

Microplastics alter multiple biological processes of marine benthic fauna

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1 Microplastics alter multiple biological processes of marine benthic fauna 2 3 Running page head: Microplastic impacts on benthic fauna. 4 5 Victoria G. Mason, Martin W. Skov, Jan Geert Hiddink, Mark Walton 6 7 School of Ocean Sciences, Bangor University, Isle of Anglesey. LL59 5AB. UK. 8 9 Email: torimason@hotmail.co.uk ABSTRACT 10

11 Marine sediments are a sink for microplastics, making seabed organisms particularly

12 exposed. We used meta-analysis to reveal general patterns in a surge in experimental studies 13 and to test for microplastic impact on biological processes including invertebrate feeding, 14 survival and energetics. Using Hedge's effect size (g), which assesses the mean response of 15 organisms exposed to microplastics compared to control groups, we found negative impacts 16 (significant negative g values) across all life stages (overall effect size (g) = -0.57 95% CI [-17 0.76, -0.38]), with embryos most strongly affected (g = -1.47 [-2.21, -0.74]). Six of seven 18 biological process rates were negatively impacted by microplastic exposure, including 19 development, reproduction, growth and feeding. Survival strongly decreased (g = -0.69 [-20 1.21, -0.17]), likely due to cumulative effects on other processes such as feeding and growth. 21 Among feeding habits, omnivores and deposit feeders were most negatively impacted (g = -0.93 [-1.69, -0.16] and -0.92 [-1.53, -0.31], respectively). The study incorporated the first 22 23 meta-analysis to contrast the effects of leachates, virgin, aged and contaminated particles. 24 Exposure to leachates had by far the strongest negative effects (g = -0.93 [-1.35, -0.51]), showing studies of contaminants and leachates are critical to future research. Overall, our 25

meta-analysis reveals stronger and more consistent negative impacts of microplastics on seabed invertebrates than recorded for other marine biota. Seabed invertebrates are numerous and diverse, and crucial to bottom-up processes, including nutrient remineralisation, benthopelagic coupling and energy transfer through the ocean food web. Marine sediments will store microplastics over long timescales. The reveal that microplastics impinge on multiple fundamental biological processes of seabed fauna implies plastic pollution could have significant and enduring effects on the functioning of the ocean.

33 Key Words

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Systematic review • Benthos • Functional traits • Survival • Development • Meta-analysis 34

1.0 INTRODUCTION

The problem of plastic pollution is growing, resulting from an average annual increase of 9% in plastic manufacturing between 1950 and 2009 (Hammer et al. 2012). The input of plastics into the marine environment, both directly and indirectly through riverine inputs, is also increasing. An estimated 4.7 to 12.8 million tonnes of plastic enters the marine environment every year (Agamuthu et al. 2019). The fate of much of this plastic is unknown; the term 'missing plastic' was coined to describe the shortfall in the estimated volume of plastics found in the water column compared to inputs (Wayman & Niemann 2021). It is thought that deep-water and sediment storage of plastics and microplastics, in particular, make up the majority of this 'missing plastic' (Zhang 2017). Here, we assess the impact of accruing microplastics on invertebrate animals of the seafloor.

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The definition of microplastics is inconsistent throughout the existing literature, but most commonly includes plastic particles of any shape from 0.1 µm to 5mm (Auta et al. 2017). Within this category exist intentionally manufactured primary microplastics, such as highly 50 prevalent pre-production plastic 'nurdles' (Jiang et al. 2021), as well as secondary 51 microplastics, resulting from the UV or physical degradation of marine macroplastics 52 (Efimova et al. 2018). Microplastic prevalence in the ocean was recently estimated at 2.41 53 million tonnes across the Atlantic, Indian and Pacific subtropical gyres (Vazquez & Rahman 54 2021). This prevalence is likely to increase with inputs not only from terrestrial activity, but 55 also from the breakdown of plastics already present in the marine environment (Kooi et al. 56 2017). Microplastics are subject to further change upon entering the marine environment; 57 they may be further broken down into nanoplastic particles (<0.1µm) or experience 58 biofouling (Zhang 2017). Biofouling of microplastics occurs predominantly as a result of the 59 attraction of organic substances to the hydrophobic surface of the particle (Kaiser et al. 60 2017). Cózar et al. (2014) showed that the specific density of most microplastics is lower 61 than that of seawater, so particles should remain buoyant. However, settling of microplastics 62 on the seafloor has been documented, with Zhang (2017) suggesting sinking rates of 63 approximately 4mm per day. Sinking is stimulated by the biofouling of microplastic particles 64 which increases the specific density, although studies have also suggested the influence of 65 microplastic shape and size on the sinking rate of a particle (Melkebeke et al. 2020). Using 66 Environmental Risk Assessment modelling, Everaert et al. (2018) found species had varying sensitivity to microplastic, but that sediment concentrations <540 microplastic particles kg⁻¹ 67 68 were 'safe' and unlikely to have negative impact. The same study reported a current 69 concentration of 32-144 particles kg⁻¹ in marine intertidal sediments, suggesting that the safe 70 threshold is likely to be exceeded in the latter half of the 21st century. Estimates of 71 microplastics in seawater itself vary widely and Xu et al. (2020) reported seawater 72 concentrations ranging from 0.33 to 3252 particles m⁻³ globally. The vast majority (>90%) of 73 marine microplastics have been reported to accumulate on the seafloor (Melkebeke et al. 74 2020). In the southern North Sea, for example, sediment microplastics have been reported to

range in concentration from 2.8 to 1188.8 particles kg⁻¹ dry weight (Lorenz et al. 2019).

Microplastics are therefore likely to become a ubiquitous component of seabed sediments and

thus the influence of microplastics on benthic habitats must be considered.

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Gall and Thompson (2015) reported over 44000 interactions of marine fauna with plastic debris, across 693 species. Larger plastic fragments impact fauna predominantly through ingestion and entanglement. A systematic review of 747 studies quantifying the interactions of plastics with marine megafauna found 701 species had ingested plastics and 354 species had experienced entanglement (Kühn & van Freneker 2020). Microplastics can impact marine organisms through a wider range of mechanisms, as shown in many experimental laboratory studies. Microplastic exposure caused abnormal embryo development in the brown mussel Perna perna (Gandara e Silva et al. 2016). The lugworm Arenicola marina reduced its feeding rate with increasing microplastic dosage (Besseling et al. 2013). Reduced feeding can be the result of a false sense of fullness, damage or blockages to the digestive tract or confusing microplastics for prey (de Sá et al. 2015). Numerous studies have found cellular level impacts of microplastics, for example, microplastic consumption influenced cellular pathway signalling, diminished growth and induced toxicity and oxidative stress in rotifers (Jeong et al. 2016). Such impacts may lead to behavioural changes, growth inhibition and, ultimately, increased mortality (de Sá et al. 2018). Microplastics also have the potential to cause adverse reactions via persistent organic pollutants (POPs), which adhere to plastic particles. Particularly hazardous are endocrine disruptor chemicals (EDCs), which accumulate in fatty tissues, altering hormone production and potentially causing thyroid problems, reduced reproductive success and hormone-sensitive cancers (Gallo et al. 2018). While the study of POPs so far has focussed primarily on the impacts on human health, effects on marine fauna have been observed, one example being reduced survival rate and

jump height in beach hoppers (Tosetto et al. 2016). Microplastics encountered in nature are often contaminated, giving them the potential to be more toxic than virgin microplastics. Abnormal development was found in 23% of brown mussel *Perna perna* embryos from virgin pellets compared to 100% abnormal development from pellets sourced from beach sediments (Gandara e Silva et al. 2016). Despite such indications of impact to benthic organisms, there is no overview of implications to the breadth of seabed organisms. In a systematic review of 220 studies published prior to the year 2010, Ajith et al. (2020) found that 38% of existing studies on the impacts of microplastics had used fish as the study organism, followed by 18% studies targeting molluscs. This leaves a knowledge gap surrounding the majority of benthic invertebrate species. Here, we make use of a rapid increase in publications on marine benthos since 2019 (Figure S2, Supplementary Materials) and new data for a total of 6 taxa to generate a comprehensive overview across seabed taxonomic and functional groups.

As a means of quantifying the impacts of microplastics on marine fauna, recent studies have considered the impact of microplastics on what was termed the 'functional traits' of organisms (Berlino et al. 2021, Salerno et al. 2021), albeit several 'traits' are more correctly perceived as the rates of important biological processes like growth, reproduction and survival (See Supplementary Materials, Table S1). Focusing on biological rates offers insights into the impacts of microplastics on wider organismal and ecosystem functioning. Since many impacts of microplastics result directly from the ingestion of particles, feeding strategy in particular may contribute to variation in the magnitude of impacts. Thus, among fish and invertebrates, predators and deposit feeders contained more plastics than filter feeders and, sometimes, deposit feeders (Bour et al. 2018, Naji et al. 2018). It stands to

reason that if the ingestion of microplastics varies by feeding strategy, so might the effects on biological processes.

There is a lack of consensus of the impacts of microplastics on marine benthic fauna, particularly in terms of the range of factors which might be contributing to the variation in effects. Here, we make use of a rapid increase in publications on marine benthos since 2019 with new data for a total of 6 phyla to generate a comprehensive overview of the impacts of microplastics across seabed taxonomic and functional groups. Using a systematic review and associated meta-analysis of extracted data we quantify the impacts of microplastics on marine benthic fauna and identify knowledge gaps and potential bias in the current state of the art. We hypothesised that microplastics would have an overall negative effect on the performance of marine benthic fauna, which would increase with exposure concentration. We expected the effects of microplastics to vary amongst feeding habits, with predators at risk of stronger effects resulting from trophic transfer of microplastic particles. Microplastic characteristics, including size, shape, exposure duration and concentration, were expected to be primary drivers of any variation in effect size.

2.0 MATERIALS AND METHODS

2.1 STUDY DESIGN

The study used a systematic review and meta-analysis to assess the impacts of experimental exposure to microplastics particles (hereinafter, MPP will refer to microplastic particles) on marine benthic fauna. Only laboratory studies that included a control (no MPP) and one or more MPP exposure levels were included, so that overall mean effect sizes could be determined. Studies focusing on MPP ingestion but not impacts on biological processes were

excluded. The review had no geographical or temporal limits. Two search engines, Web of Science and the Wiley online library, were used in order to include papers from a range of sources, including grey literature, and minimise publication bias otherwise arising from restricting search results to peer-reviewed journals favouring studies with significant results (Sterne et al. 2000). Ultimately, all studies included in the analysis were from peer-reviewed journals. The study considered the influence of potential contributing factors, such as phylum, feeding strategy and microplastic composition, on variation in the magnitude of microplastic impacts (see Supplementary Materials, Figure S1).

2.2 LITERATURE SEARCH AND DATA EXTRACTION

The systematic literature search was conducted on the 7th June 2021, following the methodology of Pullin and Stewart (2006) and O'Dea et al. (2021). The search string had three components using the Boolean operators "AND" and "OR". Each component of the string was designed to address an area (impact, microplastics or biological processes) of the research question and to include studies on any marine benthic fauna. The string of search terms was tested to ensure it delivered relevant literature hits (tested using 10 pre-identified highly relevant key references. Table S2). The final string of search terms was as follows:

impact* OR response* OR effect* OR interaction* OR consequence* OR implication* OR contamination* OR ingestion* OR consumption* OR consume* OR uptake* OR "taken up"

OR accumulation OR contamination OR transfer

169 AND

Microplastic* OR "micro plastic" OR "micro-plastic" OR microfilament* OR filament* OR "plastic pellet*" OR nurdle*

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trait* OR "functional trait*" OR growth OR feeding OR reproduction OR fecundity OR behaviour* OR development OR hatching OR health OR survival OR digestion

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A total of 3,650 search results (studies, papers) were identified on Web of Science, with a further 166 from the secondary Wiley Online Library. For each paper, the title, then abstract and then the full-text content were screened for relevance (Table S3) according to the following criteria. Studies that purely addressed the distribution or sources of microplastics were excluded, as were observational work documenting only the ingestion of microplastics, qualitative and systematic reviews. Changes to feeding rates following microplastic consumption were included, but ingestion rates of microplastic particles themselves were not included as a change to a biological process. Experimental studies with a focus on cellular impacts were also excluded, unless the impact could be tied directly to one of the biological processes we evaluated (e.g. O₂ consumption, representing respiration and energy demand). Only studies focussing on marine benthic organisms were considered. Freshwater organisms or those from inland saltwater were excluded, while both intertidal and subtidal marine and estuarine organisms were included, where the species was determined to spend the majority of its lifecycle on, or buried within, the seafloor. Experiments which used microplastics of sizes outside of the predetermined range (0.1µm - 5mm) were excluded. A final list of 72 papers (Table S3) was selected for meta-analysis.

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Data were extracted directly from paper text, tables and figures, the latter using Automeris WebPlotDigitizer Version 4.4. Types of data extracted from each study were study identifiers, meta-data and data for quantitative synthesis (control and experimental mean, standard deviation, SD, and number of replicates, n (Table S4). Examples of response variables which were used for biological traits are outlined in Table S1. A total of 701 case

studies (independent experiments included in the same study. For example, multiple exposure concentrations or species tested) were extracted from the 72 papers.

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2.3 DATA ANALYSIS

Data extracted from papers required standardisation before analysis to overcome the use of different units and approaches among studies. The data were standardised to common units of microplastic exposure concentration, duration and particle size in order to allow comparison of the experimental conditions that test animals were exposed to. Microplastic particles were classified into: fibre, fluff (usually derived from clothing fibres), fragment, pellet, square, sphere (including microbeads) or powder, plus leachates and leachates adsorbed to microplastics, according to how they were described by the authors (see e.g. Gray and Weinstein 2017). Microplastic exposure units which could not be standardised into common units (e.g. % sediment weight, fibres per prey individual) were excluded from concentration analysis (18 studies). Remaining microplastic exposure units from 54 studies were standardised into common units of g L⁻¹. Concentrations given in particles L⁻¹ were converted using mass_{particle} = density × volume (Everaert et al. 2018), using a standard density of marine microplastics of 0.925g cm⁻³, determined by Van Cauwenberghe (2016). Density of plastic particles was not available for the microplastics used in most studies and using this standard density was the most appropriate approach. Particle volumes were calculated for spheres (and for fragments, with assumptions of largely spherical shape) using $V = 4/3\pi r^3$, where the radius of the particle was provided in the original study. Microplastic concentration was log transformed for analysis to allow patterns to be more clearly seen, since data were skewed towards very small values. Where necessary, medians and interquartile ranges (IQR) were converted into means and standard deviations (SD), where SD was taken to equal IQR/1.35, assuming normal distribution of data (Higgins et al. 2019). Any 95% confidence intervals

(CI) were also converted into SD, where SD=CI/3.92, multiplied by the square root of the sample size (n) (Higgins et al. 2019). Data were explored for patterns in the number of studies per geographical region, taxa (phylum of organism), feeding strategy (predator, deposit feeder, scavenger, filter feeder, omnivore) and microplastic characteristic (shape, size, polymer type) to generate an overview of the geographical distribution of research and to identify potential bias within the results, such as a high proportion of studies published in one geographic region.

Effect size for each study was calculated as Hedge's g (Borenstein et al. 2009):

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$$Hedge's g = \frac{m_{c-}m_e}{SD_{pooled}} \times J$$

Where m_c was the control mean, m_e was the experimental mean, SD_{pooled} was the pooled standard deviation across the samples and J was the correction factor used to account for bias arising from variation in sample size.

Hedge's g values were interpreted using the recommended thresholds from Cohen (2013), where \sim 0.2 indicated a small effect, \sim 0.5 indicated a moderate effect and >0.8 indicated a larger effect. A negative Hedge's g represents a negative impact of the experimental condition relative to the mean. Directionality of effect sizes was corrected to ensure g values were representative of the effects shown by studies and as described by the authors (Table S5). For example, an increased time to find a new shell (automatically a positive effect) was corrected to be negative, when the authors noted this represented a negative impact on the organism (Crump et al. 2020). We checked for any influence of publication bias by applying the non-parametric trim and fill method (Duval and Tweedie 2000) to an rma.uni model of

our data, whereby the number of missing studies at either extreme positive or negative values could be estimated. This showed that publication bias was likely to have had a negligible effect on the outcome of our meta-analysis (Table S6).

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Once an effect size had been determined for each case study (k = 701, where k signifies independent experiments, or case studies, included in the same study), a pooled effect size was calculated for all values and each biological process, using a random effects model with the "rma.mv" function of the "metafor" package (Viechtbauer 2010) in Rstudio Version 1.3.1093 (Rstudio Team 2020). In each model, we included 'Study ID' of the published study to account for non-independence of data extracted from the same study (Viechtbauer 2007). To evaluate data compliance with test assumptions, an I² value was produced by Wald's test for heterogeneity of variance between studies (Borenstein et al. 2009) and a Cochran's Q value determined the level and significance of heterogeneity (Cochran 1954). Since results from the random effects model indicated significant heterogeneity between studies, subgroup analyses (categorical data) and meta-regressions (continuous data) were conducted using random effects models in metafor (R statistics) to identify moderator variables which may have been driving the variation. Organism traits such as taxa, feeding type and life stage and experimental variables including microplastic size, shape, polymer type and concentration, were investigated for contribution to heterogeneity as well as the pooled effect size for each variable. Effect sizes were given with 95% confidence intervals.

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3.0 RESULTS

3.1 SUMMARY AND DISTRIBUTION OF FINDINGS

While no temporal limits of publication were implemented, all papers were published from 2013 onwards, with 79.2% published since 2018 and nearly half (43.1%) published in the last

1.5 years covered by our systematic review (Figure S2). Published findings were from 6 continents, leaving only Antarctica absent, with the most research having occurred in Europe (n = 35) and Asia (n = 17) (Figure S3).

Experiments involved 6 animal phyla and 6 feeding strategies (Figure 1a), with the majority of studies focused on filter feeders (n = 39). A wide range of experimental conditions were used by studies. Exposure concentrations were reported in a multitude of units, of which 'g l' and 'particles l'' were the most common, with less frequently used units including '% of feed' and '% of sediment weight'. Approaches to reporting microplastic leachates were varied, since some studies used leachates adsorbed to particles and others used leachates independently (reported as concentration in the water column). The majority of studies (n = 26) exposed organisms to microplastic spheres, although 30 studies did not state the shape of particles (Figure 1b). Out of 19 types and combinations of polymers used for exposure, polystyrene and polyethylene were the most commonly used (n = 25 and n = 10, respectively).

3.2 EFFECTS OF MICROPLASTICS ON BIOLOGICAL PROCESSES

The effect size for all organisms pooled indicated a moderate, but significant overall negative effect of microplastics on biological processes (g = -0.57 [-0.76, -0.38], p < 0.001) (Figure 2). Significant negative effects were also seen for all categories of biological processes, except energy use (e.g. respiration). Large negative effects of microplastic particles (MPP) on animal development, reproduction and survival were seen (Figure 2). A small and non-significant effect of MPP on energy processes was found. Significant heterogeneity of variance was found between studies ($I^2 = 61.4\%$, $Q_{700} = 2668.9$, p < 0.001), including within every biological process category (Table 1), indicating unexplained variance beyond the

effect of biological process and supporting the need for a sub-group analysis to investigate other drivers of effect size.

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3.3 SUB-GROUP ANALYSIS

3.3.1 Organism Characteristics. The taxonomic group of organisms explained a significant amount of heterogeneity of variance in the dataset ($Q_{\text{moderators}, 6} = 39.87, p < 0.001$). Microplastic exposure had a large and significantly negative effect on all phyla, with chordates (ascidians) most significantly affected (g = -1.79 [-3.47, -0.12], p = 0.04), although this result originated from only one study (Anderson and Shenkar 2021). Echinoderms, crustaceans and molluscs were less, but still significantly, impacted by microplastic exposure, while impacts on annelids and cnidarians were not significant (Figure 3). Species-level effects were also statistically significant ($Q_{\text{moderators}, 61} = 160.81, p < 0.001$). The greatest negative plastics effect on a single species was in the sea urchin Lytechinus variegatus (g = -11.57[-16.21, -6.92], p < 0.001, k = 2), followed by the coral Acropora formosa (g = -4.67)7.22, -2.11], p < 0.001, k = 5). Feeding strategy of the organism contributed significantly to heterogeneity between studies $(Q_{\text{moderators, 6}} = 42.15, p < 0.001)$ (Figure 4). Omnivores and deposit feeders experienced the largest negative effects from MPP (g = -0.93 [-1.69, -0.16] and -0.92 [-1.53, -0.31], respectively), while all other feeding strategies except scavengers were also negatively impacted (Figure 4). Every life stage of organism was significantly negatively impacted by MPP, with earlier life stages most strongly affected, particularly embryos (g = -1.47 [-2.21, -0.74], p < 0.001) (Figure 5).

3.3.2 Microplastic exposure. Microplastic exposure concentration ranged from 1.21×10^{-11} to 1000 g L^{-1} (median = $4.84 \times 10^{-4} \text{ g L}^{-1}$) but did not contribute significantly to between-study heterogeneity ($R^2 = 0.99$, $Q_{moderators, 1} = 0.0077$, p = 0.93, Figure S4). However, analysis of the distribution of data showed higher variability in effect sizes at higher concentrations, particularly for fragments (Figure 6).

The duration for which organisms were exposed to microplastics ranged from 0.17 to 5760 hours, with a median duration of 120 hours. Meta-regression showed that duration of exposure to microplastics did not explain a significant amount of heterogeneity (R^2 = 0.02, $Q_{moderator, 1}$ = 0.13, p = 0.72) (Figure S5a) and the size of microplastic particle did not contribute to variation in effect size (R^2 = 0.10, $Q_{moderator, 1}$ = 0.08, p = 0.77) (Figure S5b), although the effects of nanoparticles (<0.1 μ m) were not explored in this study.

Microplastic shape accounted for significant heterogeneity in the data (mixed-effect modelling: $Q_{\text{moderators, }10} = 47.10$, p < 0.001), although there was significant residual heterogeneity ($Q_{residual, 691} = 2543.67$, p < 0.001) (Figure 7). Microplastic fibres, fragments, leachates and spheres had significant negative effects (Figure 7). Effects driven by microplastic fluff, leachates adsorbed onto microplastics, pellets, powders and squares were not significant, although there were only 3 effect sizes of leachates adsorbed to particles, all from one study (Gu et al. 2020). The most negative significant effect resulted from leachates (no longer adsorbed onto microplastics) (g = -0.93 [-1.35, -0.51], p < 0.001), followed by fragments (g = -0.70 [-1.14, -0.26], p < 0.001).

From all exposure conditions analysed (MPP concentration, size, shape, exposure duration and polymer type), polymer type contributed the most to between-study heterogeneity

(Q_{moderators, 19} = 68.93, p < 0.001) (Figure S6). Polybrominated biphenyl ether had the most
 negative significant effect (g = -4.69 [-6.88, -2.51], p < 0.001) (Figure S6).

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4.0 DISCUSSION

4.1 BIOLOGICAL PROCESSES

This study offers the strongest and most consistent evidence to date of an overrridingly negative impact of microplastics on marine invertebrates. We found highly significant negative effects of microplastics on the biological process rates of marine benthic fauna. Every life stage was negatively impacted, with the strongest effects on early life stages, especially embryos. There were negative impacts on six out of seven fundamental biological processes including survival, development, reproduction, growth and feeding. Among feeding habits, omnivores and deposit feeders were particularly hard hit. Our study differs from previous reviews in that it documents substantially stronger and more consistently negative impacts of microplastics on a much greater variety of animal life-processes. For instance, Foley et al. (2018) described more neutral than negative effects of microplastics on growth, consumption, reproduction on the survival of fishes and aquatic invertebrates. Previous studies differed in focal organisms from the present study by including freshwater species or fishes (Foley et al. 2018, Salerno et al. 2021, Berlino et al. 2021). Yet, the primary cause for greater predominance of negative impact in the present meta-analysis is likely that the rapid increase in experimental studies over the past two years has offered greater statistical power for detecting the impacts of microplastics on marine animals; the present study synthesised data from 72 studies compared to 41 studies in the most recent previous review (Berlino et al. 2021). Certainly, the documentation of negative impacts has become more frequent in recent reviews (Foley et al. 2018, Salerno et al. 2020, Berlino et al. 2021).

Our findings of stronger impacts on benthic organisms compared to pelagic and freshwater organisms emphasises the need to improve research efforts in this area.

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The reveal that multiple organismal processes and traits are affected by plastics is not surprising. The biological rates of an organism are intrinsically linked and it is unlikely that the effects of microplastics would act independently on each of these. Figure 8 explores this principle of interlinkages: commencing with the process of feeding, which can be impacted by microplastics as a result of intestinal blockages, false sense of fullness or confusion with prey (Cole et al. 2011), reduced feeding will limit energy availability for morphological change, gonad development and movement. The suppression of feeding indirectly affects somatic growth, development and reproduction (Foley et al. 2018, Salerno et al. 2021), in addition to direct cellular effects or other growth altering processes such as tissue incorporation (Hierl et al. 2021). The observation that survival was significantly negatively impacted indicates a synergistic effect of plastics on the organism as a whole, wherein the impacts on different processes interact to create a larger combined effect than expected from the sum of individual impacts (Figure 8). Energy was the only response not significantly impacted by microplastic exposure, which may in part be due to the methodological difficulties in ascribing effects on energetic processes as either positive or negative (Table S5).

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4.2 ORGANISM CHARACTERISTICS

Effects of microplastics on benthic taxonomic groups were universally negative, although not significant for annelids and cnidarians. Across multiple taxonomic groups, a reduction in growth was documented, most likely the result of reduced energy reserves as reported by Wright et al. 2013. In that study, a range of exposure concentrations were used, up to 5%

sediment weight. This is likely to be higher than environmentally realistic concentrations of microplastics, perhaps causing more extreme impacts. However, impacts on growth have been seen more widely; previously, 58.8% of nematodes were shown to suffer energy loss from consuming microplastic particles, particularly fibres (Hodgson 2018). Growth inhibition may also have resulted from changes in cellular activity (Prinz & Korez 2020), for instance through cellular modifications (e.g. penetration of microplastics into cell structures) and oxidative stress, although this study focused on organismal level processes rather than cellular. Further research into cellular level effects is therefore strongly recommended.

For several species the strength of impact can be explained by the life stage investigated, although it was not possible to fully disentangle the effects of life stage from species through meta-analysis. The effects of microplastics tends to increase with decrease in organismal size (Salerno et al. 2021), with earlier life stages (gametes, embryos, larvae and juveniles) more severely affected than adults, as recorded here. Thus, the strongest negative effects we recorded were for the larvae of the sea urchin *Lytechinus variegatus*, where abnormal development increased 58.1-66.5% after microplastic exposure (Nobre et al. 2015). Smaller invertebrates are often numerous and crucial to bottom-up processes in natural ecosystems. Their study is therefore particularly important to predicting the influences of plastic pollution on whole-ecosystem functioning.

The severity of impact from plastics varied with feeding strategy. Omnivores and deposit feeders were most greatly affected, with filter feeders experiencing weaker, but nonetheless significant, negative impacts. Microplastic ingestion varies by feeding strategy (Bour et al. 2018, Naji et al. 2018), with 16% more microplastics ingested by predators and deposit feeders compared to filter feeders (Bour et al. 2018). The greater ingestion of MPP by

Our findings were in keeping with Berlino et al. (2021), which also found that benthic filter feeders were negatively impacted by microplastics although, in the earlier study, omnivores, predators and grazers were not. The strong effects on grazers in the present meta-analysis likely resulted from high microplastic concentration on the sediment surface or, in experimental conditions, on the tank floor. Microplastics naturally congregate on the seafloor, with the majority of benthic microplastics found in the top 0.5cm sediment (Martin et al. 2017), where grazers (and some omnivores) predominantly feed (Duchêne and Rosenberg 2001). Strong effects of microplastics on predators and omnivores could result from the trophic transfer of microplastics through the food chain, with microplastic fragments being most prone to bioaccumulation (Gray & Weinstein 2017). The majority of our 72 studies, however, were short-term laboratory experiments, in which study organisms were purchased from aquaria and exposed directly to microplastics, suggesting that trophic transfer would not have influenced our results and demonstrating a need for more environmentally realistic laboratory experiments.

4.3 MICROPLASTIC CHARACTERISTICS

While organismal characteristics were the primary causes for variation in microplastic impact, microplastic shape and polymer type significantly contributed to variation in effect size. We found no effect of microplastic size, exposure concentration and exposure duration, despite individual studies documenting stronger negative impacts at higher exposures (Green et al. 2016, Lo & Chan 2018). The recorded influence of polymer type conflicted with findings of Lei et al. (2018), where the size of microplastic particle determined toxicity in nematodes and zebrafish and the polymer composition was less important. However, polymer type of a microplastic influences the specific density and hydrophobicity of a particle and

thus the biofouling and sinking rates (Kaiser et al. 2017). It is therefore logical that polymer type will influence the availability of both the microplastic itself and its leachates to an organism. In terms of shape, fragments and fibres had larger effects than spheres and squares, potentially, in the case of fragments, due to sharp edges that cause damage following ingestion (Pirsaheb et al. 2020). Fragments and fibres are likely to become the most prevalent microplastics in marine ecosystems, already constituting 48.5% and 31%, respectively, of microplastics in sediment and water (Kooi & Koelmans 2019). The high prevalence of fragments and fibres in marine ecosystems makes the effects of these shapes, compared with spheres, for example, far more environmentally realistic, suggesting that the strong negative impacts of these particle shapes could have widespread implications for benthic ecosystems. Microplastic dosage had less influence over impacts than microplastic shape or polymer type. This may in part be due to the focus of meta-analytical techniques on average responses, since the influence of microplastic concentration may be more pronounced at extreme values. However, since extreme values are likely to be less environmentally realistic, we consider the use of average values was not detrimental to our conclusions. Of the polymer types investigated, microplastic leachates which had been separated from their microplastic substrates had the strongest negative impacts on fauna. The impacts of leachates on benthic fauna have not been previously investigated by meta-analyses. Leachates included contaminants such as persistent organic pollutants (POPs) which had adsorbed onto the microplastic surface and later been separated, as well as chemicals which has leached directly from the microplastic. Leachates had negative impacts on reproduction, development and feeding of echinoderms. Leachate endocrine disruptor chemicals (EDCs) can alter hormone production, causing issues such as reduced reproductive success (Gallo et al. 2018).

Microplastics with adsorbed benzo[a]pyrene and perfluorooctane sulfonic acid cause more

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damage to gill tissues and digestive glands compared to non-contaminated microplastics (O'Donovan et al. 2018). On a cellular level, changes to enzyme activity in gobies have been seen following exposure to the antibiotic celafexin (Fonte et al. 2016), while microplastic associated polychlorinated biphenyls (PCBs) have been shown to contribute to effects such as hepatic stress, tissue accumulation of chemicals, reduced feeding activity and increased mortality (Besseling et al. 2013, Rochman et al. 2013, Herzke et al. 2016). Adsorbed chemicals may therefore have contributed to the negative impacts on feeding activity found by the present study.

4.4 DISTRIBUTION OF LITERATURE USED

There was a skew in the number of studies by geographic location and sampling taxa. Most studies were published in Europe (49%) or Asia (24%), with Africa, North America and South America somewhat underrepresented, resulting in a lack of knowledge surrounding native and commercially important species in these regions. The majority of studies analysed were conducted on molluscs that had relevance to human food supply, usually commercially important bivalve species such as the blue mussel, *Mytilus edulis*. For a comprehensive overview to be representative of global impacts, funding should be directed towards addressing the knowledge gaps surrounding continents such as Africa and less commercially important organisms such as polychaetes, for which there is a lack of data. The numbers of relevant studies are increasing rapidly, indicating an opportunity for these knowledge gaps to be filled. Crucially, for findings to be truly comparable there is a need for standardisation of sampling methodology and units of expression, a point widely made in past papers (Hermsen et al. 2016, Miller at al. 2017, Ajith et al. 2020).

5.0 CONCLUSIONS

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- Microplastic exposure has significant negative impact on multiple biological processes of marine benthic fauna assessed.
- This study provides the first meta-analytical evidence that microplastic leachates have more severe impacts on benthic fauna than microplastic particles themselves. Clearly, microplastic management should consider the fate of microplastic already within the marine system, alongside minimising further input.
- Significant knowledge gaps remain surrounding certain geographic regions and species without commercial interest. Future research should be directed towards addressing these gaps.
- A rapid increase in microplastic studies since 2019 caused this study to reveal stronger and more consistently negative effects of microplastics than previous metaanalyses. There is an undeniable and urgent call to address the microplastic crisis within waste management systems globally.

510 6.0 FIGURES AND TABLES WITH CAPTIONS

Table 1. Heterogeneity of effect sizes of microplastics on marine benthic fauna, given as: Wald's Value (I^2) , Cochran's value (Q), and the degrees of freedom (DF) and p-value pertaining to Cochran's Q.

I ² (%)	Q	DF	p-value	
61.4	2668.9	700	< 0.001	
75.2	658.3	72	< 0.001	
74.4	368.5	102	< 0.001	
59.4	278.4	131	< 0.001	
34.2	179.7	109	< 0.001	
47.0	602.2	158	< 0.001	
80.3	149.4	38	< 0.001	
73.9	304.2	84	< 0.001	
	61.4 75.2 74.4 59.4 34.2 47.0 80.3	61.4 2668.9 75.2 658.3 74.4 368.5 59.4 278.4 34.2 179.7 47.0 602.2 80.3 149.4	61.4 2668.9 700 75.2 658.3 72 74.4 368.5 102 59.4 278.4 131 34.2 179.7 109 47.0 602.2 158 80.3 149.4 38	61.4 2668.9 700 <0.001

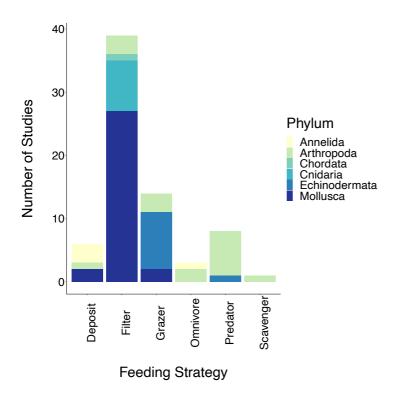


Figure 1. The frequency of animal feeding strategy by phylum used in 72 experimental studies.

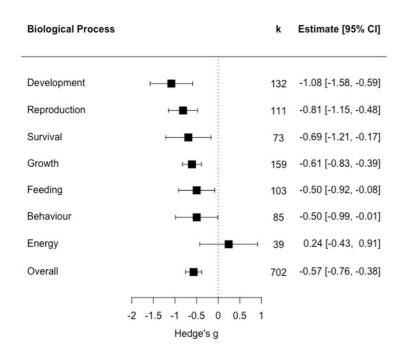


Figure 2. The effects of microplastic exposure on biological processes of marine benthic fauna. Effects on each of 7 processes and overall, as indicated from random-effects modelling. Boxes and error bars represent pooled Hedge's g values and 95% confidence

intervals, respectively. K represents the number of case studies, or independent experiments within the same study. Overlap of confidence intervals with 0 indicate non-significance.

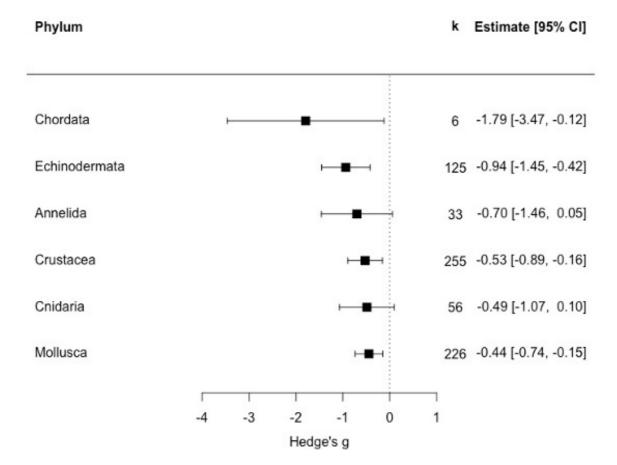


Figure 3. The effects of microplastic exposure on phyla of marine benthic fauna. Effects on each of 6 phyla as indicated from mixed effects modelling. Boxes and error bars represent pooled Hedge's g values and 95% confidence intervals, respectively. K represents the number of case studies.

Feeding Strategy k Estimate [95% CI]

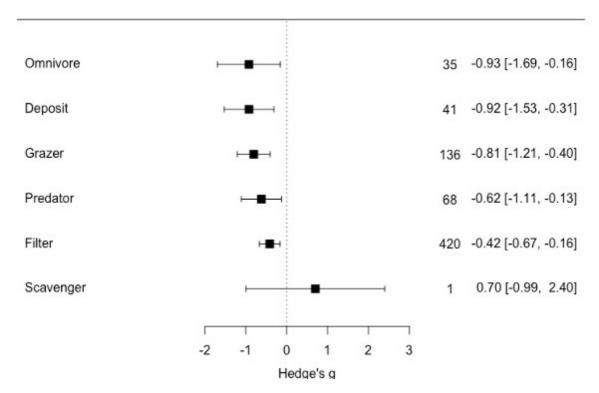


Figure 4. The effects of microplastic exposure on feeding strategies of marine benthic fauna. Effects on each of 6 feeding strategies, as indicated from mixed-effects modelling. Boxes and error bars represent pooled Hedge's g values and 95% confidence intervals, respectively. K represents the number of case studies.

Life Stage k Estimate [95% CI]

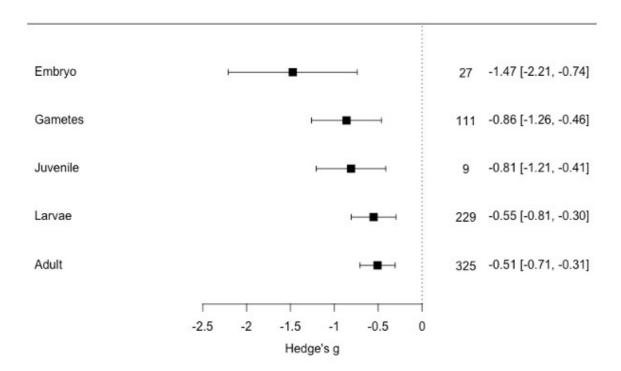


Figure 5. The effects of microplastic exposure on life stages of marine benthic fauna.

Effects on each of 5 life stages as indicated from mixed-effects modelling. Boxes and error bars represent pooled Hedge's g values and 95% confidence intervals, respectively. K represents the number of case studies.

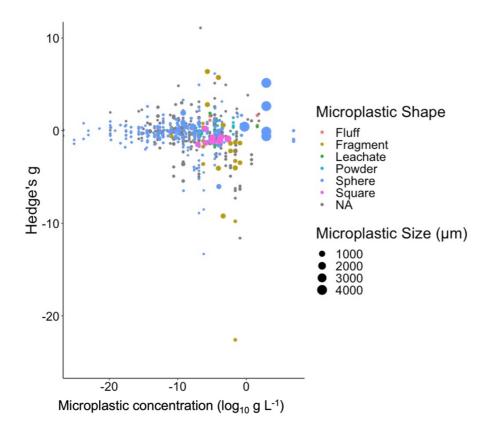


Figure 6. Effect of microplastic exposure concentration on the biological processes of marine benthos. Effect size indicated by Hedge's g value. Point size is indicative of microplastic particle size, while colour represents the shape of the particle.

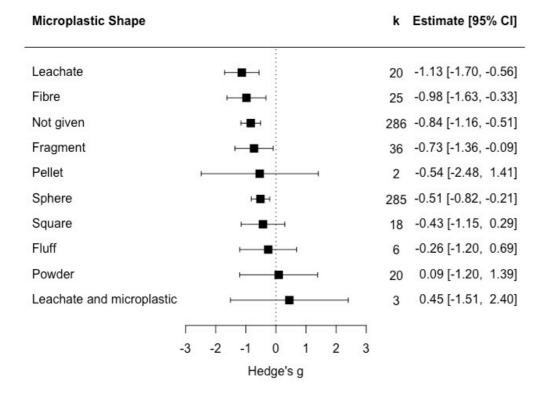


Figure 7. Responses of benthic fauna to the shape of microplastics used by experiments. Responses indicated from mixed effects modelling. Boxes and error bars represent pooled Hedge's g values and 95% confidence intervals, respectively. K represents the number of case studies. 'Leachate and microplastic' refers to microplastic particles with adsorped leachates, while 'leachate' refers to leachate which is not adsorped to a particle.

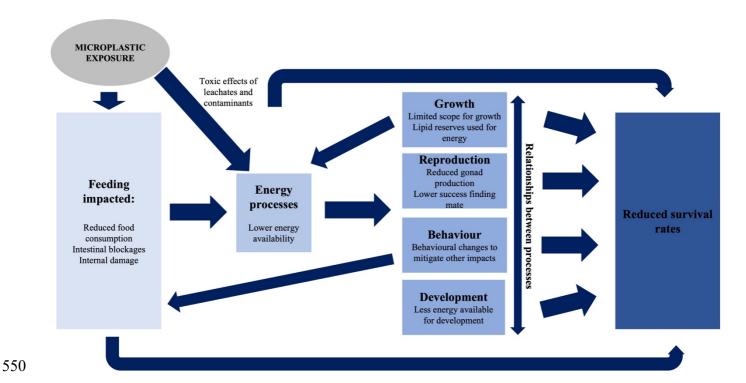


Figure 8. Interactions of impacts on different biological processes of marine benthic fauna, as a result of microplastic exposure. Interactions demonstrated by arrows, culminating in a synergistic effect and overall reduction in survival rate.

7.0 ACKNOWLEDGEMENTS

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Appendix: Supplementary Materials 1 2 3 Microplastics alter multiple biological processes of marine benthic fauna 4 5 Running page head: Microplastic impacts on benthic fauna. 6 7 Victoria G. Mason, Martin W. Skov, Jan Geert Hiddink, Mark Walton 8 9 School of Ocean Sciences, Bangor University, Isle of Anglesey. LL59 5AB. UK. 10 11 Email: torimason@hotmail.co.uk 12 **OVERVIEW OF CONTENT:** 13 14 A systematic review and meta-analysis were conducted to quantify the impacts of 15 microplastics on the biological processes (Table S1) of marine benthic fauna (Figure S1). The 16 influence of organism and microplastic characteristics were also investigated. Search terms for the systematic review were scoped using 9 test searches, where the relevance of hits was 17 18 evaluated based on the inclusion of 10 pre-determined key reference studies (Table S2). 19 Studies were then screened by title, abstract and full text to produce a final list of 72 20 publications (Table S3). Data were extracted from the final 72 studies (Table S4). Reference 21 numbers were recorded and included for each study to allow tracing through the stages and 22 identification of any replicate studies. Hedge's g value was calculated to quantify the effect 23 size in each study, using the data extracted (Table S4). The directionality of effect was 24 changed from positive to negative for study results where an increase in a response variable

represented a negative impact on the organism (Table S5). Number of studies published over

time and by region were plotted to visualise the distribution of the data temporally and spatially (Figure S2, S3). The potential effect of publication bias was assessed using the 'trim and fill' method (Duval and Tweedie 2000), with the results shown in Table S6. Adjusting the estimated pooled effect size in our study had little effect on the overall outcome and indicated that publication bias was likely to have had a negligible effect on our results.

Random effects modelling was then used to analyse the influence of drivers such as phylum, life stage and microplastic exposure characteristics. The most significant results were found from phylum, feeding strategy, microplastic duration, shape and polymer type, as outlined in the main text. Further, less significant results such as the influence of microplastic size and duration were included in these supplementary materials (Figure S4), as well as a sub-group relationships of effect size in each taxonomic group with microplastic exposure concentration (Figure S5). Effect sizes for exposure to different polymer types are shown in Figure S6.

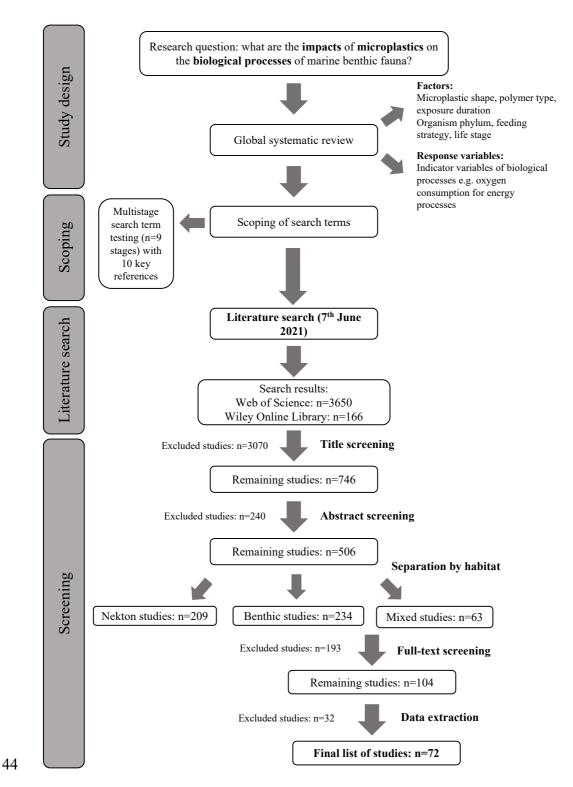
39 1.0 INTRODUCTORY TABLES

40 *Table S1.* Biological rates used in this study, with trait type, indicator variables and source.

41 Based on definitions by Violle et al. (2007).

Biological rate	Definition	Examples of indicator
		variables
Survival	Number of individuals surviving over	Mortality rate, survival rate,
	time with exposure to microplastic	number/% of live
	treatment	individuals
Growth	Physical increase in body size of an	Somatic growth rate, length
	organism (somatic growth)	increase, weight increase
Reproduction	Ability of an organism to	Reproductive success, %
	successfully produce viable young	live young, sperm velocity,
		oocyte number, fecundity
Development	The development of specific body	% normal development, %
	parts or progression of an organism	larval abnormalities,
	through life stages	development time, segment
		regeneration time
Behaviour	Characteristics of organism	Righting time, byssal thread
	behaviour relating to movement,	production, cirral beating
	boldness and activity	frequency, swimming speed
Feeding	Ability of an organism to	Prey consumption rate,
	successfully consume food sources or	algal clearance rate, %
	capture prey	ingestion success
Energy	Processes involving the generation of	Respiration rate (oxygen
consumption	energy in an organism, usually	consumption), energy
	respiration	consumption

43 2.0 SYSTEMATIC REVIEW METHODOLOGY



45 *Figure S1.* Flow chart depicting study design and methodology of the present study through 46 the scoping literature search and screening processes. One scoping stage refers to one test 47 search of the search string.

Table S2. Key references used when scoping potential search terms to assess for relevance of results. Studies given with author, publication date, study organism and the number of citations. Number of citations as given by Web of Science on 27th May 2021 (benthic studies) and 4th June 2021 (nekton studies). Studies selected for relevance, range of study organisms and number of citations.

	Authors	Year of	Study Organism	Number of
		Publication		Citations
Benthic	Murray and Cowie	2011	Nephrops norvegicus	448
	Farrell and Nelson	2013	Mytilus edulis	569
	Setälä et al.	2014	Macoma balthica	149
			Mytilus trossolus	
			Gammarus spp.	
			Mysid shrimps	
			Monoporeia affinis	
			Marenzelleria spp.	
	Van Cauwenberghe and	2014	Crassostrea gigas	653
	Janssen		Mytilus edulis	
	Van Cauwenberghe et al.	2015	Mytilus edulis	429
			Arenicola marina	
Nekton	Bourdages et al.	2020	Seals (range)	6
	Egbeocha et al.	2018	Range	20
	Hu et al.	2020	Oryzias latipes	7
	Le Bihanic et al.	2020	Oryzias melastigma	12
	Critchell and	2018	Acanthochromis	66
	Hoogenboom		polyacanthus	

Table S3. Final list of papers (n=72) from which data were extracted for meta-analysis,

- 55 following title, abstract and full text screening. Papers are given with reference number from
- 56 the original search results (7th June 2021), title, authors, publication year and DOI.

Ref No	Authors	Article Title	Year	DOI	Number of
					Observations
11	Berry, KLE; Epstein, HE; Lewis,	Microplastic Contamination Has	2019	10.3390/d111	30
	PJ; Hall, NM; Negri, AP	Limited Effects on Coral Fertilisation		20228	
		and Larvae			
21	Reichert, J; Arnold, AL;	Impacts of microplastics on growth and	2019	10.1016/j.env	4
	Hoogenboom, MO; Schubert, P;	health of hermatypic corals are species-		pol.2019.1130	
	Wilke, T	specific		74	
29	Horn, DA; Granek, EF; Steele, CL	Effects of environmentally relevant	2020	10.1002/lol2.	2
		concentrations of microplastic fibers on		10137	
		Pacific mole crab (Emerita analoga)			
		mortality and reproduction			
31	Seuront, 1	Microplastic leachates impair	2018	10.1098/rsbl.2	2
		behavioural vigilance and predator		018.0453	
		avoidance in a temperate intertidal			
		gastropod			
43	Tosetto, l; Brown, C; Williamson,	Microplastics on beaches: ingestion	2016	10.1007/s002	3
	JE	and behavioural consequences for		27-016-2973-	
		beachhoppers		0	
58	Crump, A; Mullens, C; Bethell, EJ;	Microplastics disrupt hermit crab shell	2020	10.1098/rsbl.2	1
	Cunningham, EM; Arnott, G	selection		020.0030	
69	Santana, MFM; Moreira, FT;	Continuous Exposure to Microplastics	2018	10.1007/s002	2
	Pereira, CDS; Abessa, DMS; Turra,	Does Not Cause Physiological Effects		44-018-0504-	
	A	in the Cultivated Mussel Perna perna		3	

93	Corinaldesi, C; Canensi, S;	Multiple impacts of microplastics can	2021	10.1038/s420	6
	Dell'Anno, A; Tangherlini, M; Di	threaten marine habitat-forming species	3	03-021-	
	Capua, I; Varrella, S; Willis, TJ;			01961-1	
	Cerrano, C; Danovaro, R				
108	Sussarellu, R; Suquet, M; Thomas,	Oyster reproduction is affected by	2016	10.1073/pnas.	1
	Y; Lambert, C; Fabioux, C; Pernet,	exposure to polystyrene microplastics		1519019113	
	MEJ; Le Goic, N; Quillien, V;				
	Mingant, C; Epelboin, Y;				
	Corporeau, C; Guyomarch, J;				
	Robbens, J; Paul-Pont, I; Soudant,				
	P; Huvet, A				
110	Torn, K	Microplastics uptake and accumulation	2020	10.3176/proc.	2
		in the digestive system of the mud crab		2020.1.04	
		Rhithropanopeus harrisii			
161	Yu, P; Liu, ZQ; Wu, DL; Chen,	Accumulation of polystyrene	2018	10.1016/j.aqu	4
	MH; Lv, WW; Zhao, YL	microplastics in juvenile Eriocheir		atox.2018.04.	
		sinensis and oxidative stress effects in		015	
		the liver			
239	Seuront, l; Nicastro, KR; McQuaid,	Microplastic leachates induce species-	2021	10.1002/eap.2	4
	CD; Zardi, GI	specific trait strengthening in intertidal		222	
		mussels			
252	Welden, NAC; Cowie, PR	Long-term microplastic retention	2016	10.1016/j.env	2
		causes reduced body condition in the		pol.2016.08.0	
		langoustine, Nephrops norvegicus		20	
254	Xu, XY; Lee, WT; Chan, AKY; Lo,	Microplastic ingestion reduces energy	2017	10.1016/j.mar	6
	HS; Shin, PKS; Cheung, SG	intake in the clam Atactodea striata		polbul.2016.1	
				2.027	

266	Kaposi, KL; Mos, B; Kelaher, BP;	Ingestion of Microplastic Has Limited	2014	10.1021/es40	8
	Dworjanyn, SA	Impact on a Marine Larva		4295e	
289	Green, DS; Boots, B; Sigwart, J;	Effects of conventional and	2016b	10.1016/j.env	18
	Jiang, S; Rocha, C	biodegradable microplastics on a		pol.2015.10.0	
		marine ecosystem engineer (Arenicola		10	
		marina) and sediment nutrient cycling			
291	Mouchi, V; Chapron, l; Peru, E;	Long-term aquaria study suggests	2019	10.1016/j.env	4
	Pruski, AM; Meistertzheim, AL;	species-specific responses of two cold-		pol.2019.07.0	
	Vetion, G; Galand, PE; Lartaud, F	water corals to macro-and		24	
		microplastics exposure			
303	Green, DS; Colgan, TJ; Thompson,	Exposure to microplastics reduces	2019	10.1016/j.env	2
	RC; Carolan, JC	attachment strength and alters the		pol.2018.12.0	
		haemolymph proteome of blue mussels		17	
		(Mytilus edulis)			
314	Opitz, T; Benitez, S; Fernandez, C;	Minimal impact at current	2021	10.1016/j.mar	6
	Osores, S; Navarro, JM; Rodriguez-	environmental concentrations of		polbul.2020.1	
	Romero, A; Lohrmann, KB;	microplastics on energy balance and		11834	
	Lardies, MA	physiological rates of the giant mussel			
		Choromytilus chorus			
360	Besseling, E; Wegner, A; Foekema,	Effects of Microplastic on Fitness and	2013	10.1021/es30	6
	EM; van den Heuvel-Greve, MJ;	PCB Bioaccumulation by the Lugworm	1	2763x	
	Koelmans, AA	Arenicola marina (l.)			
402	Gambardella, C; Morgana, S;	Ecotoxicological effects of polystyrene	2018	10.1016/j.mar	3
	Bramini, M; Rotini, A; Manfra, 1;	microbeads in a battery of marine		envres.2018.0	
	Migliore, 1; Piazza, V; Garaventa,	organisms belonging to different		9.023	
	F; Faimali, M	trophic levels			

532	Silva, PPGE; Nobre, CR; Resaffe,	Leachate from microplastics impairs	2016	10.1016/j.wat	3
	P; Pereira, CDS; Gusmao, F	larval development in brown mussels		res.2016.10.0	
				16	
562	Woods, MN; Hong, TJ; Baughman,	Accumulation and effects of	2020	10.1016/j.mar	3
	D; Andrews, G; Fields, DM;	microplastic fibers in American lobster		polbul.2020.1	
	Matrai, PA	larvae (Homarus americanus)		11280	
570	Leung, J; Chan, KYK	Microplastics reduced posterior	2018	10.1016/j.mar	5
		segment regeneration rate of the		polbul.2017.1	
		polychaete Perinereis aibuhitensis		0.072	
586	Webb, S; Gaw, S; Marsden, ID;	Biomarker responses in New Zealand	2020	10.1016/j.eco	4
	Mcrae, NK	green-lipped mussels Perna		env.2020.110	
		canaliculus exposed to microplastics		871	
		and triclosan			
588	Hankins, C; Moso, E; Lasseigne, D	Microplastics impair growth in two	2021	10.1016/j.env	2
		atlantic scleractinian coral species,		pol.2021.1166	
		Pseudodiploria clivosa and Acropora		49	
		cervicornis			
591	Trifuoggi, M; Pagano, G; Oral, R;	Microplastic-induced damage in early	2019	10.1016/j.env	15
	Pavicic-Hamer, D; Buric, P;	embryonal development of sea urchin		res.2019.1088	
	Kovacic, I; Siciliano, A; Toscanesi,	Sphaerechinus granularis		15	
	M; Thomas, PJ; Paduano, 1; Guida,				
	M; Lyons, DM				
621	Yap, VHS; Chase, Z; Wright, JT;	A comparison with natural particles	2020	10.1016/j.mar	18
	Hurd, CL; Lavers, JL; Lenz, M	reveals a small specific effect of PVC		polbul.2020.1	
		microplastics on mussel performance		11703	
662	Cole, M; Galloway, TS	Ingestion of Nanoplastics and	2015	10.1021/acs.e	10
		Microplastics by Pacific Oyster Larvae		st.5b04099	

676	Luan, LP; Wang, X; Zheng, H; Liu,	Differential toxicity of functionalized	2019	10.1016/j.mar	26
	LQ; Luo, XX; Li, FM	polystyrene microplastics to clams		polbul.2019.0	
		(Meretrix meretrix) at three key		1.003	
		development stages of life history			
717	Missawi, O; Bousserrhine, N;	Uptake, accumulation and associated	2021	10.1016/j.jhaz	4
	Zitouni, N; Maisano, M;	cellular alterations of environmental		mat.2020.124	
	Boughattas, I; De Marco, G;	samples of microplastics in the		287	
	Cappello, T; Belbekhouche, S;	seaworm Hediste diversicolor			
	Guerrouache, M; Alphonse, V;				
	Banni, M				
721	Rist, SE; Assidqi, K; Zamani, NP;	Suspended micro-sized PVC particles	2016	10.1016/j.mar	11
	Appel, D; Perschke, M; Huhn, M;	impair the performance and decrease		polbul.2016.0	
	Lenz, M	survival in the Asian green mussel		7.006	
		Perna viridis			
729	Nobre, CR; Santana, MFM; Maluf,	Assessment of microplastic toxicity to	2015	10.1016/j.mar	2
	A; Cortez, FS; Cesar, A; Pereira,	embryonic development of the sea		polbul.2014.1	
	CDS; Turra, A	urchin Lytechinus variegatus		2.050	
		(Echinodermata: Echinoidea)			
766	Leads, RR; Burnett, KG; Weinstein,	The Effect of Microplastic Ingestion on	2019	10.1002/etc.4	5
	JE	Survival of the Grass Shrimp		545	
		Palaemonetes pugio (Holthuis, 1949)			
		Challenged with Vibrio campbellii			
776	Green, DS	Effects of microplastics on European	2016a	10.1016/j.env	10
		flat oysters, Ostrea edulis and their		pol.2016.05.0	
		associated benthic communities		43	
787	Wang, X; Liu, LQQ; Zheng, H;	Polystyrene microplastics impaired the	2020	10.1016/j.mar	24
	Wang, MX; Fu, YX; Luo, XX; Li,	feeding and swimming behavior of		polbul.2019.1	
	FM; Wang, ZY	mysid shrimp Neomysis japonica		10660	

791	Rist, S; Baun, A; Almeda, R;	Ingestion and effects of micro- and	2019	10.1016/j.mar	12
	Hartmann, NB	nanoplastics in blue mussel (Mytilus		polbul.2019.0	
		edulis) larvae		1.069	
802	Tallec, K; Huvet, A; Di Poi, C;	Nanoplastics impaired oyster free	2018	10.1016/j.env	16
	Gonzalez-Fernandez, C; Lambert,	living stages, gametes and embryos		pol.2018.08.0	
	C; Petton, B; Le Goic, N; Berchel,			20	
	M; Soudant, P; Paul-Pont, I				
812	Carrasco, A; Pulgar, J; Quintanilla-	The influence of microplastics	2019	10.1016/j.mar	2
	Ahumada, D; Perez-Venegas, D;	pollution on the feeding behavior of a		polbul.2019.0	
	Quijon, PA; Duarte, C	prominent sandy beach amphipod,		5.018	
		Orchestoidea tuberculata (Nicolet,			
		1849)			
827	Syakti, AD; Jaya, JV; Rahman, A;	Bleaching and necrosis of staghorn	2019	10.1016/j.che	5
	Hidayati, NV; Raza'i, TS; Idris, F;	coral (Acropora formosa) in laboratory		mosphere.201	
	Trenggono, M; Doumenq, P; Chou,	assays: Immediate impact of LDPE		9.04.156	
	LM	microplastics			
881	Bertucci, JI; Bellas, J	Combined effect of microplastics and	2021	10.1016/j.scit	2
		global warming factors on early growth	l	otenv.2021.14	
		and development of the sea urchin		6888	
		(Paracentrotus lividus)			
930	Watts, AJR; Urbina, MA; Corr, S;	Ingestion of Plastic Microfibers by the	2015	10.1021/acs.e	3
	Lewis, C; Galloway, TS	Crab Carcinus maenas and Its Effect		st.5b04026	
		on Food Consumption and Energy			
		Balance			
971	Capolupo, M; Franzellitti, S;	Uptake and transcriptional effects of	2018	10.1016/j.env	6
	Valbonesi, P; Lanzas, CS; Fabbri, E	polystyrene microplastics in larval		pol.2018.06.0	
		stages of the Mediterranean mussel		35	
		Mytilus galloprovincialis			

1036	Urban-Malinga, B; Jakubowska, M;	Response of sediment-dwelling	2021	10.1016/j.scit	5
	Bialowas, M	bivalves to microplastics and its		otenv.2020.14	
		potential implications for benthic		4302	
		processes			
1124	Gardon, T; Reisser, C; Soyez, C;	Microplastics Affect Energy Balance	2018	10.1021/acs.e	6
	Quillien, V; Le Moullac, G	and Gametogenesis in the Pearl Oyster		st.8b00168	
		Pinctada margaritifera			
1132	Mohsen, M; Zhang, LB; Sun, LN;	Effect of chronic exposure to	2021	10.1016/j.eco	6
	Lin, CG; Wang, Q; Liu, SL; Sun,	microplastic fibre ingestion in the sea		env.2020.111	
	JC; Yang, HS	cucumber Apostichopus japonicus		794	
1209	Detree, C; Gallardo-Escarate, C	Single and repetitive microplastics	2018	10.1016/j.fsi.2	1
		exposures induce immune system		018.09.018	
		modulation and homeostasis alteration			
		in the edible mussel Mytilus			
		galloprovincialis			
1224	Thomas, PJ; Oral, R; Pagano, G;	Mild toxicity of polystyrene and	2020	10.1016/j.mar	58
	Tez, S; Toscanesi, M; Ranieri, P;	polymethylmethacrylate microplastics		envres.2020.1	
	Trifuoggi, M; Lyons, DM	in Paracentrotus lividus early life		05132	
		stages			
1247	Mendrik, FM; Henry, TB; Burdett,	Species-specific impact of	2021	10.1016/j.env	2
	H; Hackney, CR; Waller, C;	microplastics on coral physiology		pol.2020.1162	
	Parsons, DR; Hennige, SJ			38	
1321	Sikdokur, E; Belivermis, M; Sezer,	Effects of microplastics and mercury	2020	10.1016/j.env	2
	N; Pekmez, M; Bulan, OK; Kilic, O	on manila clam Ruditapes		pol.2020.1142	
		philippinarum: Feeding rate,		47	
		immunomodulation, histopathology			
		and oxidative stress			

1362	Green, DS; Boots, B; O'Connor,	Microplastics Affect the Ecological	2017	10.1021/acs.e	8
	NE; Thompson, R	Functioning of an Important Biogenic		st.6b04496	
		Habitat			
1393	Wang, SX; Zhong, Z; Li, ZQ;	Physiological effects of plastic	2021	10.1016/j.jhaz	4
	Wang, XH; Gu, HX; Huang, W;	particles on mussels are mediated by		mat.2020.124	
	Fang, JKH; Shi, HH; Hu, MH;	food presence		136	
	Wang, YJ				
1441	Anderson, G; Shenkar, N	Potential effects of biodegradable	2021	10.1016/j.env	6
		single-use items in the sea: Polylactic		pol.2020.1153	
		acid (PLA) and solitary ascidians		64	
1457	Gonzalez-Soto, N; Hatfield, J;	Impacts of dietary exposure to different	2019	10.1016/j.scit	8
	Katsumiti, A; Duroudier, N;	sized polystyrene microplastics alone		otenv.2019.05	
	Lacave, JM; Bilbao, E; Orbea, A;	and with sorbed benzo[a]pyrene on		.161	
	Navarro, E; Cajaraville, MP	biomarkers and whole organism			
		responses in mussels Mytilus			
		galloprovincialis			
1474	Hope, JA; Coco, G; Thrush, SF	Effects of Polyester Microfibers on	2020	10.1021/acs.e	3
		Microphytobenthos and Sediment-		st.0c00514	
		Dwelling Infauna			
1479	Martinez-Gomez, C; Leon, VM;	The adverse effects of virgin	2017	10.1016/j.mar	24
	Calles, S; Gomariz-Olcina, M;	microplastics on the fertilization and		envres.2017.0	
	Vethaak, AD	larval development of sea urchins		6.016	
1506	Gu, HX; Wei, SS; Hu, MH; Wei, H;	Microplastics aggravate the adverse	2020	10.1016/j.jhaz	7
	Wang, XH; Shang, YY; Li, LA;	effects of BDE-47 on physiological and		mat.2020.122	
	Shi, HH; Wang, YJ	defense performance in mussels		909	
1539	Suckling, CC	Responses to environmentally relevant	2021	10.1016/j.scit	4
		microplastics are species-specific with		otenv.2020.14	
				2341	
-					

		dietary habit as a potential sensitivity			
		indicator			
1590	Diana, Z; Sawickij, N; Rivera, NA;	Plastic pellets trigger feeding responses	2020	10.1016/j.aqu	3
	Hsu-Kim, H; Rittschof, D	in sea anemones		atox.2020.105	
				447	
60	Korez, S; Gutow, 1; Saborowski, R	Feeding and digestion of the marine	2019	10.1016/j.cbp	1
		isopod Idotea emarginata challenged		c.2019.10858	
		by poor food quality and microplastics		6	
79	Yu, SP; Chan, BKK	Effects of polystyrene microplastics on	2020b	10.1016/j.eco	47
		larval development, settlement, and		env.2020.110	
		metamorphosis of the intertidal		362	
		barnacle Amphibalanus amphitrite			
83	Bruck, S; Ford, AT	Chronic ingestion of polystyrene	2018	10.1016/j.env	6
		microparticles in low doses has no		pol.2017.10.0	
		effect on food consumption and growth		15	
		to the intertidal amphipod			
		Echinogammarus marinus?			
181	Lo, HKA; Chan, KYK	Negative effects of microplastic	2018	10.1016/j.env	8
		exposure on growth and development		pol.2017.10.0	
		of Crepidula onyx		95	
342	Yu, SP; Chan, BKK	Intergenerational microplastics impact	2020c	10.1016/j.env	96
		the intertidal barnacle Amphibalanus		pol.2020.1155	
		amphitrite during the planktonic larval		60	
		and benthic adult stages			
392	Van Colen, C; Vanhove, B; Diem,	Does microplastic ingestion by	2020	10.1016/j.env	9
	A; Moens, T	zooplankton affect predator-prey		pol.2019.1134	
		interactions? An experimental study on		79	

larviphagy

1010	Bringer, A; Thomas, H; Prunier, G;	High density polyethylene (HDPE)	2020	10.1016/j.env	15
	Dubillot, E; Bossut, N; Churlaud,	microplastics impair development and		pol.2020.1139	
	C; Clerandeau, C; Le Bihanic, F;	swimming activity of Pacific oyster D-		78	
	Cachot, J	larvae, Crassostrea gigas, depending			
		on particle size			
1028	Bringer, A; Cachot, J; Prunier, G;	Experimental ingestion of fluorescent	2020	10.1016/j.che	9
	Dubillot, E; Clerandeau, C;	microplastics by pacific oysters,		mosphere.202	
	Thomas, H	Crassostrea gigas, and their effects on		0.126793	
		the behaviour and development at early			
		stages			
1153	Beiras, R; Bellas, J; Cachot, J;	Ingestion and contact with	2018	10.1016/j.jhaz	3
	Cormier, B; Cousin, X; Engwall,	polyethylene microplastics does not		mat.2018.07.1	
	M; Gambardella, C; Garaventa, F;	cause acute toxicity on marine		01	
	Keiter, S; Le Bihanic, F; Lopez-	zooplankton			
	Ibanez, S; Piazza, V; Rial, D; Tato,				
	T; Vidal-Linan, l				
1794	Beiras, R; Tato, T	Microplastics do not increase toxicity	2019	10.1016/j.mar	2
		of a hydrophobic organic chemical to		polbul.2018.1	
		marine plankton		1.029	
13	Yu, J; Tian, JY; Xu, R; Zhang, ZY;	Effects of microplastics exposure on	2020a	10.1016/j.env	28
	Yang, GP; Wang, XD; Lai, JG;	ingestion, fecundity, development, and		pol.2020.1154	
	Chen, R	dimethylsulfide production in		29	
		Tigriopus japonicus (Harpacticoida,			
		copepod)			
498	Lee, DH; Lee, S; Rhee, JS	Consistent exposure to microplastics	2021	10.1016/j.mar	20
		induces age-specific physiological and		polbul.2020.1	
		biochemical changes in a marine mysid		11850	

508	Li, ZC; Zhou, H; Liu, Y; Zhan, JJ;	Acute and chronic combined effect of	2020	10.1016/j.che	6
	Li, WT; Yang, KM; Yi, XL	polystyrene microplastics and dibutyl		mosphere.202	
		phthalate on the marine copepod		0.127711	
		Tigriopus japonicus			

Table S4. Data extracted from the final list of papers (n=72) for meta-analysis of the impacts

- of microplastics on the functional traits of marine benthic fauna, categorised by study
- 62 identifiers, meta-data and data for quantitative synthesis.

Study Identifier	Meta-data	Data for quantitative synthesis
Reference number	Location (continent, country,	Biological rate indicator (e.g.
Case study (a, b, c etc.)	region)	growth rate, respiration rate):
Author	Date of experiment	Control group (mean, standard
Publication Type	Study organism (phylum,	deviation, number of replicates,
Publication Year	species, life stage, feeding	units)
	strategy)	Experimental group (mean,
	Exposure conditions (duration,	standard deviation, number of
	microplastic concentration,	replicates, units)
	polymer type, microplastic	
	shape, microplastic size, added	
	contaminants)	

Table S5. Measured response variables of biological rates for which the units measured were converted from a positive to a negative value in this study (prior to calculation of effect size) in order to signify a negative impact on fauna. For example, mortality was measured as a positive value, but converted into a negative value as it was deemed negative for the animal.

Biological	Study	Year	Measured response and units
Survival	Wang et al.	2020	Mortality (%)
	Lo and Chan	2018	Mortality (individuals day ⁻¹)

Development	Berry et al.	2019	Embryo abnormality (%)
	Gandara e Silva et al.	2016	Abnormal larvae (%)
	Rist et al.	2019	Malformations (individuals/10)
	Thomas et al.	2020	Developmental defects (%)
	Martínez-Gómez et al.	2017	Abnormality (%)
	Bringer et al.	2020	Larval abnormalities (%)
	Yu et al.	2020	Development time (days)
Behaviour	Seuront	2018	Righting time (minutes)
	Crump et al.	2020	Time to enter shell (seconds)
	Gambardella et al.	2018	Swimming speed change (%)
	Hope et al.	2020	Burial time (hours)
	Suckling	2021	Righting time (seconds)
Growth	Wang et al.	2020	Growth inhibition (%)

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Table S6. Results of testing for publication bias using the 'trim and fill' method on rma.uni
 model. Result indicates assessment of balance of positive and negative effect size studies.
 Estimated effect size (in bold) indicates overall pooled Hedge's g effect size of microplastics
 on biological processes of benthic fauna, with Hedge's g adjusted for potential publication
 bias (trim and fill) and with our data (random effects model).

Test	Result	p-value	Estimated	Effect type	Model
			effect size		Reference
			(Hedge's g)		
Trim and fill	17 positive	< 0.0001	-0.61	Moderate	Duval and
with random	effect studies			negative	Tweedie (2000)
effects model	filled in (SE =				
	6.00)				
Rma.mv model		< 0.0001	-0.57	Moderate	Viechtbauer
				negative	(2010)

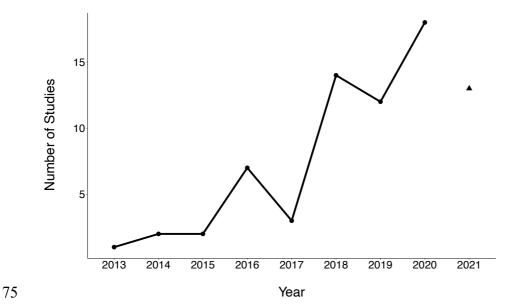


Figure S2. Number of studies related to the impact of microplastics on the biological rates of marine benthic fauna per publication year, from 2013-20. The triangle represents studies published in 2021 up until date of final search (7th June 2021).

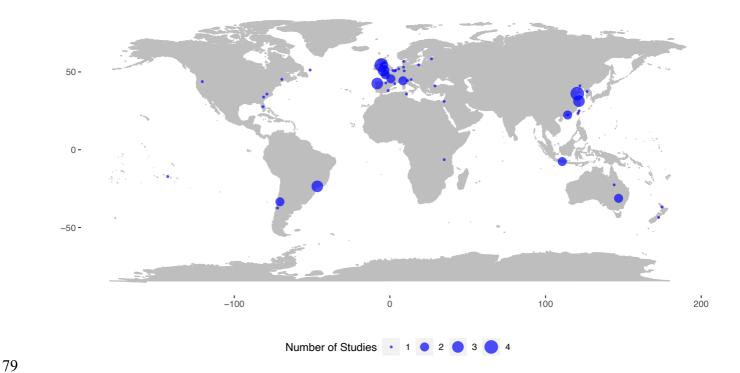


Figure S3. World map showing the number of publications related to the impact of microplastics on the functional traits of marine benthic fauna in each region. Circle size is proportional to the number of studies. Studies represented were published from 2013-21.

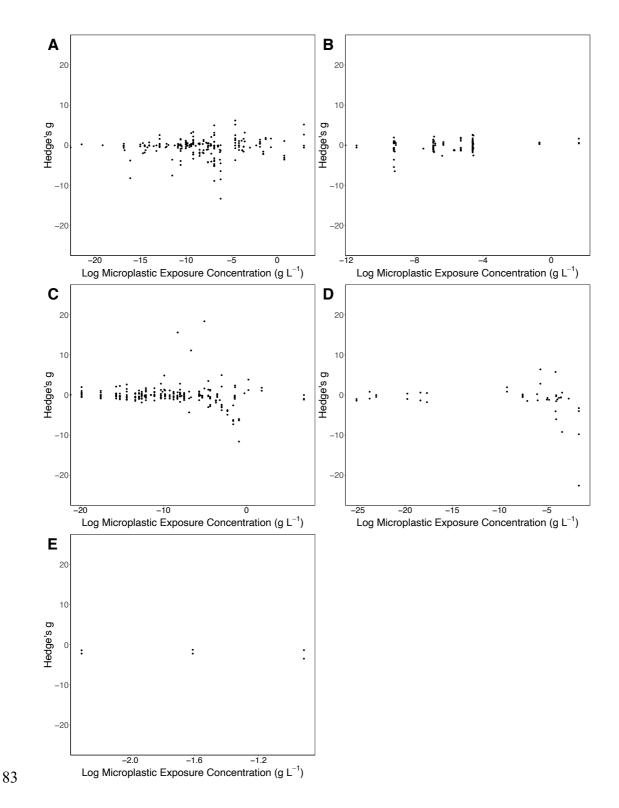
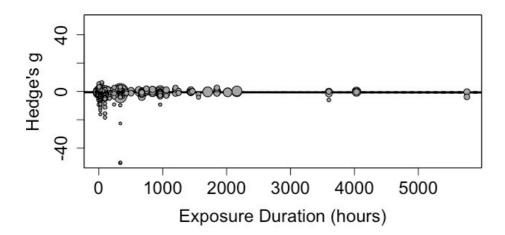


Figure S4. Relationship between log transformed microplastic exposure concentration (g L^{-1}) and effect size on marine benthic fauna (Hedge's g) for a) molluscs, b) echinoderms, c) crustaceans, d) cnidarians and e) chordates using studies from 2013-2021 which reported standardisable exposure concentration units (n = 54).



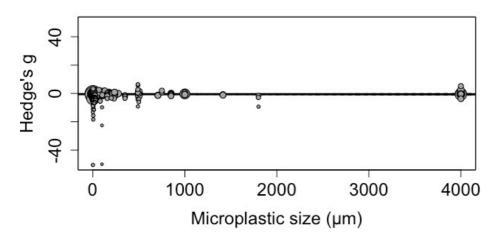


Figure S5. Meta-regression of a) exposure duration and b) microplastic size with Hedge's g effect size. The size of each point is proportional to the weight of the study (studies with larger sample sizes given greater weight), with smaller points given less weight. Regressions were produced based on the results of mixed-effects modelling using a) exposure duration and b) microplastic size as moderators.

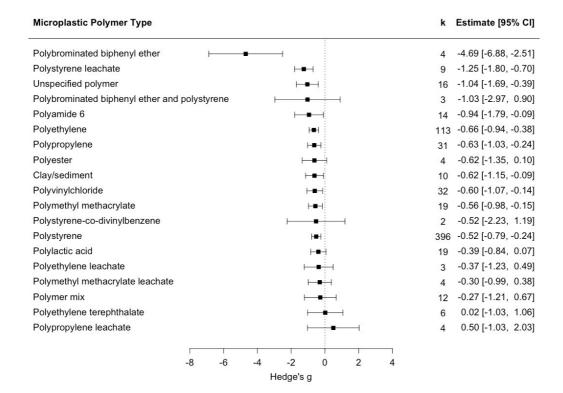


Figure S6. Influence of microplastic polymer type on marine benthic fauna. Influence indicated from mixed effects modelling, clay/sediment represents control. Boxes and error bars represent pooled Hedge's g values and 95% confidence intervals, respectively. K represents the number of case studies.

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- Meta-analysis revealed microplastics weaken multiple processes fundamental to seabed life
- Surge in research helps establish that plastic impacts are stronger than thought
- Severity of impact depends on feeding strategy, life stage and taxonomic group
- Early life stages are most strongly impacted by microplastic exposure
- Leaking chemicals generate stronger responses than plastic particles themselves