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**On their way to the North: larval performance of *Hemigrapsus sanguineus*
invasive to the European coast. A comparison with the native European
population of *Carcinus maenas***

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ABSTRACT

The Asian shore crab *Hemigrapsus sanguineus* has become invasive in North Europe and it co-occurs and competes with the native European shore crab *Carcinus maenas*. Both species develop through a feeding and dispersive larval phase characterised by several zoeal and a settling megalopa stage. Larvae of marine crabs are vulnerable to food limitation and warming has the potential to exacerbate the negative effects of food limitation on survival and growth. We quantified the combined effects of temperature and food limitation on larval performance (survival and growth) of *H. sanguineus* and we compared our results with those reported on performance of *C. maenas* larvae, under the same experimental design and methodology. Larvae from four females of *H. sanguineus* collected on Helgoland (North Sea) were experimentally reared from hatching to megalopa, at four temperatures (range 15-24°C) and two food conditions (permanent vs daily limited access to food). Larval survival of *H. sanguineus* was low at 15 °C and increased with temperature, in contrast to the high survival reported for *C. maenas* larvae in the range 15-24 °C. Food limitation reduced survival and body mass of *H. sanguineus* larvae at all temperatures, but without evidence of the exacerbating effect caused by high temperatures and reported for *C. maenas*. By contrast, high temperature (24°C) mitigated the negative effect of food limitation on body mass on *H. sanguineus* larvae. Advantages of *H. sanguineus* over *C. maenas* appear especially under the increased temperatures expected from climate change.

Keywords: food limitation, warming, invasive vs native crabs, survival, growth rates

58 INTRODUCTION

59 Increasing trade and travel in the last decades has led to biological invasions, i.e. the dispersion
60 and introduction of many species to geographical areas located outside their native range (Hulme 2011;
61 Bailey et al. 2020). Dispersion of organisms due to anthropogenic activities occurs through shipping
62 (ballast water, fouling), construction of corridors, aquaculture, fisheries, and food trade (Carlton 2002,
63 Hulme et al. 2008; Molnar et al. 2008; Katsanevakis et al. 2013). The introduction of a new species can
64 lead to the decline in native species richness and abundances, loss of genetic variation in the native
65 community, and an increase in the homogeneity of the invaded communities (Rahel 2000; Pyšek et al.
66 2012; Geburzi et al. 2018). Whenever the “introduced” species causes an impact on the host ecosystem,
67 it is considered “invasive”. Biological invasions are one of the most important threats to biodiversity
68 after changes in land and sea use, animal exploitation, climate change, and pollution (Brondizio et al.
69 2019; IPCC 2019). Invasive species affect the dynamics of native communities at different scales
70 (Hulme 2017), by means of different interactions including competition, predation, and introduction of
71 new diseases (Ruiz et al. 2000; Sakai et al. 2001; Jeschke et al. 2012). Biological invasions have been
72 reported for most marine and estuarine habitats in the world (Katsanevakis et al. 2013; Chan and Briski
73 2017; Pyšek et al. 2020). Many hypotheses have been proposed to explain biological invasions and the
74 success of the invaders (Simberloff and Von Holle 1999; Gurevitch et al. 2011; Kelley 2014), yet there
75 is still little information on the processes involved in dispersion and establishment of alien species in
76 novel habitats (Bailey et al. 2020; Rato et al. 2021).

77 Global climate change is causing an increase in the number and the impact of biological
78 invasions (Hulme 2017; González-Ortegón et al. 2020; Pyšek et al. 2020), because of the steady increase
79 in temperature (IPCC 2021), and the increased frequency of extreme events (e.g. heatwaves, Meehl and
80 Tebaldi 2004; Smale et al. 2019). These changes combined with other human activities can act together
81 to help the dispersal and establishment of new species. For example, the oyster *Crassostrea gigas* was
82 first introduced in the North Sea for aquaculture purposes and was able to establish there due to above-
83 average summer temperatures that helped the species to spread (Diederich et al. 2004; Smaal et al.
84 2009). Global warming has also led to the poleward expansion of many species (Sorte et al. 2010;
85 Poloczanska et al. 2013; Giménez et al. 2020).

86 Global warming could affect both invasive species and their native competitors in different
87 ways. How different species respond to changes in environmental variables (e.g. temperature) will
88 ultimately determine individual performance and species interactions. Increases in temperature cause
89 rises in metabolic demands, which should be met by food supply (Gillooly et al. 2001; Somero 2002).
90 However, for organisms living in the marine pelagic realm, where the food distribution is patchy
91 (Paffenhöfer et al. 1987, McManus et al. 2003) increased temperature combined with food limitation
92 could impair growth and survival. The combined effect of food limitation and increased temperature

93 may particularly impact life history stages characterised by high growth rates (Foster and Hirst 2012;
94 Otto et al. 2020). Crustacean larvae are an example where high growth rates are sustained by high
95 feeding rates, which in turn demand higher food availability (Anger 2001). In such case, one would
96 expect that increased temperatures combined with limited access to food may cause reductions in
97 growth rate and survival (Torres and Giménez 2020). However, the nature of the effect could vary
98 between native and exotic competitors because of differences in thermal optimum ranges (Griffith et
99 al. 2021). Hence, a critical question to understand current invasion and future population spread is how
100 increased temperature combined with food limitation drive the performance of both native and exotic
101 species.

102 The Asian shore crab *Hemigrapsus sanguineus* is native to the east coast of Asia (20° - 50° N)
103 (Takahashi et al. 1985; Fukui et al. 1989; Hwang et al. 1993). It is one of the most abundant crab species
104 on rocky beaches and occupies the upper and middle intertidal zones (Kikuchi et al. 1981; Fukui 1988).
105 It has successfully invaded the coasts of North America, North Europe, and the Adriatic and Black Seas
106 (Schubart 2003; Micu et al. 2010) via ballast water (Ai-yun and Yang 1991; Kraemer et al. 2007;
107 Epifanio 2013). In northern Europe, *H. sanguineus* was first recorded in France and the Dutch Delta
108 system in 1999 (Breton et al. 2002) and then spread to the North Sea and Scandinavia (Wolff 2005;
109 Kerckhof et al. 2007; Dauvin and Dufossé 2011; Gittenberg et al. 2010; Gothland et al. 2013; Seeley et
110 al. 2015; Karlsson et al. 2019). *H. sanguineus* was occasionally found in the Mediterranean and Black
111 Sea (Schubart 2003; Ben Souissi et al. 2004; Micu et al 2010; Ounifi-Ben Amor et al. 2016a,b;
112 GBIF.org). There are several factors likely to drive or limit the expansion of *H. sanguineus*, including
113 the presence of congeneric competitors (e.g. US North Pacific, Steinberg and Epifanio 2011, Lord 2017)
114 and low summer temperatures (Stephenson et al. 2009). In the Atlantic coast of North America, the
115 northern distribution limit of *H. sanguineus* is determined by the larval thermal tolerance to low
116 temperatures (Epifanio et al. 1998; Stephenson et al. 2009). Hence, as a result of warming, the
117 distribution of *H. sanguineus* is likely to expand further north (Epifanio 2013; Giménez et al. 2020). *H.*
118 *sanguineus* co-occurs and competes with the shore crab *Carcinus maenas* in Europe and in North
119 America. Both species overlap in diet and habitat use, with juvenile/adults of *H. sanguineus*
120 outcompeting *C. maenas* in the use of space and resources. *H. sanguineus* also predated on *C. maenas*
121 juveniles affecting their recruitment success (Lohrer and Whitlatch 2002; Jensen et al. 2002; Geburzi et
122 al. 2018). For example, in Southern New England *H. sanguineus* has significantly reduced the
123 recruitment of *C. maenas* due to direct predation, leading to a decline in densities by 40 - 90 % (Lohrer
124 and Whitlatch 2002). In addition, when in sympatry individuals of *C. maenas* migrate towards the
125 subtidal zone (Geburzi et al. 2018). The above-mentioned factors, help explain the displacement of *C.*
126 *maenas* from environments where it was previously more abundant.

127 We compared the performance of *H. sanguineus* and *C. maenas* from the perspective of the
128 larval phase for co-occurring populations of the German Bight (Helgoland, North Sea). We focus on

129 larvae because larval survival and recruitment are critical contributors to the propagule pressure by *H.*
130 *sanguineus* (Simberloff 2009) and for the persistence of populations of *C. maenas*. Propagule pressure,
131 i.e. a group of individuals of a species arriving in a region to which they are not native, drives the
132 establishment and spreading of invasive populations (Simberloff 2009). Marine larvae in particular,
133 tend to be more sensitive to environmental fluctuations than juveniles or adults (Pandori and Sorte 2019)
134 and larval survival is central to the recovery of populations after environmental disturbances (Cowen
135 and Sponaugle 2009; Pineda et al. 2009; Giménez et al. 2020). In particular, differences in larval
136 survival among co-occurring species may affect the balance of competition, either exacerbating or
137 counteracting the outcome. In theory, counteracting effects may occur in cases of trade-offs between
138 competition and dispersal abilities (Seifan et al. 2013). Because *H. sanguineus* was first reported in the
139 German Bight very recently (2008- Scrosati et al. 2011, 2009- Jungblut et al. 2017), it is not clear yet
140 whether *H. sanguineus* would be able to outcompete *C. maenas* at that local habitat. Unlike *C. maenas*,
141 larvae of *H. sanguineus* cannot develop at temperatures below 13-15 °C (Epifanio 2013) which
142 characterise the spring and early summers in the German Bight and coastal North Sea (Giménez et al.
143 2020). For *H. sanguineus*, larvae appear to be released when the temperature surpasses 15 °C which is
144 early/ mid- June depending on the year (Giménez et al. 2020). In the case of *C. maenas*, larval release
145 starts in May (Harms et al. 1994), but the full larval season of both species partially overlap. We know
146 that under food limitation, zoea I of *H. sanguineus* is more tolerant to short thermal fluctuations than
147 *C. maenas* (Giménez et al. 2021). However, over the entire larval phase and under low temperatures,
148 food limitation may produce a stronger negative effect on survival of *H. sanguineus* than that observed
149 in *C. maenas*.

150 Here we compared the responses of larvae of *Hemigrapsus sanguineus* and *Carcinus maenas*
151 to food limitation under increased temperature, reared in comparable experimental conditions. We first
152 quantified the combined effects of temperature and food limitation on survival and performance of
153 larvae of *H. sanguineus*. For the first time, we documented growth and survival responses in any
154 population of this species to food limitation over a wide range of temperatures. In particular, we
155 quantified the correlated responses of growth and development, driving size, and reserves at
156 metamorphosis, which for benthic invertebrates, are known to drive the performance of the post-
157 metamorphic stages in the benthic habitat (Giménez et al. 2004; Pechenik 2006; Giménez 2010; Torres
158 et al. 2016). Second, we compared our results on *H. sanguineus* with those obtained by Torres and
159 Giménez (2020) for a co-occurring population of *C. maenas*.

160

161

162

163 MATERIALS AND METHODS

164 Animal husbandry, larval rearing and experimental design

165 Berried females of *H. sanguineus* (carapace width 15.8 - 17.2 mm) were collected on the island
166 of Helgoland (North Sea, German Bight, 54°10'40.9" N 7°53'32.4" E) during their reproductive season
167 (July - September) and transported to the laboratory. Females were kept individually in 2-L aquaria
168 with UV-treated filtered (0.2 µm) seawater (32.5 ‰) permanently aerated, in a temperature-controlled
169 room at 18 °C (± 0.5 °C) with a 12:12 h light: dark cycle. Females were fed every 3 days with shrimps
170 (*Crangon crangon*) and water was changed daily to ensure high water quality at hatching.

171 The experimental setup comprised a factorial design, in which we exposed larvae obtained from
172 each given female to different combinations of temperatures and access to food (following Torres and
173 Giménez 2020). This experimental procedure was repeated four times, i.e. once for each of the hatches
174 obtained from four different females, in order to assess potential variations in responses driven by
175 maternal influence. Freshly hatched zoeae were distributed in 8 treatments (4 replicate vessels per
176 treatment, see below for more details), combination of 2 levels of daily access to food (6 or 24 h / day,
177 provided *ad libitum*) and 4 temperatures (15, 18, 21 and 24 °C). The different temperatures were chosen
178 based on its natural variability in the German Bight: 15 and 18 °C represent temperatures recorded
179 during the larval season of *H. sanguineus* (Giménez et al. 2020); this corresponds to summer
180 temperatures around the local population (Wiltshire and Manly, 2004). Temperatures > 20 °C are
181 expected as the consequence of steady warming due to climate change (Schrum et al. 2016), and as the
182 consequence of the expected increment in the frequency of warm summers (Christidis et al. 2015).

183 Experiments were carried out in temperature-controlled rooms and using natural UV-treated
184 filtered (0.2 µm) seawater. When hatching occurred, 50 larvae were sorted into each of 500-mL rearing
185 vessels (4 replicate vessels per treatment for each of the four females) in UV-treated filtered seawater
186 at the temperature of hatching. Freshly hatched *Artemia* sp. nauplii (Great Salt Lake Artemia, Sanders,
187 USA) were provided as food for the larvae in densities of ~5 nauplii/mL (Torres et al. 2021). In the
188 treatment of limited access to food, *Artemia* sp. nauplii were available for 6 hours each day (between 9
189 a.m. and 3 p.m., following Giménez and Anger 2005, Torres and Giménez 2020). By contrast, in the
190 treatment of permanent access to food, *Artemia* sp. nauplii were available all day. Water in all treatments
191 was changed daily following standard procedures for larval rearing (Torres et al. 2021). During the
192 daily water change, live larvae were staged and recorded, dead ones were also recorded and removed
193 from the experiments; in addition, we checked that remaining food was present in each rearing vessel.

194 Body mass and elemental composition (carbon and nitrogen) were measured in freshly hatched
195 larvae (3 replicates, 50 zoea each at the start of each experiment) and in freshly moulted megalopae
196 (sampled within 24 h after moulting). Carbon content is used as a proxy for lipid reserves, used by
197 crustacean larvae to sustain periods of food limitation (Dawirs et al. 1986; Anger and Harms 1990);
198 nitrogen content is used as a proxy for protein content (Dawirs 1996; Dawirs et al. 1986). The number

199 of individual megalopae sampled in each of the 4-replicate rearing vessel was on average 5 (Table S1).
200 Larvae were pipetted onto a filter, rinsed with distilled water, gently blotted dry with filter paper, and
201 stored in pre-weighed tin cartridges at -20 °C for later analysis. To determine the dry weight, samples
202 were freeze-dried for 48 h and weighed using a microbalance (Sartorius SC2, precision 1 µg). Carbon
203 and nitrogen content were then determined using an elemental analyser (vario MICRO cube CHNS
204 analyser, Elementar Analysensysteme).

205

206 **Data analysis**

207 The response variables were survival, duration of development, body mass, elemental
208 composition (carbon and nitrogen), and instantaneous growth at the megalopa. Survival to each zoeal
209 stage was calculated as the percentage of survivors in relation to the number of organisms at the start
210 of each experiment. Duration of development was calculated as the time elapsed from hatching to reach
211 each developmental stage. Growth rates were estimated as $G = \log(W_f/W_0)/t$. In this formula W_0 is the
212 average mass (dry weight, carbon or nitrogen) at hatching, W_f is the corresponding mass of each
213 individual megalopa collected in each rearing replicate and t is the corresponding duration of
214 development of each individual megalopa.

215 Mixed modelling was carried out in R (function lme and gls from package nlme, Pinheiro et al.
216 2018, R Core Team 2013) to assess the responses to the different combinations of food availability and
217 temperatures on survival and duration of development. The models contained temperature and food
218 availability as fixed factors and female of origin as a random factor. We performed backwards model
219 selection (Zuur et al. 2009) in two steps. In a first step, we tested the random terms using restricted
220 maximum likelihood (REML), we compared the different models through the corrected Akaike
221 information criteria (AICc) and ranked them. The model with the lowest AICc was selected for further
222 analysis. When the difference between two models was $\Delta AICc < 3$ and the most complex model had
223 the lower AICc we applied hypothesis testing (likelihood ratio tests). When the models differed
224 significantly ($p < 0.05$), we chose the model with the lowest AICc and when the difference was not
225 significant we chose the simpler model (with the lowest number of parameters). In a second step, the
226 fixed terms were analysed through maximum likelihood (ML). For dry mass, elemental composition,
227 and growth rates at the megalopa we did not get sufficient data in the food limited treatment at 15 °C
228 for female 4 (F4). We therefore, analysed the data using two different starting models: (1) considering
229 all females but without 15 °C and (2) considering all treatment combinations but without F4.

230 Data for survival was analysed in the logistic (Warton and Hui 2011) and logarithmic scale; as
231 a first step, proportions (p) were re-scaled using the formula $p' = [p(50-1) + 0.5]/50$, to avoid
232 inconsistencies associated to $\log(0)$ values. Logarithmically-transformed proportions were used to test
233 the multiplicative model, whereby temperature and food limitation would act independently on the

234 survival rates (Piggot et al. 2015). Survival responses consistent with a multiplicative model cannot be
 235 tested when the proportions are expressed in the logistic scale (Torres and Giménez 2020). Duration of
 236 development was analysed in the raw and the logarithmical scale to test if the effects were additive,
 237 multiplicative or interactive. Body mass and elemental composition were analysed in the raw scale.
 238 Tukey's honestly significant difference tests (Tukey's HSD) were performed to test differences among
 239 the different treatments.

240

241 **Comparison performance *H. sanguineus* and *C. maenas***

242 We compared the performance of *H. sanguineus* and *C. maenas* at different temperatures and
 243 food conditions through two means. First, we compared the integrative response of body mass and
 244 developmental time. We calculated the ratios between the body mass under food limitation and those
 245 observed under permanent access to food for each temperature; the same calculation was made for
 246 duration of development. For comparison, these standardised values were plotted against those of
 247 *Carcinus maenas*. In this plot, the unit corresponds to the values of body mass (and duration of
 248 development) under permanent access to food at each temperature.

249 We also calculated the ratios of survival (S_R) and growth rates (G_R) between species, i.e. as S_R
 250 $=S_H/S_C$ and $G_R = G_H/G_C$, where S_H and S_C are the survival to the megalopa of *H. sanguineus* and *C.*
 251 *maenas*, respectively and G_H and G_C the respective growth rates (from hatching to megalopa). We
 252 calculated average ratios and used simulations (details in: Supplementary material, Materials and
 253 Methods, Section 2. Data analysis: details on model simulation) to incorporate the intraspecific
 254 variation in survival and growth associated to variations within and among families. First, for each
 255 species, we simulated 1000 values of the survival and growth rates. Survival was simulated from the
 256 statistical model fitted to the responses of both species to the different combinations of temperature and
 257 access to food; i.e. there were 1000 values for each combination of species, response variable,
 258 temperature, and food condition. Growth was simulated from an additional statistical model which also
 259 considers survival as covariate; this model incorporates correlations between average survival and
 260 growth associated to female-to-female variation in larval performance. The data and models used for
 261 *C. maenas* correspond to Torres and Giménez (2020). The model used for survival of *H. sanguineus*
 262 was that of Tables S2; for growth we fitted an additional model based on female-to-female averages
 263 (Supplementary material, Materials and Methods, Section 2. Data analysis: details on model
 264 simulation). Simulations were performed using the function *simulate* of the package nlme (R Core Team
 265 2013) applied to the best fitted models for each variable. Second, for each combination of temperature
 266 and food condition a pair of values of survival and growth was randomly sampled (function *sample*, R
 267 Core Team 2013) for each female of each species. The ratio of survival (or growth) was calculated
 268 between two randomly chosen females (one per species). This procedure was repeated 4000 times in

269 order to obtain distributions of ratios of survival (and growth) for each combination of temperature and
270 food condition. We then calculated the average of four ratios, comparing the performance of four
271 hatches of *H. sanguineus* and four of *C. maenas*; this calculation takes into account that larvae obtained
272 from four different females were used for each study. In addition, this procedure to calculate the average
273 maintained the correlation between survival and growth. Using the simulated ratios of survival and
274 growth, we calculated the centroid and the 90% confidence ellipses which were plotted for each
275 combination of temperature and food (see Fig. 5).

276

277 **RESULTS**

278 We first describe the survival, development and growth rates for *Hemigrapsus sanguineus* and we
279 compare them to the data from *Carcinus maenas* already published (Torres and Giménez 2020,
280 PANGAEA: <https://doi.org/10.1594/PANGAEA.918056>). The graphs showing the results
281 corresponding to already published data of *C. maenas*, are highlighted with a grey background in figures
282 1, 2 and 3.

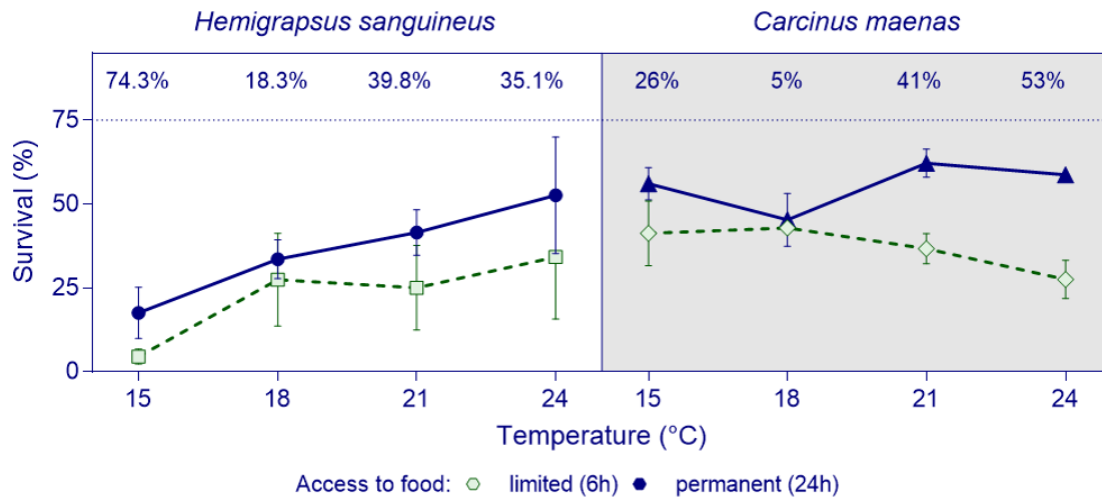
283 **Survival, development, and growth**

284 Survival to megalopa in *H. sanguineus* decreased towards lower temperatures and under limited
285 access to food (Fig. 1, left panel); while at 15 °C, food limitation reduced survival by 74 %, at 24 °C
286 the survival reduction was 35 %. Best models retained food availability and temperature operating in
287 additive or interactive ways depending on the stage (Table S3), but with variations among larvae from
288 different females (Table S4). Survival to megalopa under limited access to food ranged from 0 - 74 %
289 depending on temperature and female of origin, with 0 % survival occurring at 15 °C and 24 °C for
290 female 3 (Fig. S5). The effects of temperature and limited access to food on survival to zoeae II to IV
291 were small, but increased for survival to ZV and especially to the megalopa (Fig. S6). The sensitivity
292 to temperature varied among larvae from different females; survival at 15 °C was consistently low (13.2
293 %) but survival at 24 °C ranged from 19 to 92 % (larvae under permanent access to food) depending on
294 the female (Fig. S5).

295 The combined effects of temperature and food limitation on the overall survival to megalopa
296 (Fig. 1, left panel) were consistent with a multiplicative model (additive model retained in the
297 logarithmic scale: Table S3). The multiplicative model can be illustrated considering the combination
298 of 24 °C and permanent access to food as the optimal condition. In this case, the observed proportion
299 of survivors under the combination of two stressors experienced simultaneously (15 °C and limited
300 access to food: 0.05) is close to that expected by the product of the proportions observed when the
301 stressors were experienced in isolation (limited access to food: 0.3; 15 °C: 0.2). On a female-by-female
302 basis, the effects ranged from synergistic with a strong effect of temperature (F1 and F2) to antagonistic
303 with a strong effect of limited access to food (F3 and F4).

304 The overall response of *H. sanguineus* was clearly different from that of *C. maenas* (Fig. 1).
305 The increased survival of *H. sanguineus* with temperature irrespective of the food condition contrasts
306 with the reduced survival of *C. maenas* under food limitation and high temperatures. The response of
307 *H. sanguineus* to food limitation and temperature, consistent with a multiplicative model, also differs
308 from the strong interactive effect shown by *C. maenas* where high temperatures exacerbated the effect
309 of food limitation on survival (Fig. 1).

310



311

312 **Figure 1.** *Hemigrapsus sanguineus* and *Carcinus maenas*. Average survival from hatching to megalopa as a response to
 313 temperature and food availability. Data shown as average values \pm SE for the four females of origin. Permanent access to food:
 314 blue symbols and continuous line; limited access to food: green symbols with dashed line. Percentages on top (only when
 315 significantly different): percent difference in survival between permanent and limited access to food treatments for each
 316 temperature. Already published data for *Carcinus maenas* is presented in the right panel, grey background (Torres and
 317 Giménez 2020).

318

319 Duration of development to megalopa in *H. sanguineus* increased with decreasing temperatures
 320 in a non-linear pattern (Fig. 2a, left panel); the best model retained the interactive effect of temperature
 321 and limited access to food (Table S5). At low temperatures, the effects of food limitation were weak,
 322 producing a delay in the metamorphosis of fewer than 3 days (representing less than a 5 % change
 323 between the two food conditions). However, at 21 and 24 °C the delay was longer than 3 days,
 324 representing 14 - 16 % change (Fig. 2a, left panel). The effect of food limitation was weak at early
 325 stages and then it became stronger in the zoea IV (Fig. S7); best models retained temperature in
 326 development to stages ZII and ZIII, temperature and food availability operating in an additive way in
 327 development to ZIV, and interactively in development to ZV and megalopa (Table S5). In larvae from
 328 all females reared at 15 °C, there was an extra zoeal stage (zoea VI) regardless of food availability.

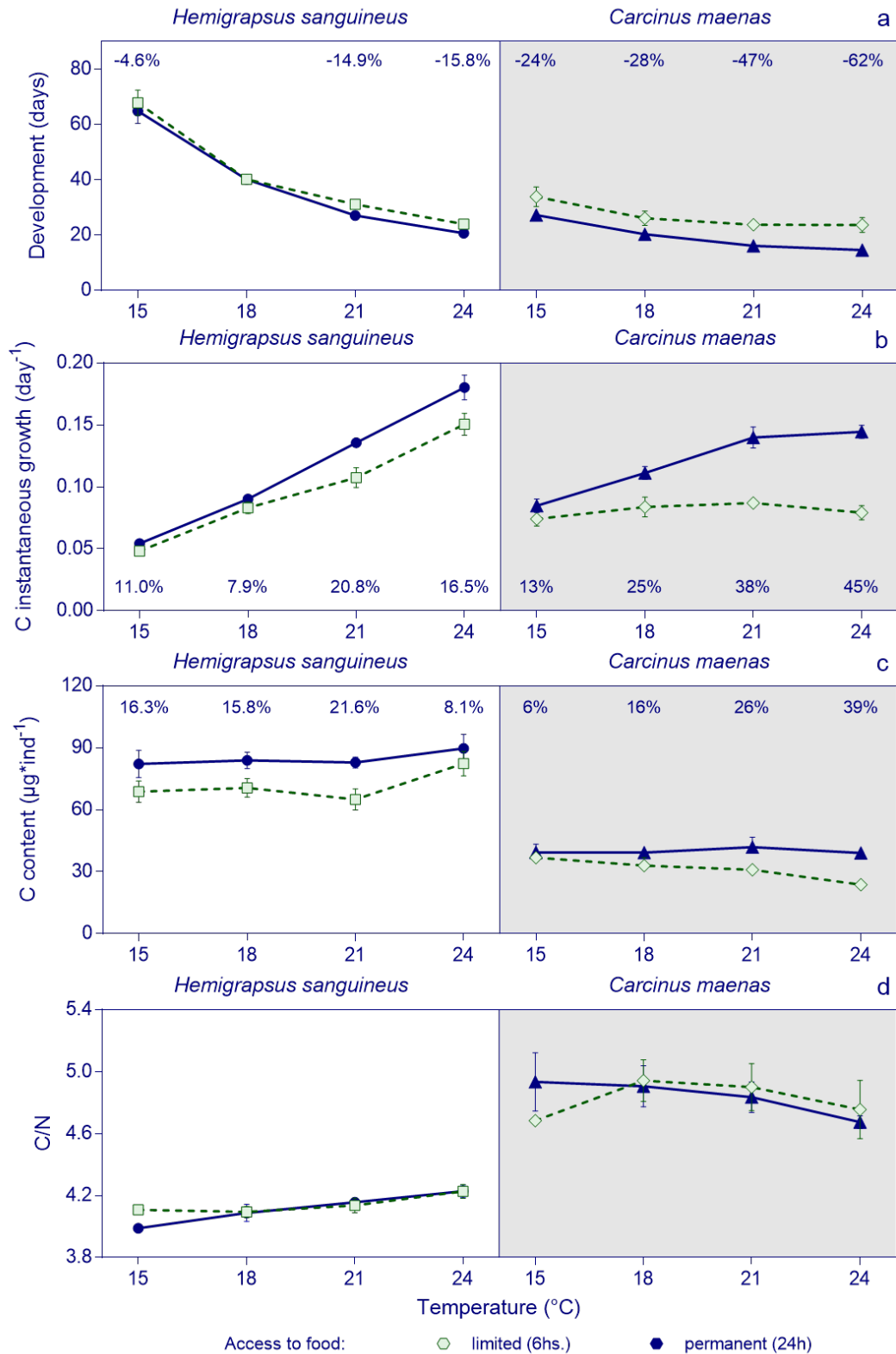
329 Carbon growth rates of *H. sanguineus* increased with increasing temperatures in both food
 330 conditions; food limitation caused a reduction in carbon growth rates, and the effect was stronger at
 331 higher temperatures (Fig. 2b, left panel); best models retained the interaction of food availability and
 332 temperature (Table S6). Similar effects were also found in terms of dry mass and nitrogen content (Fig.
 333 S8 a, c). Exposure to food limitation resulted in a reduction in body mass, carbon and nitrogen content
 334 (Figs. 2c left panel, S8 b, d), but the magnitude of the effect depended on temperature and varied among
 335 females. Best models retained food availability and temperature operating interactively (Table S6).
 336 Consistently for all females, the effect of food limitation on carbon content was strong in the range 15

337 - 21 °C as compared with 24 °C (e.g. 16 % vs 8 % decrease in carbon content). Similar effects were
338 found for dry mass and nitrogen content (Fig. S8 b, d). Food limitation reduced carbon and nitrogen
339 contents in similar proportions among temperatures resulting in comparable C/N ratios (Fig. 2d, left
340 panel); the exception was 15 °C where the reduction in nitrogen (19 %) was higher than that of carbon
341 (16 %).

342 Duration of development and growth of *H. sanguineus* larvae were more affected by
343 temperature which contrasts to those of *C. maenas*, that are more sensitive to food limitation (Fig. 2).
344 In terms of duration of development, larvae of *H. sanguineus* were more sensitive to temperature than
345 *C. maenas*, especially at low temperatures; at 15 °C *H. sanguineus* reaches the megalopa in ca 70 days
346 while *C. maenas* needs ca 30 days (Fig. 2a). By contrast, limited access to food increased developmental
347 time in a lesser extent in *H. sanguineus* (< 16 % change between the two food conditions) than in *C.*
348 *maenas* (> 20 %). While growth rates of *H. sanguineus* increased with temperature irrespective of the
349 food condition, those of *C. maenas* decreased with temperature but remained constantly low in the food
350 limited treatment (Fig. 2b). *H. sanguineus* megalopa had higher carbon content and lower C/N ratios
351 than *C. maenas* (Fig. 2d). The weakest effect of food limitation on carbon content found at the highest
352 temperature in *H. sanguineus* contrasts to the pattern found in *C. maenas*, where the effect was weakest
353 at the lowest temperature (Fig. 2c).

354

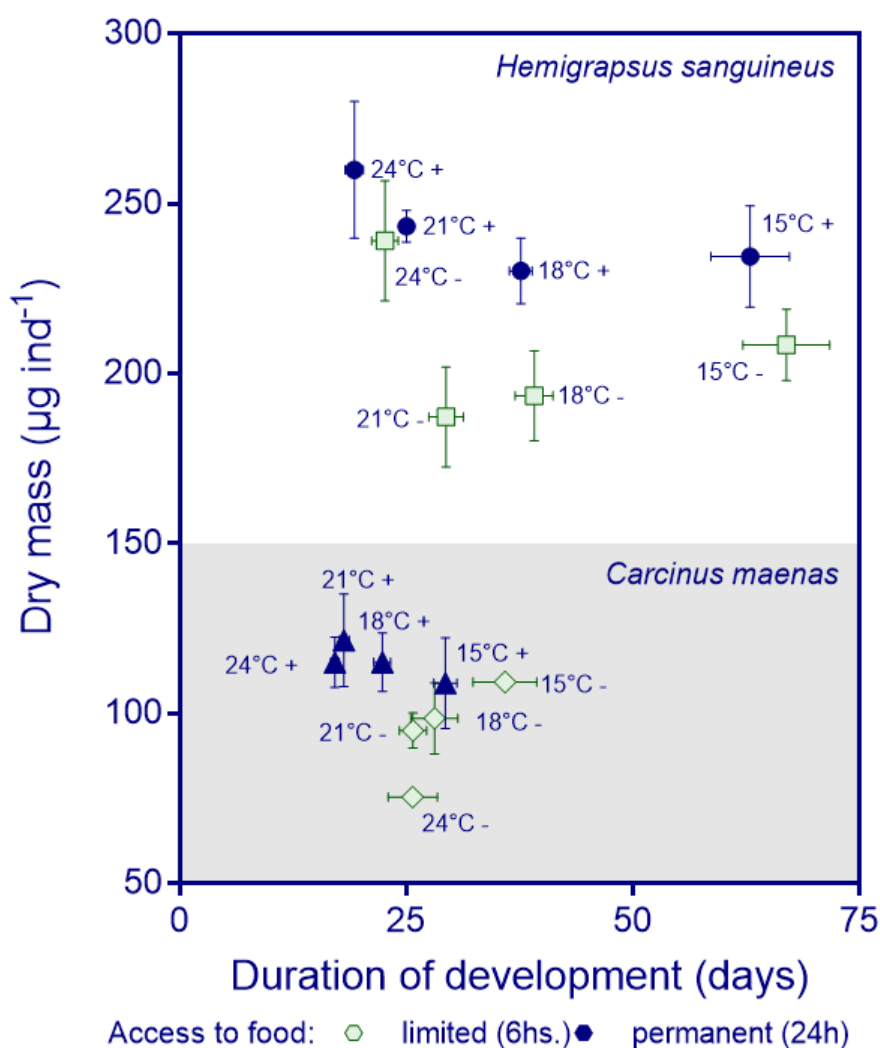
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356

357 **Figure 2.** *Hemigrapsus sanguineus* and *Carcinus maenas*. (a) Average duration of development. (b) Average growth rates. (c)
 358 Average carbon content. (d) Average C/N ratio. Data corresponds to the responses, from hatching to megalopa, to temperature
 359 and access to food. Data shown as average values \pm SE. Symbols as in Figure 1. Percentages on top or below (only when
 360 significantly different): percent difference in development time, C growth, C content, and C/N between permanent and limited
 361 access to food treatments for each temperature.

362 Integrated growth responses to megalopa of *H. sanguineus* were characterised by a strong
 363 decrease in body mass under food limitation rather than a long delay in development. The delay in
 364 metamorphosis did not compensate the effects of food limitation on growth rates. At 24 °C and under
 365 permanent access to food larvae reached a maximum threshold of body mass (= 260 µg/ ind), and
 366 differences between food conditions were small (limited access to food = 240 µg/ ind). However, lower
 367 thresholds were reached at lower temperatures, especially under limited food availability (Fig. 3, upper
 368 side of graph). In addition, *H. sanguineus* larvae did not compensate for the effect of low temperature
 369 on body mass even under permanent access to food (Fig. 3, upper side of graph). Similar patterns
 370 characterised the integrated responses in terms of carbon and nitrogen content (Fig. S9).



371

372 **Figure 3.** *Hemigrapsus sanguineus* and *Carcinus maenas*. Integrated responses of body mass and duration of development
 373 under the different treatments of temperature and food availability. Data shown as average values \pm SE for both variables.
 374 Symbols: permanent access to food is represented with blue symbols (indicated as '+') and limited access to food with green
 375 symbols (indicated as '-'), temperature is indicated in the graph next to the symbols.

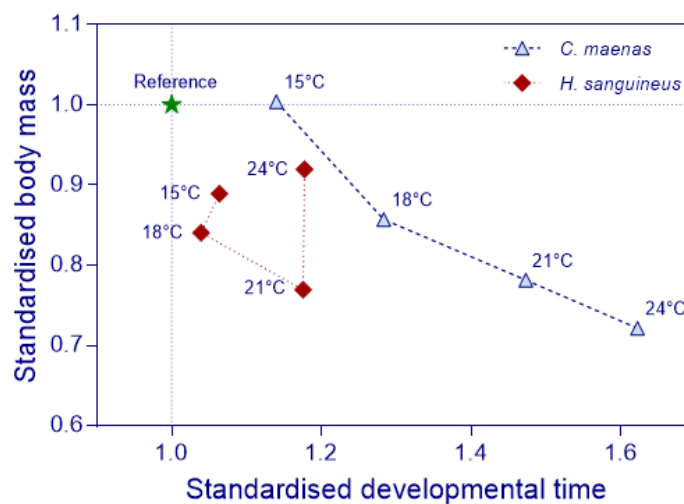
376 The range of variation in the integrative response of *H. sanguineus* is much higher than that of
 377 *C. maenas* larvae (Fig. 3). However, when the duration of development and the body mass are
 378 standardised, the opposite pattern arises. Figure 4 compares the effect of food limitation on the
 379 integrated responses of body mass and developmental time, for each temperature, of both *Carcinus*
 380 *maenas* and *H. sanguineus*. Each variable (V: body mass or duration of development) was expressed
 381 for each temperature (T) as a ratio, R:

$$382 \quad R_T = \frac{V_{T,L}}{V_{T,P}} \quad (1)$$

383 where the subindices L and P represent limited and permanent access to food, respectively. In this
 384 representation, the condition of permanent access to food is set to one irrespective of the temperature
 385 because the formula becomes:

$$386 \quad R_T = \frac{V_{T,P}}{V_{T,P}} = 1 \quad (2)$$

387 Figure 4 shows that as compared with *C. maenas*, *H. sanguineus* extended the development in a small
 388 fraction in response to limited access to food. In *C. maenas*, the proportional reduction in body mass
 389 and delay in metamorphosis became larger with temperature (Fig. 4, see also Torres and Giménez
 390 2020); by contrast in *H. sanguineus* both reduction in body mass and delays in metamorphosis do not
 391 show any consistent trend.



392

393 **Figure 4.** *Hemigrapsus sanguineus* and *Carcinus maenas*. Summary of standardised responses to the different temperatures
 394 under food limited conditions (blue triangles and red diamonds, see equation (1)). For the standardised values of body mass
 395 and developmental time, the values for permanent food conditions represent the unit (green star, see equation (2)). At the
 396 lowest temperature tested, *C. maenas* extended the developmental in response to food limitation; larvae compensated for the
 397 limited access to food, metamorphosing with high body mass. *H. sanguineus* did not compensate for the limited access to food
 398 and metamorphosed with lower body mass. Symbols: *C. maenas*: blue triangles, *H. sanguineus*: red diamonds. Data for *H.*
 399 *sanguineus* this study, data *C. maenas* from Torres and Giménez 2020.

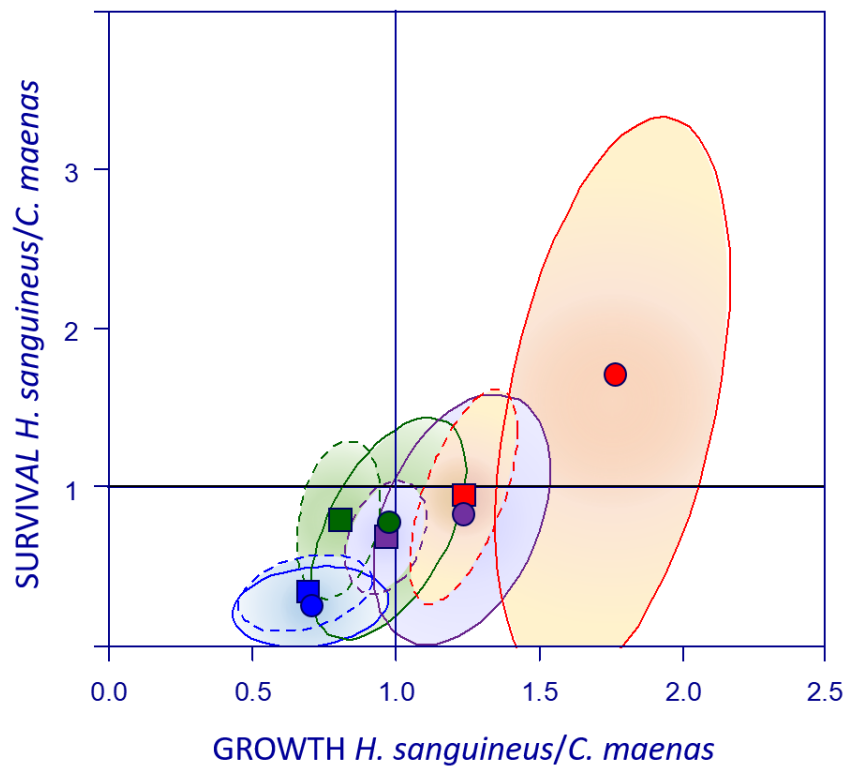


Figure 5. *H. sanguineus* and *C. maenas* simulated survival and growth ratios under different food and temperature conditions. Squares and circles denote centroids of 95% confidence ellipses in shown colour. Data for permanent access to food (circles) shown with ellipses as continuous lines and for limited access to food (squares) as dashed lines. Blue: 15 °C, green: 18 °C, purple: 21 °C and red: 24 °C.

The results of the simulated ratios of survival vs growth rates of both species showed that at most of the temperatures and food conditions (15 °C and 18 °C permanent and limited access to food and 21 °C limited access to food) larvae of *C. maenas* will be favoured; but at higher temperatures and especially under conditions of high access to food, larvae of *H. sanguineus* may be favoured (Fig. 5). The ellipses, indicating the importance of intraspecific variation in the performance of both species, show that *H. sanguineus* might perform better at higher temperatures, but it is not likely to outperform *C. maenas* at the lower temperatures that characterise the German Bight.

430 DISCUSSION

431 There were three main findings out of our experiments. First, *Hemigrapsus sanguineus* is able
432 to metamorphose to megalopa under limited daily access to food over a wide thermal range (15 - 24
433 °C). Second, within the range of temperatures experienced in the local habitat (German Bight, North
434 Sea), low temperature (15 °C) appears to be the primary limiting driver of larval performance, in terms
435 of survival and growth. Third, when the performance is quantified in relation to that of *Carcinus*
436 *maenas*, unfavourable scenarios for *H. sanguineus* are projected for temperature characterising cool
437 summers. In turn, favourable scenarios are projected for summers characterised by long heatwaves and
438 after coastal warming.

439 Temperature and food limitation

440 *H. sanguineus* larvae were able to metamorphose to megalopa under limited access to food over
441 a wide range of temperatures, with some variations among families (i.e. larvae originated from different
442 females). This is consistent with other studies on decapod crustacean larvae and strengthen the general
443 hypothesis that such larvae can reach metamorphosis as long as they access food patches for a short
444 period of time every day (Sulkin et al. 1998; Giménez and Anger 2005; González-Ortegón and Giménez
445 2014; D'Urban Jackson et al. 2014; Torres and Giménez 2020). Limited daily access to food is expected
446 in habitats characterised by food patchiness (Paffenhöfer et al. 1987; Durham and Stocker 2012; Prairie
447 et al. 2012, Robinson et al. 2021) and in scenarios where larvae perform diel or tidal migrations
448 (Forward et al. 2001; Queiroga and Blanton 2005; dos Santos et al. 2008, Thygesen and Patterson 2019).
449 In temperate latitudes such as the one of the German Bight, access to prey during night time should be
450 limited to a few hours because *H. sanguineus* larvae develop in summer when the darker period is
451 restricted to few hours. Hence, *H. sanguineus*, as other species including *C. maenas* should be able to
452 survive the larval phase as long as they access prey for a limited amount of time every day.

453 Food limitation had important consequences in terms of body mass and reserves at
454 metamorphosis to the megalopa as well as growth rates and developmental time. In species with
455 complex life cycles, traits at metamorphosis are relevant for post-metamorphic survival and are the
456 basis for trait-mediated effects of the pelagic environment on recruitment in the benthic habitat
457 (Giménez 2004; Pechenik 2006; Marshall et al. 2008; Torres et al. 2016). Larvae of *H. sanguineus*
458 under food-limited conditions metamorphosed to megalopae with reduced body mass at all
459 temperatures, suggesting that impacts of food limitation are consistent across the thermal range
460 expected for cool and warm summers. It is important to understand the mechanisms driving body size
461 at metamorphosis as they should mediate effects of warming on species responses (Lowe et al. 2021;
462 Song et al. 2021). While there is a good amount of information on mechanisms driving metamorphosis
463 in species with complex life cycles (Werner 1988; Emler 1988; Hentschel and Emler 2000), only a
464 handful of studies have investigated how such mechanisms respond to warming (D'Urban et al. 2014;

465 Torres and Giménez 2020; Griffith et al. 2021). Body mass is a plastic trait, driven by variations in
466 larval growth rates and developmental time, but body mass should vary within upper and lower
467 thresholds set by fitness costs (Werner 1988; Gotthard and Nylin 1995; Hentschel and Emler 2000;
468 Gotthard et al. 2000). Beyond the upper threshold, costs are associated to the high growth rates needed
469 to achieve a large body mass (e.g. predation). Longer developmental time contributes to larger body
470 mass but also determines the period when larvae are exposed to pelagic mortality risks (Eckert 2003)
471 and the conditions experienced after settlement (Miron et al. 1999; Jarrett 2003). Because post-
472 settlement conditions fluctuate in seasonal environments, the specific conditions experienced by
473 juveniles are indirectly determined by the duration of the larval phase. For instance, in *H. sanguineus*,
474 late settlement may result in reduced juvenile growth as individuals may miss most of the summer
475 season where growth rates are enhanced by high temperatures in the intertidal zone. During summers,
476 temperatures in the intertidal zone may boost growth because they are much higher than water
477 temperatures given the exposure to sunlight during low tides (Stephenson 1942; Lewis 1964; Somero
478 2002). In addition, late settlement may expose individuals to cannibalism (Moksnes et al. 1997;
479 Moksnes 2002; Moksnes 2004) or predation by e.g. juvenile *C. maenas*. According to life history
480 theory, costs associated to trait changes should drive the evolution of plastic responses to environmental
481 variation; because of trade-offs associated to such costs, the less responsive traits should be those with
482 higher associated costs (Gotthard and Nylin 1995; Gotthard 2000, 2004). In the case of *H. sanguineus*,
483 larvae responded to food limitation mainly through reduction of growth rates; the extension of
484 development time, found to partially compensate effects of food limitation in *C. maenas* (Torres and
485 Giménez 2020) was rather short. By contrast, *H. sanguineus* larvae showed a strong plasticity to
486 temperature by extending the development from < 20 days at 24 °C to > 60 days at 15 °C. Given that at
487 the local population, *H. sanguineus* females appear to release larvae in early summer, only after
488 temperatures reach 15 °C (Giménez et al. 2020), summers with water temperatures in the range of 15 -
489 18 °C would result in late settlement if larvae were to extend further the developmental time. Hence, in
490 the range 15 - 18 °C and under food limitation, further extension of the larval phase may have higher
491 fitness costs than metamorphosing with a smaller size (but profiting from a warm growing period).

492 **Low temperatures**

493 Our study is in line with others (Stephenson et al. 2009; Giménez et al. 2020), showing that low
494 larval survival occurs at low temperatures (range 12 - 15 °C). Reduced body mass at metamorphosis
495 found at 15 °C is expected because body mass at stage, peaks at temperatures where physiological
496 performance is optimal (Anger 1998; Anger 2001). Perhaps the body mass of megalopa of *H.*
497 *sanguineus* would decrease at temperatures higher than 24 °C, as expected from the temperature-size
498 rule (Atkinson 1994).

499 Under low temperature and food limitation, larvae hatching from all females showed a stronger
500 reduction in nitrogen as compared to carbon. This is unlike previously observed responses to stressors
501 in other species (Harms et al. 1994; Torres and Giménez 2020; Torres et al. 2021) where the main
502 characteristic is a stronger reduction in carbon than in nitrogen content. In decapod crustaceans, carbon
503 content is considered a proxy for lipid reserves (Anger and Harms 1990); hence, stress responses of that
504 type are interpreted as a reduction in the accumulation of lipid reserves, but not in the proteins needed
505 to sustain activity or the enzymatic machinery. Given that in decapods, nitrogen is a proxy for protein
506 levels, we hypothesise that the reduction observed in *H. sanguineus* reflects a thermal limitation in the
507 rate of protein synthesis. Protein synthesis accounts for a great proportion of the specific dynamic action
508 (SDA, Brody 1964) in crustaceans, i.e. the energetic costs incurred by physiological processes related
509 to feeding (including e.g. ingestion, digestion, assimilation and synthesis: Jobling 1993; Wieser 1994),
510 which increase after a meal (Houlihan et al. 1990, Robertson et al. 2001). For example, in *Carcinus*
511 *maenas*, protein synthesis accounts for 20-37 % of the post-meal oxygen rise (Houlihan et al. 1991).
512 Temperature affects rates of protein synthesis in fasted and inactive crustaceans (McMillan and
513 Houlihan 1988; Whiteley et al. 1996; El Haj and Whiteley 1997). Hence, one would expect that rates
514 of protein synthesis are compromised at lower than optimal temperatures, due to limitations in covering
515 the associated costs (Whiteley et al. 1997, 2001). In this study, the lowest temperatures tested are
516 suboptimal for *H. sanguineus*, but not for species that are native to North European Seas (e.g. D'Urban
517 et al. 2014; Torres and Giménez 2020). Hence, the differences between *H. sanguineus* and e.g. *C.*
518 *maenas*, in how C:N ratios respond to limited access to prey may reflect interspecific differences in the
519 thermal tolerance range.

520

521 **Performance of *H. sanguineus* relative to *C. maenas***

522 We found that larval performance (e.g. survival) of *H. sanguineus* at low temperatures is low
523 in relation to that of *C. maenas* also at low temperatures (Fig. 1). This is relevant to understand the
524 outcome of the balance between larval settlement, and competition between *C. maenas* and *H.*
525 *sanguineus* in the benthos. *C. maenas* larvae can complete larval development at temperatures as low
526 as 12 °C (Dawirs 1985; Nagaraj 1993). The larval season of both species partially overlap, *C. maenas*
527 larvae are released from May onwards and megalopae settle in the intertidal zone from the end of June
528 until the end of August (Giménez and Dirk, 2007). Interspecific competition occurs because juveniles
529 of both species develop in the intertidal zone during summer (Geburzi et al. 2018) The balance of larval
530 supply and competition of *C. maenas* and *H. sanguineus* must be evaluated considering both spring and
531 summer conditions. For example, a large number of *C. maenas* juveniles would survive competition in
532 years with strong settlement of *C. maenas* followed by weak settlement of *H. sanguineus* due to cool
533 summers (temperature ~ 15 °C).

534 We hypothesise that summers characterised by long heatwaves and high temperatures due to
535 warming would increase performance of *H. sanguineus* larvae and exacerbate current competitive
536 advantages already exhibited by the juvenile and adult stages in the benthos (Lohrer and Whitlatch 2002;
537 Jensen et al. 2002; Geburzi et al. 2018). Marine heatwaves, i.e. periods of time (> 3 - 5 days) when
538 temperatures are above a predefined threshold (Meehl and Tebaldi 2004; Hobday et al. 2016), can have
539 drastic consequences in structure and functioning of marine ecosystems (Garrabou et al. 2009; Marbà
540 and Duarte 2010; Wernberg et al. 2013; Mills et al. 2013) and those in the German Bight have become
541 more frequent since the 1990's (Giménez et al. 2022). During summer heatwaves such as those
542 occurring in 2018, water temperatures around the local population reached values above 18 °C
543 (Giménez et al. 2020); temperatures in waters of the coastal Wadden Sea (where *C. maenas* and *H.*
544 *sanguineus* co-occur) were much higher (BSH, 2019).

545 Another important aspect considered in our analysis concerns the role of intraspecific variation
546 in performance (IVP). IVP is common in invertebrate larvae (e.g. Appelbaum et al. 2014; Spitzner et
547 al. 2019; Torres et al. 2020), and expected from genetic variation (Marshall et al. 2008; Durrant et al.
548 2013) or parental effects (Pond et al. 1996; Shama et al. 2014). Important IVP was found in this study
549 as variation in survival, developmental time, and growth among larvae from different females (e.g. Fig.
550 S5), which is also expressed in Figure 5 as wide ellipses. The ellipses are spread across regions where
551 the relative performance switches from being stronger in *H. sanguineus* to become stronger in *C.*
552 *maenas*. The ellipses surround the area where 90 % of the 1000 simulated events for each factor
553 combination are located. Hence, Figure 5 depicts the importance of the intraspecific variation in the
554 performance of the species, as opposed to what means show; the ellipses show that, for example, there
555 is a possibility for *H. sanguineus* larvae to be able to perform (i.e. survive and grow) better than *C.*
556 *maenas* at low temperatures. Likewise for *Carcinus maenas*, there is room for their larvae to perform
557 better than those of *H. sanguineus* at high temperatures under food limitation. Species coexistence is
558 one of the important ecological consequences of intraspecific variation (Bolnick et al. 2011; Appelbaum
559 et al. 2014).

560 In synthesis, larvae from the invasive crab *H. sanguineus* are able to complete their
561 development under a wide range of temperatures and under daily limited access to food (for a period as
562 short as 6 h), with variations among families; hence, *H. sanguineus* larvae should be able to survive
563 starvation periods as long as they can access food patches for a brief time every day. *H. sanguineus*
564 responds to food limitation through lengthening the development to a much smaller degree as compared
565 to *C. maenas*; this occurs at expenses of metamorphosing to megalopa with lower body mass. It remains
566 to be seen how such responses affect the balance of costs and benefits of metamorphosing with lower
567 body mass but still profiting from the warm season increasing juvenile growth rates. *H. sanguineus*
568 shows different responses to high temperatures and food limitation from the native *C. maenas* (Torres
569 and Giménez 2020). Under cool summer conditions, we would expect limited survival of *H. sanguineus*

570 larvae which could favour *C. maenas*. By contrast, under a warming scenario, *H. sanguineus* should
571 benefit through high larval growth and survival rates. Under this scenario, increased rates of survival
572 and growth in the pelagic habitat should enhance propagule pressure of *H. sanguineus*, magnifying the
573 effect produced by being the dominant competitor in the intertidal zone. Overall, our study emphasises
574 the importance of integrative studies comparing the performance among native and invasive species
575 across their life cycles, and extending our study towards the juvenile - adult phase. Such approach will
576 help us to understand and predict effects of warming on species replacement.

577

578

579

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588 **Compliance with Ethical Standards:** The research presented in this paper complies with
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590 European parliament and of the Council of 22nd September 2010) on the protection of animals
591 used for scientific purposes.

592 **Data accessibility:** All data for this paper will be available from PANGAEA ®Data Publisher
593 <https://www.pangaea.de>

594

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