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Optics Letters

Published: 01/01/2025

Peer reviewed version

Cyswllt i'r cyhoeddiad / Link to publication

Dyfyniad o'r fersiwn a gyhoeddwyd / Citation for published version (APA): Ye, R., Zhang, X., Zhi, L., Cao, Y., Melinte, S., Wang, Z., & Yang, S. (2025). Microsphere-assisted super-resolution polarized light microscopy for polarization-sensitive materials. *Optics* Letters, 50(3), 908-911.

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Microsphere-assisted super-resolution polarized light microscopy for polarization-sensitive materials

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Polarization sensitivity is a fundamental phenomenon observed in nature, and its application is vital for advancing scientific discoveries. Here, we present a microsphere-assisted polarized light microscopy method that enables high-quality, label-free super-resolution imaging of polarization-sensitive materials. We investigated the imaging performance of this method using various samples, including colloidal crystals and grating structures. Both simulations and experiments were performed to explore the underlying physical mechanisms driving the enhancement in imaging quality. The results of this work have potential values for the fields of polarized light microscopy and super-resolution imaging.

The diffraction limit represents a fundamental barrier in conventional optical microscopy, restricting lateral resolution to half the wavelength of the illumination. Several methods have been proposed to overcome this barrier, including as structured illumination microscopy [1], superlenses [2,3], and computational microscopic imaging [4,5]. It has been demonstrated that lateral resolution beyond the diffraction limit can be achieved by depositing microspheres on the sample surface [6]. The application of microsphere-assisted super-resolution imaging for confocal imaging [7], biological imaging [8,9], and spectral imaging [10,11] demonstrates its advantages such as low cost, ease of use, and eliminating the need for fluorescent dyes. Imaging mechanisms for microspheres have been linked to the near-field evanescent wave [12], photonic hooks [13,14], photonic nanojets (PNJs) [15,16], and other mechanisms [17].

A PNJ is a focused spot formed on the shadow side of the microsphere by illuminating light projected onto the microsphere. They have waists smaller than the diffraction limit and propagate over several optical wavelengths without significant diffraction [18]. PNJs generated by microspheres have been applied on nanopatterning [19], optical inspection [20], and

spectrum measurement [21]. Another important characteristic of PNJs is polarization invariance [22]. It is reported that the PNJ generated by microspheres is basically in the same direction of polarization as that of the incident light, making it possible to perform polarized light illumination in a microsphere-assisted imaging system.

Birefringence is an inherent property of many natural and artificial materials [23]. Polarized light microscopy provides unique insights for analyzing the structures with polarization sensitivity. By measuring changes in polarization state, we can investigate the birefringent properties of anisotropic samples at the molecular level. Polarized fluorescence microscopy has been extensively used for single-molecule observations of both material and biological structures [24]. However, label-free super-resolution imaging of these structures presents challenges. In this Letter, we present a microsphere-assisted super-resolution polarized light microscopy (MS-PM) technique, which achieves high-quality super-resolution imaging for polarization-sensitive materials with the help of polarized PNJs. We applied this technique to various samples, demonstrating its superior performance over conventional microsphere-assisted unpolarized microscopy (MS-uPM). The polarization of PNJs was also theoretically and experimentally studied in this work.

As illustrated in Fig. 1(a), the MS-PM system consists of a microsphere lens integrated with a polarized light microscopic imaging system. To demonstrate the compatibility of the proposed technique with conventional optical microscopes, we employed a commercially available light microscope (Axio Imager.A2m, Carl Zeiss) in this study. The microscope is equipped with a fixed polarizer (453264, Carl Zeiss) and a rotatable analyzer (428103, Carl Zeiss). A halogen lamp (HAL 100, Carl Zeiss) was installed in the setup to provide white-light illumination with a central wavelength of 550 nm. For the microsphere lenses, we chose spherical barium titanate glass (BTG) particles due to their wide application in super-resolution microscopy. The BTG particles were purchased from Microspheres-Nanospheres (USA) and have a refractive index of ~1.90 and a diameter of around 20 um. In the experiments, the microsphere lenses were placed

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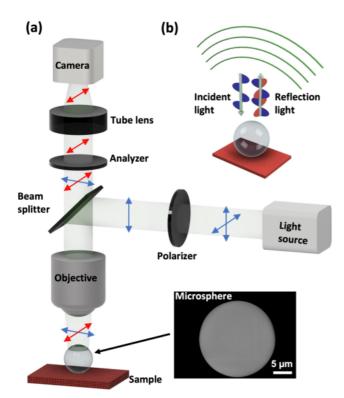


Fig. 1. (a) Diagram of the microsphere-integrated polarized light microscope (MS-PM), the inset on the right is the scanning electron microscopic (SEM) image of a barium titanate glass (BTG) microsphere. (b) Schematic drawing to show the polarization of the incident and reflection light.

in contact with the observation samples, forming a magnified real image above the sample surface. A $20\times$ objective (0.40 NA, EC Epiplan, Zeiss) was used to capture these images. The resolution of the objective is ~688 nm, as calculated with Abbe's equation: $R = \lambda/2NA$, where λ is the wavelength of illumination and NA is the numerical aperture of the objective. Finally, optical microscopic images were recorded using a high-speed scientific complementary metal—oxide—semiconductor (CMOS) camera (DFC295, Leica) for further analysis.

The principle of the MS-PM method combines the superresolution capabilities of microspheres with the contrastenhancing capabilities of polarized light microscopy. As shown in Fig. 1(b), in a polarized light imaging system, light interacts with the sample and undergoes polarization changes based on the material properties. The analyzer, positioned before the camera, blocks unaltered light and transmits only the light that has undergone polarization changes caused by the sample. By setting the polarizer and analyzer orthogonally, the system isolates the light that has experienced polarization changes, highlighting subtle differences in the sample's interaction with light. This maximizes contrast in the image by selectively allowing light with the desired polarization state to pass through. The role of the microsphere lens is to collect the near-field high-frequency signals and transfer them to the objective, so the resolution of the optical microscope can be enhanced [17].

To demonstrate the advantages of the MS-PM method over MS-uPM, the imaging performance of BTG microspheres was

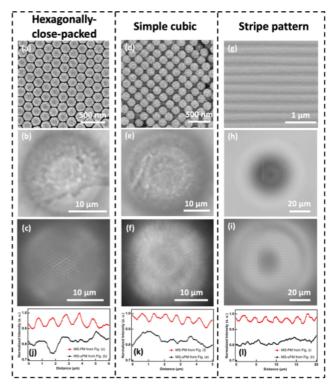


Fig. 2. (a) and (d) SEM images of silica particle arrays with (a) hexagonally close-packed (hcp) structures and (d) simple cubic structures; (g) SEM image of a BD disk; (b), (e), and (h) optical microscopic images of the (b) hcp structures, (e) simple cubic structures and (h) stripe patterns obtained with MS-uPM; (c), (f), and (i) microscopic images of the same structures obtained with MS-PM. (j)–(l) Intensity profiles across the dash lines in the corresponding images.

compared under polarized and unpolarized microscopy conditions, with the polarizer and analyzer aligned orthogonally for polarized microscopy. The first sample we used is 200-nmdiameter silica particle arrays with hexagonally close-packed (hcp) structures [Fig. 2(a)]. This sample was prepared from colloidal suspensions (Nanorainbow, China) using a gravityassisted self-assembly method [25]. Before observation, we coated the sample with 10-nm-thick Ag films via physical vapor deposition to enhance its optical reflectance. As shown in Fig. 2(b), we cannot see hcp structures through the BTG microsphere with MS-uPM. However, when we switched to MS-PM and observed the same region, the silica particles can be clearly resolved, and the structure of their arrangement became visible [Fig. 2(c)]. As shown in Figs. 2(d)-2(f), for samples with simple cubic structures, the MS-PM technique again shows a better imaging performance than the MS-uPM. The corresponding intensity profiles [Figs. 2(j) and 2(k)] further validate the improved image contrast. In addition, the MS-PM technique also demonstrates an improved performance for naturally birefringent calcite particles (Fig. S1 in Supplement 1).

Blu-ray disks (BDs) are a common popular observation sample in super-resolution imaging due to their commercial availability and sub-wavelength microstructures. As shown in Fig. 2(g), the BD has a stripe pattern with 200-nm-wide tracks and 100 nm gap between two adjacent tracks. These dimensions exceed the resolution limit of our imaging system (0.40 NA objective). Figures 2(h) and 2(i) show microscopic images of

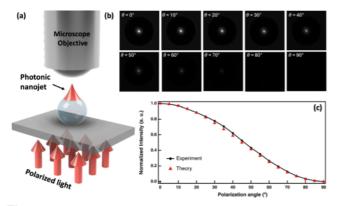


Fig. 5. (a) Schematic drawing of the experimental setup to demonstrate the polarization of PNJs. (b) Pictures of the polarized PNJs obtained when the analyzer was rotated gradually from 0° to 90°. (c) Maximum intensity of the polarized PNJ as a function of the rotation angle of the analyzer.

then propagated through a 20-µm-diameter BTG microsphere from the bottom. The microsphere was on a commercial sodalime glass microscope slide (No. 7101, Sail Brand, China). On the other side of the microsphere, we used a $50\times, 0.75$ NA microscope objective to capture the PNJ formed by the microsphere. A rotatable analyzer was positioned after the objective to determine the polarization of the PNJ. The intensity of the PNJ was measured by a CMOS camera.

As shown in Fig. 5(b), we can observe the PNJ as a bright spot in the recorded images. The intensity of the PNJ decreases as the angle between the polarizer and analyzer axes increases. The maximum intensity is achieved when the polarization axis of the analyzer is in parallel with that of the polarizer ($\theta = 0^{\circ}$). The PNJ gradually dims as the angle θ increases from 0° to 90° . Finally, the PNJ cannot be seen in the camera when the polarizer and analyzer are perpendicular to each other ($\theta = 90^{\circ}$).

Then, the intensity change of the PNJ was quantified by measuring the grayscale value of the bright spot. Figure 5(c) shows the intensity of PNJ as the function of the θ angle. The theoretical intensity in the figure is calculated using Malus's law: $I_t = I_0 \cdot cos^2 \theta$, where I_t is the intensity of the light transmitted through the analyzer, I_0 is the incident polarized light intensity, and θ is the angle between the polarizer and analyzer axes. We found that the experimentally measured data (black dots) is in good agreement with the theoretical values (red triangles), indicating that the PNJ of the microsphere is primarily linearly polarized, and the polarizing axis of the PNJ is same as that of incident light.

In conclusion, we have developed a microsphere-based superresolution polarized light microscopy method in this work, which can significantly enhance the imaging performance of conventional optical microscopes for polarization-sensitive objects by generating polarized PNJs from dielectric microspheres. The polarization of PNJs has been theoretically and experimentally confirmed. The imaging performance of the method has been demonstrated using various polarizationsensitive materials with hcp, simple cubic, and stripe structures. These findings contribute to advancements in polarized light microscopy and super-resolution imaging.

Funding. National Natural Science Foundation of China (62105156); Royal Society (IEC\R2\202178); BU-IIA Award (S46910); Belgian Fonds de la Recherche Scientifique-FNRS (T.0126.22, U.N027.18).

Disclosures. The authors declare no conflicts of interest.

Data availability. Data underlying the results presented in this Letter are not publicly available but may be obtained from the authors upon reasonable request.

Supplemental document. See Supplement 1 for supporting content.

REFERENCES

- 1. J. Qian, Y. Cao, Y. Bi, et al., eLight 3, 4 (2023).
- 2. J. B. Pendry, Phys. Rev. Lett. 85, 3966 (2000).
- N. Fang, H. Lee, C. Sun, et al., Science 308, 534 (2005).
- 4. Y. Shu, J. Sun, J. Lyu, et al., PhotoniX 3, 24 (2022).
- L. Lu, J. Li, Y. Shu, et al., Adv. Photonics 4, 056002 (2022).
- Z. Wang, W. Guo, L. L. B. Luk'yanchuk, et al., Nat. Commun. 2, 218 (2011).
- 7. Y. Yan, L. Li, C. Feng, et al., ACS Nano 8, 1809 (2014).
- 8. H. Yang, N. Moullan, J. Auwerx, et al., Small 10, 1712 (2013).
- A. Trukhova, M. Pavlova, O. Sinitsyna, et al., J. Biophotonics 15, e202200078 (2022).
- Y. Li, C. Qiu, H. Ji, et al., Adv. Opt. Mater. 11, 2300172 (2023).
- 11. J. Park, Y. Choi, S. Kwon, et al., Light: Sci. Appl. 13, 122 (2024).
- 12. Y. Ben-Aryeh, J. Opt. Soc. Am. A 33, 2284 (2016).
- 13. Q. Shang, F. Tang, L. Yu, et al., Photonics 8, 513 (2021).
- P. Zou, C. Xu, L. Zhi, et al., IEEE Photonics Technol. Lett. 36, 353 (2024).
- 15. H. Yang, R. Trouillon, G. Huszka, et al., Nano Lett. 16, 4862 (2016).
- 16. S. Yang, F. Wang, Y.-H. Ye, et al., Opt. Express 25, 27551 (2017).
- 17. L. Chen, Y. Zhou, Y. Li, et al., Appl. Phys. Rev. 6, 021304 (2019).
- 18. Z. Chen, A. Taflove, and V. Backman, Opt. Express 12, 1214 (2004).
- 19. Z. B. Wang, W. Guo, A. Pena, et al., Opt. Express 16, 19706 (2008).
- 20. J. Zhu, J. Liu, T. Xu, et al., Int. J. Extrem. Manuf. 4, 032001 (2022).
- 21. S. Kwon, J. Park, K. Kim, et al., ACS Photonics 11, 32 (2022).
- 22. P. B. Johnson, A. Karvounis, H. J. Singh, et al., Optica 8, 674 (2021).
- 23. M. Chen, F. Fan, S.-T. Xu, et al., Sci. Rep. 6, 38562 (2016).
- 24. A. M. Vrabioiu and T. J. Mitchison, Nature 443, 466 (2006).
- C. Fang, S. Yang, X. Wang, et al., Colloid Interface Sci. Commun. 37, 100286 (2020).