However, bats that emitted relatively more echolocation calls during emergence (Residuals of: Number of calls during emergence ~ Emergence duration (s): $r_s = 0.79$, $p < 0.001$) similarly showed more acoustic exploration after emergence (Est. \pm SE = 2.36 ± 0.57 , $p < 0.001$, Fig. 4b, Supplementary Table 6). In addition, bats that poked their heads into an adjacent chamber more often also showed more acoustic exploration (Est. \pm SE = 57.87 ± 19.16 , $p = 0.003$, Fig. 4c, Supplementary Table 6). The number of air puffs a bat emitted did not correlate with acoustic exploration (Est. \pm SE = 7.71 ± 7.85 , $p = 0.33$, Supplementary Table 6). Finally, there was a trend for bats that crawled upside down for part of the test to exhibit less acoustic exploration (Est. \pm SE = -128.15 ± 68.94 , $p = 0.06$, Supplementary Table 6).

Discussion

Animals use their sensory systems to explore (i.e., examine and investigate) environments and objects, a behavior crucial for individuals to keep up to date with the presence and distribution of resources and threats and so reduce uncertainty about their environment³⁸. Yet, most of our current understanding of variation in exploration behavior is based on where animals go rather than the information they collect. Here, we reveal a relevant new measure for exploration behavior by quantifying sensory activity relative to spatial activity and subsequently

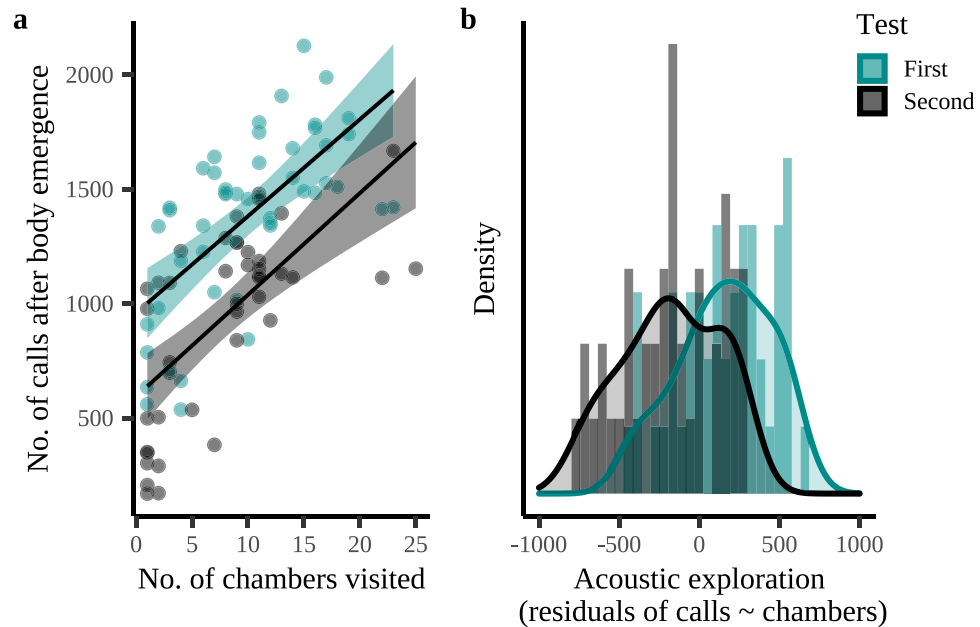


Figure 2. The number of echolocation calls emitted during the first and second test as a function of the number of chambers visited by *Nathusius' pipistrelles* in a novel environment. **(a)** Bats emitted more echolocation calls during the first 2 min in the novel environment (a maze-like arena) when they also visited more chambers. **(b)** Bats varied substantially in how much more or less they echolocated than expected based on the number of chambers they visited, i.e., the residuals of the number of echolocation calls as a function of the number of chambers visited varied considerably. Regression lines in panel **(a)** show the predicted final model values. Shaded areas around the lines reflect the 95% Confidence Interval. Lines in panel **(b)** show the density distribution while the bars show the frequency distribution.

providing evidence for consistent (short-term) variation between individual bats in how thoroughly they acoustically explore a novel environment.

With our newly developed in situ assay, we were not only able to uncover substantial intraspecific variation in bats' responses to a novel environment but also to reveal that individual bats consistently varied in these responses. Spatial movement and acoustic activity, both highly relevant behaviors in the context of orientation and navigation^{28,39}, were strongly positively correlated. In line with previous studies investigating exploration behavior in other species (e.g., fish^{21,40}, amphibians⁴¹, birds^{42,43}, mammals^{44,45}), we found spatial activity in a novel experimental arena to be individually repeatable. We add to these studies by now showing that also sensory activity (e.g., number of echolocation calls), given spatial activity, is an individually repeatable response, which we termed 'acoustic exploration'. Our repeatability estimate for acoustic exploration ($R=0.32$) was similar to those reported for a range of behavioral traits across a wide variety of animal taxa (average $R=0.37$)¹⁶. Importantly, acoustic exploration correlated with the number of times an individual poked its head into an adjacent chamber and the relative echolocation call rate during emergence, suggesting acoustic exploration to represent true exploration behavior, rather than activity. Exploration behavior frequently correlates with other behavioral traits, including aggressiveness⁴⁶ and boldness⁴⁷, forming a behavioral syndrome³⁷. Though, we did not find evidence for a correlation between acoustic exploration and emergence latency, an often-used proxy for boldness (e.g.,⁴⁷), in this study. A recent examination of in situ takeoff orientation in this species using a circular release box set-up⁴⁸ revealed emergence latency values that were highly similar (range: 6–176 s, median: 55 s) to those we report here (Supplementary Table 1). Both set-ups challenged individuals with a multi-directional choice for movement in a novel environment, raising the possibility that not only risk-aversion but also decision or orientation-speed may have influenced emergence latency, a hypothesis that requires further testing.

Interestingly, exploration behavior has also been linked to routinization⁴⁹ and learning ability⁴³. Thorough exploration likely creates a greater knowledge of the state and quality of the habitat, allowing individuals to better react to spontaneously arising environmental stimuli⁵⁰. Building forward on this, researchers have proposed the existence of a 'cognitive syndrome', based on the assumption that individuals differ in the speed and accuracy of picking up environmental cues leading to two alternative cognitive styles, namely 'fast and superficial' and 'slow but accurate'⁵¹. This theoretical framework suggests that some individuals initially learn about specific contingencies more quickly, potentially by recognizing and memorizing them faster and by being more susceptible to forming behavioral routines⁵². However, as the formation of behavioral routines is associated with some degree of inflexibility, these individuals are expected to be more challenged in tasks that require behavioral alterations in response to altered environmental conditions^{53,54}. Thorough exploring bats may have an evolutionary advantage when facing novel threats and habitat structures (e.g.,^{55,56}), as an increase in sensory update rate is thought to allow faster behavioral responses^{39,57}. Thorough explorers, i.e., individuals that echolocate more per unit of distance and therefore collect more information in a given environment, can thus be expected to detect

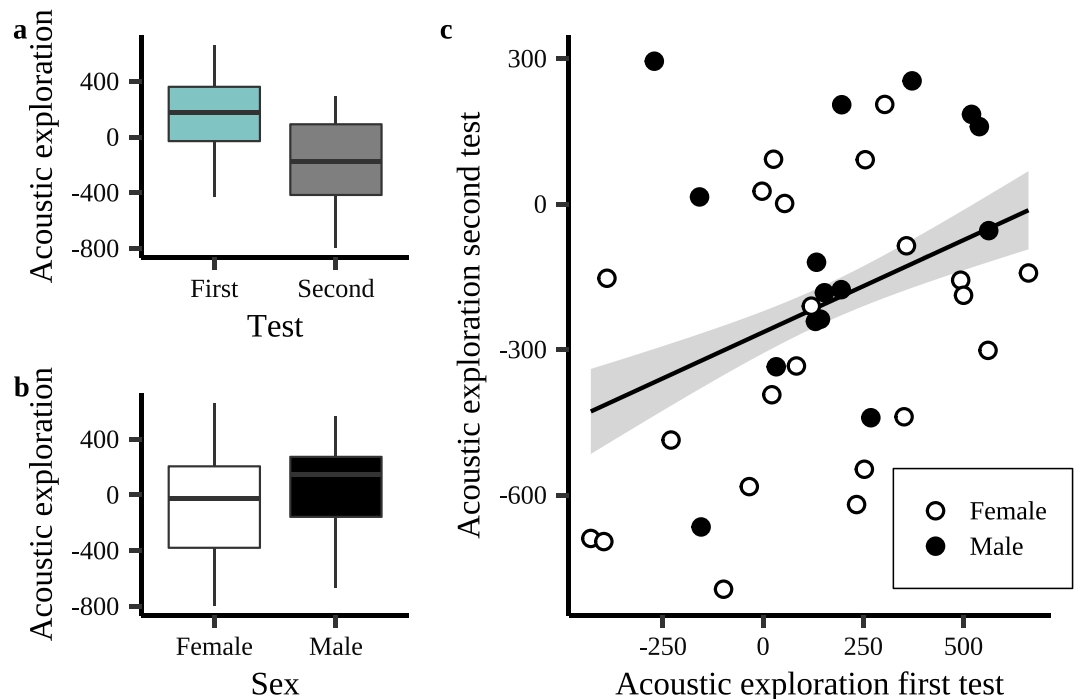


Figure 3. Acoustic exploration, i.e., residuals of the number of echolocation calls as a function of the number of chambers visited, during the first and second novel environment test with *Nathusius' pipistrelles*. (a) Individuals show more acoustic exploration during the first than the second novel environment test (a maze-like arena). (b) Males show more acoustic exploration than females. Males and females are represented by a black or white fill, respectively. (c) Acoustic exploration during the first novel environment test predicts an individual's acoustic exploration during the second test. The regression line in panel (c) shows the predicted final model values. Shaded areas around the line reflect the 95% Confidence Interval. Box plots in panels (a) and (b) show median and interquartile range with whiskers of 1.5 interquartile distances.

changes in the environment more quickly⁵⁸, increasing their adaptive potential and resilience to environmental change^{59,60}. For example, exploration behavior in a novel environment by Iberian wall lizards (*Podarcis hispanica*) was positively associated with the habituation speed to a threatening stimulus⁶¹. Linking individual variation in acoustic exploration to the ability of bats to quickly detect and respond to novelty on a wider spatial scale, or a comparative investigation of bat populations living along a gradient of habitat stability, would be exciting next steps. Furthermore, more intense use of an active sensory system, such as echolocation, is not necessarily without energetic costs⁶², so an examination of the potential costs linked to more thorough exploration in bats would likewise be insightful.

Our behavioral repeatability estimates were based on a short time interval (24 h) and ideally, longer intervals would be used. However, it is extremely challenging logistically to assess wild migratory and nocturnal animals over a longer time. We, therefore, see our study as the first step towards evidence for consistent individual differences in acoustic exploration behavior in wild migratory bats and encourage follow-up studies employing longer timespans to further investigate the stability of these differences. We also witnessed a reduction in acoustic exploration and spatial activity during the second assay which might indicate a learning process. Potentially, bats learned during the first assay that particular chambers were unrewarding, that there was no exit from the maze, and/or that the maze was not dangerous, and, consequently, reduced their exploration activity during the second assay. This latter phenomenon is referred to as habituation, where individuals learn that a certain stimulus is irrelevant^{52,63}. Habituation, and other forms of learning, are a common problem in novelty tests as repeated exposure inherently reduces novelty and creates a sense of familiarity^{64,65}. As a result, the motivation, or the need, to explore diminishes as the environment becomes more familiar^{66–68}. However, habituation also has adaptive significance, as it allows animals to separate repeated, potentially irrelevant, stimuli from important ones⁶⁹. Even though we were still able to reveal individual repeatability in exploration behavior, future studies could control for habituation effects by deploying a different type of novel environment during the second assay. Such a set-up would also account for other types of learning effects and could be adapted to investigate the presence of consistent individual differences in habituation, by repeatedly testing habituation to novelty, using a reaction-norm approach⁷⁰.

Despite their high diversity and multitude of ecosystem services, few studies have investigated the consistency of intraspecific variation in bats' behavioral responses and how this variation may influence a bat's ecology and individual fitness. Previous studies have shown intraspecific variation in foraging^{71–75} and migration strategies⁷⁶ and it would be plausible for such ecologically relevant traits to not only correlate to inter-individual morphological variation⁷⁷ but also to intrinsic behavioral differences, such as the ones measured in this study. Some

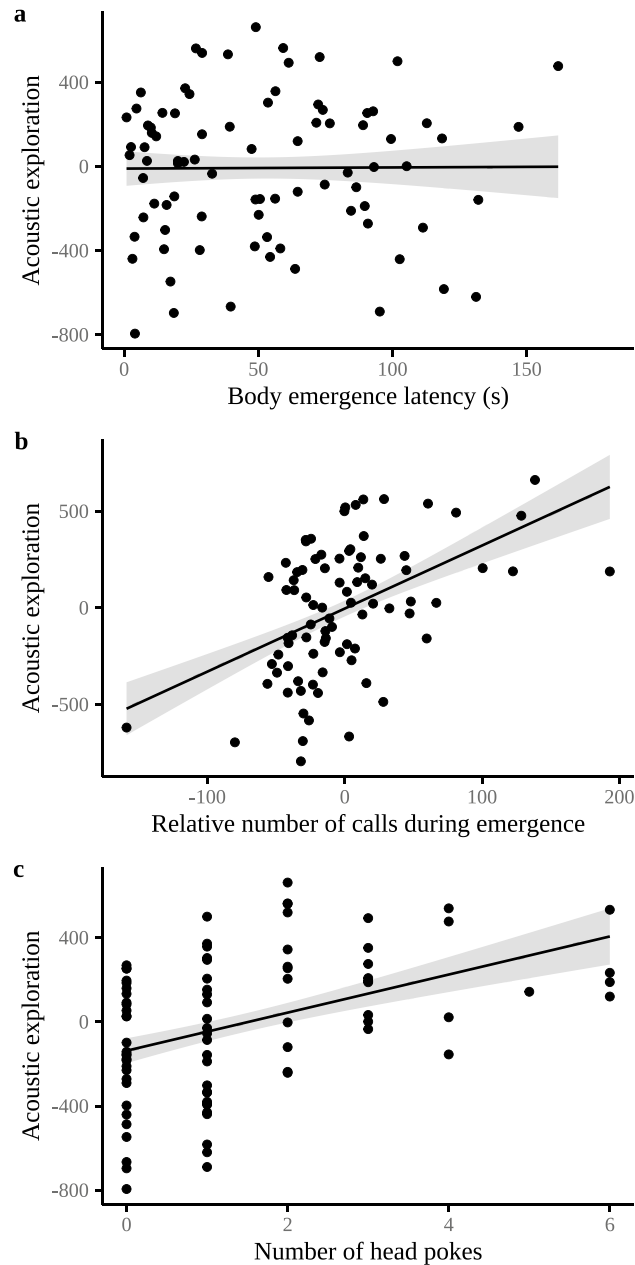


Figure 4. Acoustic exploration, i.e., residuals of the number of echolocation calls as a function of the number of chambers visited, in relation to other behavioral responses exhibited by *Nathusius' pipistrelles* in a novel environment. These behavioral responses exhibited in the novel environment (a maze-like arena) included: (a) Body emergence latency (s), (b) Acoustic exploration during emergence, i.e., the residuals of the number of echolocation calls as a function of emergence duration (time between the head out and the body out of the starting tube), (c) the number of times a bat poked its head into an adjacent chamber without entering. Regression lines show the predicted final model values. Shaded areas around the lines reflect the 95% Confidence Interval.

studies on species from the family of Vespertilionidae showed that these bats exhibit consistent intraspecific variation in (aggregate) behavioral responses linked to exploration, activity, anxiety⁷⁸, and aggressiveness⁷⁹. Moreover, this variation was linked to pathogen dynamics⁸⁰. Next to foraging and migration strategies, habitat choice is another major selection process in the lives of many bat species and earlier research suggests variation in individual habitat preferences among and within sexes⁸¹. For tree-dwelling bat species, roosts are crucial for resting⁸², reproducing⁸³, and mitigating predation risk⁸⁴. Thus roost deficits could be detrimental for bats⁸⁵. However, the large majority (97%) of European forests are managed and under commercial use⁸⁶, e.g., felling of trees and removal of deadwood, resulting in reduced habitat quality for bats⁸⁷. Especially for migrants, knowledge about adequate roosts along the route is vital and bats can be expected to remember roost locations even over hibernation⁸⁸. If trees or forest areas with suitable roosts are lost due to anthropogenic activities, these conditions

may favor high levels of exploration and/or plasticity. Thus, thorough explorers may have a selective advantage, resulting in a population-level shift in exploration behavior and associated behaviors and a potential increase in intraspecific competition⁸⁹.

For the Nathusius' pipistrelle, the time of late summer to autumn migration overlaps with the time of mating, which peaks in August in Latvia^{30,32,90}. During this period, males are typically more conspicuous, as they produce loud acoustic displays as advertisement calls during flight or while roosting, which are believed to repel other males and/or attract females⁹¹. This may partially explain the higher acoustic exploration of males in this study. It would thus be interesting to test if similar sex-differences are apparent also outside the mating season. The observed higher levels of acoustic exploration when the starting tube of the maze was facing South or East may be related to bats' migratory behavior. Nathusius' pipistrelles appear to have an intrinsic orientation, as these bats showed consistent departure flight directions in the migratory direction, even when translocated more than 10 km away from the capture site⁹². It is therefore possible that the alignment of the maze with the migratory direction stimulated enhanced acoustic exploration, but further hypothesis-based testing would be necessary to draw reliable conclusions.

Conclusion

Evaluating animal exploration is a challenging task, as this behavior is defined in terms of information acquisition⁷, but classically measured in terms of movement speed or proportion of a novel environment explored^{15,93}. Here, we address this challenge by developing an in situ assay to assess echolocation call activity relative to spatial activity while exploring a novel environment. The assessment of acoustic exploration in a tree-roosting bat species proved to be a very suitable approach to obtain high-resolution sensory-based data on individual behavior in response to a novel environment. Next to providing novel evidence for an exploration-relevant repeatable behavioral trait, we hope our approach instigates further research into intraspecific variation in environmental cue sampling, using species with a predominant use of active sensing modalities as study models, and inspire a species-specific sensory perspective on how individual animals may respond differently to environmental novelty.

Materials and methods

Study population. 60 adult migratory Nathusius' bats were caught at the Pape Ornithological Station, University of Latvia, Latvia (56°09' N, 21°03' E) between 09:00 PM and 03:00 AM on the 16th and 18th of August 2019, using a Helgoland funnel trap³² as part of the long-term Pape Station bat ringing program. Our study adhered to the ASAB/ABS guidelines for the treatment of animals and the experimental set-up and assay protocol for assessing exploration behavior in tree-roosting bats were evaluated and supported by the institutional animal care and welfare representatives of the Leibniz Institute for Zoo and Wildlife Research. Because the bats were caught as an integral part of the independent ringing program and the behavioral assays were of a non-invasive nature, the study did not entail further ethical approval. The work was conducted under permit no. 49/2019 issued by the Latvian Nature Conservation Agency to G.P. and complies with the ARRIVE guidelines. Note that this was a descriptive study and therefore did not include control groups.

General procedure. Following capture, we determined the sex, body mass (digital balance), and forearm length (manual calipers). Bats received a temporary ID-coded gummy ring for individual identification. The bats were kept together in a dark and quiet environment (dark, wooden shed) in wooden carriers with a mesh top and covered by a dark cloth (seven to eight individuals per 30 cm × 30 cm × 10 cm), mimicking natural daytime roost conditions during which bats save energy by going into torpor. The bats' general condition and body mass was checked daily, and water and food were offered, with individuals weighing less than seven grams being given supplemental feeding (Harrison's Juvenile Hand-Feeding Formula). Behavioral assays were conducted on two consecutive nights following capture and took place in a dark wooden cabin (3 m × 2 m, constant temperature of 21 ± 2 °C) within 200 m of the location of capture.

Set-up and protocol. Prior to an assay, a bat was kept in an individual cotton bag (Ecotone, Gdansk, Poland) near a wrapped-up warm water bottle to stimulate it to come out of torpor. The bat's normothermia was confirmed by measuring the skin temperature (> 30 °C) with a thermocouple (Peakmeter, PM6501; Thermocouple, Sensor SSP-1-150, Peakmeter, Shenzhen, China). Skin temperature is a non-invasive measurement that accurately reflects rectal temperature⁹⁴.

We encouraged natural exploration behavior by constructing an experimental maze-like arena, mimicking a novel environment that would be relevant for a tree-roosting bat species (Fig. 5; 40 cm × 40 cm × 5 cm). The maze, placed in a larger box (70 cm × 45 cm × 8 cm) with a transparent plastic lid, consisted of nine separate chambers accessible through small gates (3 cm × 2.5 cm) on the upper half of the walls (Fig. 5). The maze was placed horizontally to stimulate exploratory movements in all directions. The floor was covered with a plastic non-slip mat to supply the bat with a good texture for crawling, a common behavior for tree-roosting bats. A layer of insect screen covered the maze (not shown in Fig. 5), offering additional climbing opportunities and preventing the bat from escaping. This setup enabled us to capture the bat's movement via a night vision camera (Sony Digital Camcorder, DCR-SR72E, Sony, Tokyo, Japan) mounted on a tripod with a horizontal arm to allow an aerial view from 1.5 m. An infra-red flashlight (T38, Evolve Future Technology, Shenzhen, China), shining at an angle from a fixed position, provided light for the camera (the camera's light was taped off to prevent reflection). A directional USG Electret Ultrasound microphone (Avisoft Bioacoustics/Knowles FG, Berlin, Germany), receiving sound within a 180° range, connected to an ultrasound recording device (UltraSoundGate 116Hb, Avisoft Bioacoustics, Berlin, Germany) was placed within the larger box pointing to the center of the maze, to

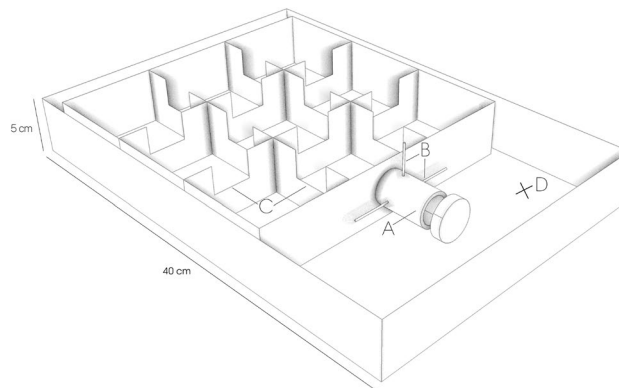


Figure 5. Schematic drawing of the maze used during the behavioral assay. (a) Opaque start tube where bats were placed at the start of each assay (b) Barriers closing the entrance to the maze (c) Gates connecting the chambers (d) Position of the microphone (USG Electret Ultrasound Microphones, Avisoft Bioacoustics/ Knowles FG, 180° field) pointing to the center of the maze. Artwork by Rebecca Scheibke.

record all vocalizations. An opaque start tube (10 cm × 3 cm) was attached to the maze but blocked by a small wooden barrier which we removed at the start of the assay (Fig. 5).

We placed the bat in the starting tube, with the exit obstructed by vertical and horizontal barriers in the form of wooden dowels. After 20 s we removed the vertical barrier and waited for a maximum of three minutes for the bat to emerge⁴⁸. The time taken for the bats to completely emerge from the start tube was recorded. If the bat did not emerge within this time, the test was terminated. After emergence, bats were given two minutes to explore the box. After each test, we sterilized the maze with a mild unscented detergent to remove potential olfactory cues left by previous subjects. Halfway through the night, we rotated the assay to control for potential orientation biases. Flight orientation was briefly assessed on the same night (different study) and controlled for in the analysis (see ‘Statistical analysis’ section). Bats were released the night of the second assay within 100 m of the original capture location.

Behavioral response quantification. We used the event-logger BORIS⁹⁵ version 7.9.1 to quantify several behavioral response measures in detail: latency to head emergence (s), latency to full body emergence (s), number of unique chambers discovered, the total number of chambers visited, number of times a bat poked its head in an adjacent chamber and whether a bat had crawled upside down through the maze. In addition, we recorded the echolocation activity during emergence as well as after head and full-body emergence and quantified the number of echolocation calls using Avisoft SASLab Pro version 5.2 (Avisoft Bioacoustics, Berlin, Germany). Prior to analysis, sampling frequency was converted from 250 to 150 kHz. The spectrogram was computed with Fast Fourier Transformation 256, parameters were set to Hamming window (bandwidth 1270 Hz, resolution 977 Hz) and volume normalized to 75%. We synchronized the audio recordings with the video recordings via spoken start and stop commands given by the observer. We used Avisoft’s call detection and template-based spectrogram comparison feature to identify distinct echolocation calls, which were verified and corrected via visual inspection (correlation corrected and uncorrected: $r_s > 0.99$, $p < 0.001$). Finally, we quantified the number of ‘air puffs’, brief audible acoustic outbursts that sound similar to sneezes. The behavioral response quantification was conducted blind, with only the observation number being known by the observer.

Statistical analysis. We conducted all analyses with R⁹⁶ version 4.0.2 in R Studio version 1.2.5033 (2009–2019 RStudio, Inc.). All statistical tests were two-sided. We first conducted Pearson (parametric) and Spearman (non-parametric) correlation tests between measures within a behavioral category (e.g., acoustic behavior), using all valid test data ($n = 111$ tests without premature emergence, 59 out of 60 individuals), to evaluate whether we could proceed with the analysis using one representative response measure and so minimize redundancy. Next, using a Pearson correlation test, we tested whether the number of echolocation calls following the first 2 min since emergence correlated to the number of chambers visited. Subsequently, we extracted the residuals for this correlation (i.e., the ‘acoustic exploration’ measure) using a linear model. We additionally tested whether the number of echolocation calls emitted during emergence was correlated to the duration of emergence (seconds between the head and full-body emergence) using a Spearman correlation test and extracted the residuals using a linear model.

Three mixed model selection procedures were conducted using the lmer and glmer functions of the ‘lme4’ package⁹⁷, to identify variables that significantly affect (1) acoustic exploration, (2) the number of chambers visited, and (3) emergence latency. These models only included data for the 52 individual bats ($n_{\text{♀}} = 31$, $n_{\text{♂}} = 21$) for which we had two valid test responses (i.e., no premature emergence or escape). We used stepwise backward model selection, assessing the significance of fixed effects by the change in deviance upon removal of the effect (Type II Wald Chi-square tests). Effects with $p < 0.1$ were kept in the final model. All models included individual ID as a random effect and all three starting models included the independent variables: test (two-level factor), sex (two-level factor), weight in grams (scaled covariate), forearm length in mm (scaled covariate), time since sunset

(scaled covariate), direction of starting tube (two integer variables: North–South with North = 1, South = -1, East and West = 0; East–West with East = 1, West = -1, North and South = 0), supplemental food (two-level factor) and orientation assessment (two-level factor). The factor ‘test’ was kept in the model at all times. Potential collinearity was assessed using the variation inflation factor (VIF) extracted with the ‘performance’ package⁹⁸ and variables above 2.5 are reported in the Supplementary Information (none of our variables of interest, but some of our control variables showed collinearity).

First, effects on acoustic exploration were analyzed using a linear mixed model (LMM). The model assumptions were evaluated using the Shapiro–Wilk test and a visual inspection of the model residuals. Second, effects on the visited number of chambers were analyzed using a generalized linear mixed model (GLMM) with a Poisson error-distribution and the bobyqa optimizer. Following overdispersion, an observation-level random effect (i.e., observation number) was added to the model. Third, effects on body emergence latency were analyzed by log-transforming emergence latency and using an LMM. Values for non-emergers were set to the minimal possible value of 181 s. Although the Shapiro–Wilk test indicated a significant deviation from normality during the model selection steps for emergence latency ($0.02 < p < 0.03$), the histogram of the residual frequency distribution did not show substantial deviations and the check_distribution function of the ‘performance’ package indicated a 100% probability of the model residuals originating from a normal distribution.

Repeatability (R_{adj}) of acoustic exploration, number of visited chambers, and emergence latency (log-transformed) was quantified with the ‘rptR’ package⁹⁹ using the final model from the model selection procedure. Confidence intervals for the repeatability estimates were bootstrapped (1000 times).

To evaluate potential correlations between acoustic exploration, and other behavioral response measures, the respective response measure was added to the final model for acoustic exploration and significance was assessed using Type II Wald Chi-square tests. To aid interpretation, we calculated the effect sizes for categorical variables with a significant effect on the unitless acoustic exploration measure using the ‘emmeans’ package¹⁰⁰. Figures were created using the ‘ggplot2’ package¹⁰¹.

Data availability

All data generated or analyzed during this study are included in this published article and the Supplementary Information files.

Received: 16 December 2020; Accepted: 31 March 2021

Published online: 14 April 2021

References

- Dangles, O., Irschick, D., Chittka, L. & Casas, J. Variability in sensory ecology: expanding the bridge between physiology and evolutionary biology. *Q. Rev. Biol.* **84**, 51–74 (2009).
- Dusenbery, D. B. *Sensory Ecology: How Organisms Acquire and Respond to Information* (W.H. Freeman, 1992).
- Dall, S. R., Giraldeau, L.-A., Olsson, O., McNamara, J. M. & Stephens, D. W. Information and its use by animals in evolutionary ecology. *Trends Ecol. Evol.* **20**, 187–193 (2005).
- Krebs, J. R. & Kacelnik, A. Decision making. In *Behavioural Ecology: An Evolutionary Approach* (eds Krebs, J. R. & Davies, N. B.) 105–136 (Blackwell Scientific Publications, 1991).
- Stevens, M. *Sensory Ecology, Behaviour, and Evolution* (OUP, 2013).
- Nelson, M. E. & MacIver, M. A. Sensory acquisition in active sensing systems. *J. Comp. Physiol. A* **192**, 573–586 (2006).
- Reader, S. M. Causes of individual differences in animal exploration and search. *Top. Cogn. Sci.* **7**, 451–468 (2015).
- Lane, J. E., Kruuk, L. E. B., Charmantier, A., Murie, J. O. & Dobson, F. S. Delayed phenology and reduced fitness associated with climate change in a wild hibernator. *Nature* **489**, 554–557 (2012).
- Nussey, D. H., Postma, E., Gienapp, P. & Visser, M. E. Selection on heritable phenotypic plasticity in a wild bird population. *Science* **310**, 304–306 (2005).
- Walther, G.-R. *et al.* Ecological responses to recent climate change. *Nature* **416**, 389–395 (2002).
- Bradshaw, W. E. & Holzapfel, C. M. Evolutionary response to rapid climate change. *Science* **312**, 1477–1478 (2006).
- Parmesan, C. Ecological and evolutionary responses to recent climate change. *Annu. Rev. Ecol. Syst.* **37**, 637–669 (2006).
- Palmer, C. Climate change, ethics, and the wildness of wild animals. In *Animal Ethics in the Age of Humans: Blurring Boundaries in Human-Animal Relationships* (eds Bovenkerk, B. & Keulartz, J.) 131–150 (Springer, 2016).
- Minderman, J., Reid, J. M., Evans, P. G. & Whittingham, M. J. Personality traits in wild starlings: exploration behavior and environmental sensitivity. *Behav. Ecol.* **20**, 830–837 (2009).
- Huang, P., Kerman, K., Sieving, K. E. & St. Mary, C. M. Evaluating the novel-environment test for measurement of exploration by bird species. *J. Ethol.* **34**, 45–51 (2016).
- Bell, A. M., Hankison, S. J. & Laskowski, K. L. The repeatability of behaviour: a meta-analysis. *Anim. Behav.* **77**, 771–783 (2009).
- Réale, D., Reader, S. M., Sol, D., McDougall, P. T. & Dingemanse, N. J. Integrating animal temperament within ecology and evolution. *Biol. Rev.* **82**, 291–318 (2007).
- Dingemanse, N. J., Both, C., Drent, P. J., van Oers, K. & van Noordwijk, A. J. Repeatability and heritability of exploratory behaviour in great tits from the wild. *Anim. Behav.* **64**, 929–938 (2002).
- Quinn, J. L., Patrick, S. C., Bouwhuis, S., Wilkin, T. A. & Sheldon, B. C. Heterogeneous selection on a heritable temperament trait in a variable environment. *J. Anim. Ecol.* **78**, 1203–1215 (2009).
- van Overveld, T. & Matthysen, E. Personality predicts spatial responses to food manipulations in free-ranging great tits (*Parus major*). *Biol. Lett.* **6**, 187–190 (2010).
- Wilson, A. D. M. & McLaughlin, R. L. Behavioural syndromes in brook charr, *Salvelinus fontinalis*: prey-search in the field corresponds with space use in novel laboratory situations. *Anim. Behav.* **74**, 689–698 (2007).
- Dingemanse, N. J., Both, C., van Noordwijk, A. J., Rutten, A. L. & Drent, P. J. Natal dispersal and personalities in great tits (*Parus major*). *Proc. R. Soc. Lond. B Biol. Sci.* **270**, 741–747 (2003).
- Cole, E. F. & Quinn, J. L. Personality and problem-solving performance explain competitive ability in the wild. *Proc. R. Soc. B Biol. Sci.* **279**, 1168–1175 (2012).
- Neuweiler, G. *The Biology of Bats* (Oxford University Press on Demand, 2000).
- Kunz, T. H. Roosting ecology of bats. In *Ecology of Bats* (ed. Kunz, T. H.) 1–55 (Springer, 1982).
- Thomas, J. A., Moss, C. F. & Vater, M. *Echolocation in Bats and Dolphins* (University of Chicago Press, 2004).

27. Knörnschild, M., Jung, K., Nagy, M., Metz, M. & Kalko, E. Bat echolocation calls facilitate social communication. *Proc. R. Soc. B Biol. Sci.* **279**, 4827–4835 (2012).
28. Schnitzler, H.-U., Moss, C. F. & Denzinger, A. From spatial orientation to food acquisition in echolocating bats. *Trends Ecol. Evol.* **18**, 386–394 (2003).
29. Yu, C., Luo, J., Wohlgenuth, M. & Moss, C. F. Echolocating bats inspect and discriminate landmark features to guide navigation. *J. Exp. Biol.* **222**, 191965 (2019).
30. Strelkov, P. P. Seasonal distribution of migratory bat species (Chiroptera, Vespertilionidae) in eastern Europe and adjacent territories: nursing area. *Myotis* **37**, 7–25 (2000).
31. Dietz, C., Von Helversen, O. & Nill, D. *Bats of Britain, Europe and Northeastern Africa* (2009).
32. Petersons, G. Seasonal migrations of north-eastern populations of Nathusius' bat *Pipistrellus nathusii* (Chiroptera). *Myotis* **41**, 29–56 (2004).
33. Alcalde, J. T. *et al.* Transcontinental 2200 km migration of a Nathusius' pipistrelle (*Pipistrellus nathusii*) across Europe. *Mammalia* **1**, (2020).
34. Seebacher, F. & Post, E. Climate change impacts on animal migration. *Clim. Change Responses* **2**, 5 (2015).
35. Voigt, C. C., Popa-Lisseanu, A. G., Niermann, I. & Kramer-Schadt, S. The catchment area of wind farms for European bats: a plea for international regulations. *Biol. Conserv.* **153**, 80–86 (2012).
36. Rydell, J. *et al.* Bat mortality at wind turbines in northwestern Europe. *Acta Chiropterol.* **12**, 261–274 (2010).
37. Sih, A., Bell, A. & Johnson, J. C. Behavioral syndromes: an ecological and evolutionary overview. *Trends Ecol. Evol.* **19**, 372–378 (2004).
38. Dall, S. R. & Johnstone, R. A. Managing uncertainty: information and insurance under the risk of starvation. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **357**, 1519–1526 (2002).
39. Geva-Sagiv, M., Las, L., Yovel, Y. & Ulanovsky, N. Spatial cognition in bats and rats: from sensory acquisition to multiscale maps and navigation. *Nat. Rev. Neurosci.* **16**, 94–108 (2015).
40. Wilson, A. D. M. & Godin, J.-G. J. Boldness and behavioral syndromes in the bluegill sunfish *Lepomis macrochirus*. *Behav. Ecol.* **20**, 231–237 (2009).
41. Videlier, M., Bonneaud, C. & Herrel, A. Intra-individual variation in exploration behaviour in a largely aquatic frog: effects of sex and personality traits. *Herpetol. J.* **28**, 10–18 (2018).
42. Verbeek, M. E. M., Drent, P. J. & Wiepkema, P. R. Consistent individual differences in early exploratory behaviour of male great tits. *Anim. Behav.* **48**, 1113–1121 (1994).
43. Guillette, L. M., Reddon, A. R., Hurd, P. L. & Sturdy, C. B. Exploration of a novel space is associated with individual differences in learning speed in black-capped chickadees *Poecile atricapillus*. *Behav. Processes.* **82**, 265–270 (2009).
44. Šlipogor, V., Oliveira, T. G., Tadić, Z., Massen, J. J. M. & Bugnyar, T. Consistent inter-individual differences in common marmosets (*Callithrix jacchus*) in boldness-shyness, stress-activity, and exploration-avoidance. *Am. J. Primatol.* **78**, 961–973 (2016).
45. Žampachová, B., Kaftanová, B., Šimánková, H., Landová, E. & Frynta, D. Consistent individual differences in standard exploration tasks in the black rat (*Rattus rattus*). *J. Comp. Psychol.* **131**, 150–162 (2017).
46. Drent, P. J., Verbeek, M. E. & Boon, A. Exploration, aggressive behaviour and dominance in pair-wise confrontations of juvenile male great tits. *Behaviour* **133**, 945–963 (1996).
47. Brown, C., Jones, F. & Braithwaite, V. A. Correlation between boldness and body mass in natural populations of the poeciliid *Brachyrhaphis episcopi*. *J. Fish Biol.* **71**, 1590–1601 (2007).
48. Lindecke, O., Elksne, A., Holland, R. A., Petersons, G. & Voigt, C. C. Orientation and flight behaviour identify the Soprano pipistrelle as a migratory bat species at the Baltic Sea coast. *J. Zool.* **308**, 56–65 (2019).
49. Marchetti, C. & Drent, P. J. Individual differences in the use of social information in foraging by captive great tits. *Anim. Behav.* **60**, 131–140 (2000).
50. DeRango, E. J. *et al.* Intraspecific variation in boldness and exploration shapes behavioral responses to stress in Galápagos sea lion pups. *Behav. Ecol. Sociobiol.* **73**, 166 (2019).
51. Sih, A. & Del Giudice, M. Linking behavioural syndromes and cognition: a behavioural ecology perspective. *Philos. Trans. R. Soc. B Biol. Sci.* **367**, 2762–2772 (2012).
52. Griffin, A. S., Guillette, L. M. & Healy, S. D. Cognition and personality: an analysis of an emerging field. *Trends Ecol. Evol.* **30**, 207–214 (2015).
53. Bebus, S. E., Small, T. W., Jones, B. C., Elderbrock, E. K. & Schoech, S. J. Associative learning is inversely related to reversal learning and varies with nestling corticosterone exposure. *Anim. Behav.* **111**, 251–260 (2016).
54. Mazza, V. *et al.* Coping with style: individual differences in responses to environmental variation. *Behav. Ecol. Sociobiol.* **73**, 142 (2019).
55. Roeleke, M., Blohm, T., Kramer-Schadt, S., Yovel, Y. & Voigt, C. C. Habitat use of bats in relation to wind turbines revealed by GPS tracking. *Sci. Rep.* **6**, 28961 (2016).
56. Lewis, S. E. Roost Fidelity of Bats: A Review. *J. Mammal.* **76**, 481–496 (1995).
57. Mitchinson, B., Martin, C. J., Grant, R. A. & Prescott, T. J. Feedback control in active sensing: rat exploratory whisking is modulated by environmental contact. *Proc. R. Soc. B Biol. Sci.* **274**, 1035–1041 (2007).
58. Mathot, K. J. & Dingemans, N. J. Plasticity and personality. In *Integrative Organismal Biology* (eds Martin, L. B. *et al.*) 55–69 (Wiley Online Library, 2015).
59. Charmantier, A. *et al.* Adaptive phenotypic plasticity in response to climate change in a wild bird population. *Science* **320**, 800–803 (2008).
60. Gross, K., Pasinelli, G. & Kunc, H. Behavioral plasticity allows short-term adjustment to a novel environment. *Am. Nat.* **176**, 456–464 (2015).
61. Rodríguez-Prieto, I., Martín, J. & Fernández-Juricic, E. Individual variation in behavioural plasticity: direct and indirect effects of boldness, exploration and sociability on habituation to predators in lizards. *Proc. R. Soc. B Biol. Sci.* **278**, 266–273 (2011).
62. Currie, S. E., Boonman, A., Troxell, S., Yovel, Y. & Voigt, C. C. Echolocation at high intensity imposes metabolic costs on flying bats. *Nat. Ecol. Evol.* **4**, 1174–1177 (2020).
63. Rankin, C. H. *et al.* Habituation revisited: an updated and revised description of the behavioral characteristics of habituation. *Neurobiol. Learn. Mem.* **92**, 135–138 (2009).
64. Vernouillet, A. & Kelly, D. M. Individual exploratory responses are not repeatable across time or context for four species of food-storing corvid. *Sci. Rep.* **10**, 1–11 (2020).
65. Daalmans, J. Habituation. In *Human Behavior in Hazardous Situations* (Elsevier, 2013).
66. Dingemans, N. J. *et al.* Variation in personality and behavioural plasticity across four populations of the great tit *Parus major*: Population variation in personality and behavioural plasticity. *J. Anim. Ecol.* **81**, 116–126 (2012).
67. Mettke-Hofmann, C., Rowe, K. C., Hayden, T. J. & Canoine, V. Effects of experience and object complexity on exploration in garden warblers (*Sylvia borin*). *J. Zool.* **268**, 405–413 (2006).
68. Martin, J. G. A. & Réale, D. Temperament, risk assessment and habituation to novelty in eastern chipmunks *Tamias striatus*. *Anim. Behav.* **75**, 309–318 (2008).
69. Peeke, H. V. S. & Bell, A. M. Individual variation in habituation: behaviour over time toward different stimuli in threespine sticklebacks (*Gasterosteus aculeatus*). *Behaviour* **149**, 1339–1365 (2012).

70. Dingemanse, N. J., Kazem, A. J. N., Réale, D. & Wright, J. Behavioural reaction norms: animal personality meets individual plasticity. *Trends Ecol. Evol.* **25**, 81–89 (2010).
71. Nachev, V. & Winter, Y. Behavioral repeatability and choice performance in wild free-flying nectarivorous bats (*Glossophaga commissarisi*). *Behav. Ecol. Sociobiol.* **73**, 24 (2019).
72. Kerches-Rogeri, P., Niebuhr, B. B., Muylaert, R. L. & Mello, M. A. R. Individual specialization in the use of space by frugivorous bats. *J. Anim. Ecol.* **89**, 2584–2595 (2020).
73. Muylaert, R. L., Matos, D. M. D. S. & Mello, M. A. R. Interindividual variations in fruit preferences of the yellow-shouldered bat *Sturnira lilium* (Chiroptera: Phyllostomidae) in a cafeteria experiment. *Mammalia* **78**, 93–101 (2014).
74. Lemke, T. O. Foraging ecology of the long-nosed bat, *Glossophaga soricina*, with respect to resource availability. *Ecology* **65**, 538–548 (1984).
75. Cryan, P. M., Stricker, C. A. & Wunder, M. B. Evidence of cryptic individual specialization in an opportunistic insectivorous bat. *J. Mammal.* **93**, 381–389 (2012).
76. Lehnert, L. S. *et al.* Variability and repeatability of noctule bat migration in Central Europe: evidence for partial and differential migration. *Proc. R. Soc. B Biol. Sci.* **285**, 20182174 (2018).
77. Magalhães de Oliveira, H. F. *et al.* Wing morphology predicts individual niche specialization in *Pteronotus mesoamericanus* (Mammalia: Chiroptera). *PLoS ONE* **15**, e0232601 (2020).
78. Menzies, A. K., Timonin, M. E., McGuire, L. P. & Willis, C. K. R. Personality variation in little brown bats. *PLoS ONE* **8**, e80230 (2013).
79. Boyer, A. M. Personality and Behavioral Syndromes of the Big Brown Bat, *Eptesicus fuscus*. (2018).
80. Webber, Q. M. R., McGuire, L. P., Smith, S. B. & Willis, C. K. R. Host behaviour, age and sex correlate with ectoparasite prevalence and intensity in a colonial mammal, the little brown bat. *Behaviour* **152**, 83–105 (2015).
81. Hillen, J. *et al.* Sex-specific habitat selection in an edge habitat specialist, the Western Barbastelle Bat. *Ann. Zool. Fenn.* **48**, 180–190 (2011).
82. Ruczyński, I., Nicholls, B., MacLeod, C. D. & Racey, P. A. Selection of roosting habitats by *Nyctalus noctula* and *Nyctalus leisleri* in Białowieża forest—adaptive response to forest management?. *For. Ecol. Manag.* **259**, 1633–1641 (2010).
83. Ruczyński, I. & Bogdanowicz, W. Summer roost selection by tree-dwelling bats *Nyctalus noctula* and *N. leisleri*: a multiscale analysis. *J. Mammal.* **89**, 942–951 (2008).
84. Ruczyński, I. & Bogdanowicz, W. Roost cavity selection by *Nyctalus noctula* and *N. leisleri* (Vespertilionidae, Chiroptera) in Białowieża Primeval Forest, Eastern Poland. *J. Mammal.* **86**, 921–930 (2005).
85. Hutson, A. M. & Mickleburgh, S. P. *Microchiropteran Bats: Global Status Survey and Conservation Action Plan*. vol. 56 (IUCN, 2001).
86. Food and Agriculture Organization of the United Nations. *State of the world's Forests 2011*. (Food and Agriculture Organization of the United Nations, 2011).
87. Erickson, J. L. & West, S. D. Associations of bats with local structure and landscape features of forested stands in western Oregon and Washington. *Biol. Conserv.* **109**, 95–102 (2003).
88. Ruczyński, I. & Siemers, B. M. Hibernation does not affect memory retention in bats. *Biol. Lett.* **7**, 153–155 (2011).
89. Werner, E. E., Mittelbach, G. G. & Hall, D. J. The role of foraging profitability and experience in habitat use by the bluegill sunfish. *Ecology* **62**, 116–125 (1981).
90. Hutterer, R. *Bat Migrations in Europe: a Review of Banding Data and Literature*. vol. 28 (Federal Agency for Nature Conservation, 2005).
91. Jahelková, H. & Horáček, I. Mating system of a migratory bat, Nathusius' Pipistrelle (*Pipistrellus nathusii*): different male strategies. *Acta Chiropterol.* **13**, 123–137 (2011).
92. Lindecke, O., Voigt, C. C., Petersons, G. & Holland, R. A. Polarized skylight does not calibrate the compass system of a migratory bat. *Biol. Lett.* **11**, 20150525 (2015).
93. Rojas-Ferrer, I., Thompson, M. J. & Morand-Ferron, J. Is exploration a metric for information gathering? Attraction to novelty and plasticity in black-capped chickadees. *Ethology* **126**, 383–392 (2020).
94. Barclay, R. M. *et al.* Can external radiotransmitters be used to assess body temperature and torpor in bats?. *J. Mammal.* **77**, 1102–1106 (1996).
95. Friard, O. & Gamba, M. BORIS: a free, versatile open-source event-logging software for video/audio coding and live observations. *Methods Ecol. Evol.* **7**, 1325–1330 (2016).
96. R Core Team. *R: A Language and Environment for Statistical Computing, Version 4.0.2*. Vienna, Austria: R Foundation for Statistical Computing. (2020).
97. Bates, D., Mächler, M., Bolker, B. & Walker, S. Fitting linear mixed-effects models using lme4. *arXiv.org. J. Stat. Softw.* **67**, 1–48 (2015).
98. Lüdecke, D., Makowski, D., Waggoner, P. & Patil, I. Performance: Assessment of Regression Models Performance [R package]. CRAN (2020).
99. Stoffel, M. A., Nakagawa, S., Schielzeth, H. & Goslee, S. rptR: repeatability estimation and variance decomposition by generalized linear mixed-effects models. *Methods Ecol. Evol.* **8**, 1639–1644 (2017).
100. Lenth, R. V. *et al.* *emmeans: Estimated Marginal Means, aka Least-Squares Means* (2021).
101. Wickham, H. *ggplot2: Elegant Graphics for Data Analysis* (Springer, 2016).

Acknowledgements

We are grateful to the many fieldworkers of the Pape Ornithological Station. We thank Shannon Currie for assistance and expert advice during the fieldwork. L.S. was funded by a Humboldt Research Fellowship for Postdoctoral Researchers (Ref 3.3 – NLD – 1192631 - HFST-P) awarded by the Alexander von Humboldt-Stiftung. We thank Rebecca Scheibke for her illustration of the novel environment assay and Gretchen Wagner for proof-reading the manuscript.

Author contributions

L.S., O.L., T.S., and C.C.V. conceived the ideas. L.S., O.L., G.P., and C.C.V. provided materials and infrastructure. L.S., O.L., L.M., G.P., and C.C.V. assisted in data collection. T.S. and S.R. quantified the behavioral responses. T.S. and L.S. conducted the statistical analyses and led the writing. All authors evaluated and approved the final draft of the article.

Funding

Open Access funding enabled and organized by Projekt DEAL.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1038/s41598-021-87588-y>.

Correspondence and requests for materials should be addressed to L.S.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2021